ISSN 0970-3209 ISSN 2231-6744 (online)

**DECEMBER 2019** | **VOL. 36** | #4

I

J

A

# INDIAN JOURNAL OF ANIMAL NUTRITION



AN OFFICIAL PUBLICATION OF
ANIMAL NUTRITION SOCIETY OF INDIA

www.indianjournals.com | www.ansi.org.in http://epubs.icar.org.in/ejournal/index.php/IJAN

#### ANIMAL NUTRITION SOCIETY OF INDIA

#### **Central Executive Committee**

President
A.K. Tyagi
Karnal, Haryana

Vice-President (North Zone) Shiv Vani Katoch

Palampur, Himachal Pradesh

Vice-President (East Zone) Pankaj Kumar Singh

Patna, Bihar

Vice-President (South Zone)
MVAN Suryanarayana
Tirupati Andhra Pradesh

Vice-President (Central Zone) Vice-President (West Zone) S.K. Verma Harish Savsani

Meerut, Uttar Pradesh

Junagarh, Gujarat

Secretary
Udeybir Singh Chahal
Ludhiana, Punjab

Chief Editor
Asit Das
Karnal, Haryana

Joint Secretary
Parminder Singh
Ludhiana Punjah

Ludhiana, Punjab

**P. Vasan** Namakkal, Tamil Nadu Chief Editor Treasurer
Asit Das Amrit Pal Singh Sethi
Karnal, Haryana Ludhiana, Punjab

Joint Treasurer Jaspal Singh Hundal

Ludhiana, Punjab

Editors
Sachin Kumar
Karnal, Haryana

Muneendra Kumar

Mathura, Uttar Pradesh

#### **CEC Members**

Ahmed Haider Ali, Srinagar, Jammu & Kashmir Amlan Kumar Patra, Kolkata, West Bengal

Jaspal Singh Lamba, Ludhiana, Punjab

Jaswinder Singh, Ludhiana, Punjab Kiran Kumari Mishra, Durg, Chhattisgarh K.S.N. Prasad, Bengaluru, Karnataka Madhu Mohini, Karnal, Haryana

Nitin Tyagi, Karnal, Haryana

Pradeep Kumar Malik, Bengaluru, Karnataka

Rajesh Nehra, Bikaner, Rajasthan Sanjay Kumar, Patna, Bihar

Sanjay Kumar Attri, Pantnagar Uttrakhand Sudarshan Singh Thakur, Karnal, Haryana Vijay Kumar Sharma, Jammu, Jammu & Kashmir Vinod Kumar Paswan, Banaras, Uttar Pradesh

#### **Nominated CEC Members**

Dr. S. S. Kundu, Haryana Dr. Tapash Kumar Ghosal, West Bangal

Dr. Atul Dhok, Maharashtra Dr. Neti A Patel, Business Head, Animal Health Division.

Dr. Sunil Jadhav, IVRI, Barelly The Himalaya Drugs Company, Bangalore

#### **Indian Journal of Animal Nutrition**

Aims and Scope: The Indian Journal of Animal Nutrition (IJAN) publishes original papers on research in the fields of animal nutrition, feed technology, feed evaluation and conservation, physiological and biochemical aspects including microbiology, biotechnology, feed and fodders and other relevant areas related to livestock feeding in developing and tropical regions. Papers describing research on given areas for ruminants and non-ruminants are welcome. The Journal is a forum for presenting peer reviewed articles on basic and applied research. Submissions addressing the new frontiers of the subject including emerging areas of nutritional genomics, modelling and interface topics such as soil plant animal and ecosystem interrelationships are given priority. Reviews on specialized topics are also included. The IJAN is published quarterly since its inception in 1983 under the aegis of Animal Nutrition Society of India (ANSI) having about twelve hundred life members around the world. ANSI has instituted two awards to be given to two selected papers published in the Journal during the biennial conferences. Authors will receive a PDF file of their articles. For inquiry regarding status of article, the request may be sent to Chief Editor at editoransi@gmail.com. The articles may be sent on line using the hyperlink: http://epubs.icar.org.in/ejournal/index.php/IJAN/about/submissions#onlineSubmissions
For any other details visit sites:- www.indianjournals.com and www.ansi.org.in

# INDIAN JOURNAL OF ANIMAL NUTRITION

(A quarterly publication)

#### **CHIEF EDITOR**

Asit Das, NDRI, Karnal, Haryana

#### **EDITORS**

P. Vasan	Sachin Kumar	Muneendra Kumar
Namakkal, Tamil Nadu	Karnal, Haryana	Mathura, Uttar Pradesh

#### EDITORIAL BOARD

Ashish Tyagi, University of Louisville, Louisville, United States

Luisa Biondi, University of Catania, Catania, Italy Dragan Sefer, Belgrade University Serbia, Serbia Nurcan Cetinkaya, Turkey

Seyed Hadi Ebrahimi, Ferdowsi University of Mashhad, Iran

Vinod Kumar Sidhu, DUVAS, Mathura, Uttar Pradesh V.B. Chaturvedi, ICAR-IVRI, Izatnagar, Uttar Pradesh, India

P.K. Nayak, ICAR-CARI, Bhubaneswar, Odisha P.M. Chatterjee, West Bengal University of Animal and Fishery Sciences, West Bengal

Goutam Mondal, ICAR-NDRI, Karnal, Haryana

K.C. Das, ICAR- CIFA, Mumbai, Maharashtra

Parimal Sardar, ICAR-CIFE, Mumbai, Maharashtra

A. Sahoo, Avikanagar, Rajasthan

B.N. Saikia, Assam

R.K. Swain, Bhubaneswar, Odisha

S.K. Mahanta, IGFRI, Jhansi, Uttar Pradesh

A.S. Mishra, ICAR, New Delhi

N.S. Maan, Hisar, Haryana

# PUBLICATION MANAGEMENT COMMITTEE

#### Asit Das, Karnal, Haryana - Chairman

A.K. Tyagi, Karnal, Haryana

T.K. Walli, Karnal, Haryana

R.C. Chopra, Karnal, Haryana

Bhupinder Singh, Karnal, Haryana

Veena Mani, Karnal, Haryana

S.V. Rama Rao, Hyderabad, Telangana

S.K. Verma, Hyderabad, Telangana

Raman Malik, Karnal, Haryana

Sachin Kumar, Karnal, Haryana

P. Vashan, Namakkal, Tamil Nadu

## IJAN is supplied free of cost to members of ANSI Life Membership of Animal Nutrition Society of India

#### Membership

A person interested in the activities of the Society who pays a lumpsum fee of Rs. 4000/- could become Life Member of the Society. For students the life membership fee is Rs. 2500/- subject to production of a proof from his guide. A Life Member will have all the rights and privileges of an Ordinary Member. The Life Membership Fee of Foreign Members for SAARC countries is US \$ 100/- and NON SAARC countries is US \$ 100/-

S. No.	Category	Amount (Annual)
1	Government and Public Institutio	n Rs. 6000/-
2	Agents fees including postage (1 year)	Rs. 7500/-
3	Benefactor members (for 10 years)	Rs. 25000/-
4	Sustaining members (for 10 years)	Rs. 25000/-
5	For SAARC countries	USD 100/-
		(postage extra)
6	For other countries	USD 100/-
		(postage extra)

All application forms and other communication may be sent to the Secretary, Animal Nutrition Society of India, Department of Animal Nutrition, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana – 141004, Punjab. Bank drafts should be drawn in favour of 'Animal Nutrition Society of India' payable at Karnal. Email: secretaryansi@gmail.com

The articles should be sent to Chief Editor using on line facility (http://epubs.icar.org.in/ejournal index.php/IJAN/about/submissions#online Submissions).

Chief Editor welcomes books for reviews; news items

**Change of Address:** May be communicated to The Secretary as well as The Chief Editor.

#### **Sustaining Member**

#### SM1. Fermenta Biotech Limited

A-1501, Thane One, DIL Complex, Ghodbunder Road, Majiwada, Thane (West) 400 610, Maharashtra, India Mobile: +91 98 92659914 | Email: keya.kulkarni@fermentabiotech.com | Web: www.fermentabiotech.com

## New Life Members of ANSI

530. Jaswandep Singh

Kalanaur, Gurdaspur-143512, Punjab

Mob. 9646661559; Email: Jashanrandhawa 1995@gmail.com

531. Vikas Phuliya

5203 A, Sector 38 West Chandigarh

Mob. 9876419632; Email:vikas.phuliya@gmail.com

532. Sunil Dev Singh

291. Sector 33, Hissar 125001, Haryana

Mob. 9466479883; Email: sdevbagla@gmail.com

533. Suresh Shankhpal Sachin Ganesh Nagar, Telephone Colony, Malkapur-443101, Maharashtra

Mob. 9898732726; Email:drshankhpalvet@gmail.com

534. Gurpreet Singh Sachdeva

JLH-10, HATHL Complex PAU, Ludiana, Punjab Phone 1612414034; Email:Gpsachdeva2000@gmail.com

535. Anil Kumar Singh

Kareha Karchhana, Allahabad, Uttar Pradesh Mob.6306088826; Email:Anilkrsingh1981@gmail.com

536. Shubham Thakur S/o Inder Singh Thakur Koon, Kot, Mandi, HP 175003

Mob.7018122717; Email:shubhi0073@gmail.com

537. Sachin Tripura South Tripura-199155

Mob. 9074117313; Email: Sachintripura 28@gm, ail.com

538. Dharmendra Chharang

48-A, Radha Nikung-I, Iskcon Road, Mansarover

Jaipur 302 020, Rajasthan

Mob. 9757216007; Email:dchharang@gmail.com

539. Manju

Department of Animal Nurtition RAJUVAS, Bikaner 302 031

 $Mob.\,9460782465; Email: dr.manju@gmail.com$ 

540. Kavipriya Jaswal

2/331 Housing Board Colony Sawai Madhopur, Rajasthan Mob. 9928011540; Email:kavipriyajaswal@gmail.com

541. Rahul Dev Mukherjee

Golanat Near Madrasha Masjid, Burdwan, Sripally, Purba

Barddhawan, West Bengal 713 103

Mob. 7044080312; Email:vetrdm@gmail.com

542. Swati Jaswal

Q.No. 558, 560, Sector-4, Type-2A Balco Nagar, Korba Chhattisgarh-495684

Mob. 7987608990; Email:Swati08jaiswal@gmail.com

543. Sudhanya Nath

Bhubaneshwar 751003, Odisha Mob. 7978205369; 9438672482 Email:sudhanyanath@yahoo.com

544. Rishabh Chugh

586, Sector-13-17, HUDA, Panipat-132103, Haryana Mob. 8770687008; Email:Rishabhchugh20@gmail.com

545. Maram Harani

D. No. 562, Nandipalli, Vempalli, Kadapa-516329 AP Mob. 7981614152; Email: Hariniredd555@gmail.com

546. Arekatla Kavya

D. No. 2404, Ayyappanagar, Yerragondapalem-5233327

Andhra Pradesh

Mob. 9346815331; Email:arekatlakavya@gmail.com

547. Gollamari Nagi Reddy

Vippariapalli, Rampicherla, Narsrgopet, Guntur-522603

Andhra Pradesh

Mob. 6281656716; Email: Golamari 027@gmail.com

548. Atluru Venkatassiva Reddy

Akottapalli, Annalur Post, Mydukur-516172 Mob. 7386349605; Email:atlurusreddy@gmail.com

549. Mannem Prasanth Sai

D. No. 1-26, Pudipatla, Tirupati Rural, Chittoor-517505,

Andhra Pradesh Mob. 8801798389

550 Kaipa Narendra Gopal Reddy

Andra Pradesh, India Mob. 8500656126

## **ACKNOWLEDGEMENT**

Animal Nutrition Society of India puts on record its sincere thanks to Indian Council of Agricultural Research, New Delhi for grant of financial support for publication of IJAN

## INDIAN JOURNAL OF ANIMAL NUTRITION

(www.indianjournals.com; www.ansi.org.in; http://epubs.icar.org.in/ejournal/index.php/IJAN)

Dece	ber, 2019 Vol. 36	#4
	CONTENTS	
	Ruminants	
1.	Effect of Feeding Straw Based Densified Complete Feed Blocks Contain Dhanwantharam Oil Residue and Tapioca Starch Waste on Body Weight, Dry Mat Intake, Feed Conversion Efficiency and Nutrient Digestibility in Crossbred Hei Calves  H.S. Sunil Gowda, Sajith Purushothaman, K. Ally, Deepa Ananth and Shibu Sin	tter ifer 329
2.	Nutrient Intake, Digestibility, Rumen Parameters and Blood Metabolites of Kaca Goats Fed Silage of Forage Mixture Produced from Intercropping of Sorgh Differing in Planting Space with Butterfly Pea ( <i>Clitoria ternatea</i> ) E. Hartati, G.A.Y. Lestari, M.M. Kleden and I.G.N. Jelantik	O
3.	Effect of Supplementing Syzygium cumini (Jamun) Fruit Shreds to Total Mixed Rate on the Performance of Lactating Crossbred Cows  A.H. Mayuresh, M. Wadhwa, J.S. Hundal, M.P.S. Bakshi, Amit Sharr Simerjeet Kaur and B.K. Bansal	342
4.	Comparative Evaluation of Oat Hay and Silage Based Rations on Nutrient Utilizat and Methane Emissions in Murrah Buffaloes U.B. Sontakke, Sonali Prusty, S.S. Kundu and Vijay Kumar Sharma	ion 347
5.	Effect of Depotash Vinasse on Rumen Fermentation Kinetics in vitro Gaurav Pratap Singh, Vandana Kumari Leitanthem, Amit N. Sharma, Madhu Moh Naresh Arora and Goutam Mondal	353 nini,
6.	The Effects of Dietary ω-3 and ω-6 Fatty Acids on Nutrient Utilization and Grove Performance in Sahiwal Heifers  Sushil Kumar, Sajjan Sihag, Zile Singh Sihag, C.S. Patil, Surender Singh Dha and Anand Kumar Pandey	358
	Non-Ruminants	
7.	A Newly Developed Mixture of Herbal Plants and Spices Enriched with Spec Extracts and Essential Oils Enhances Feed Utilisation, Growth Performance a Lowers Harmful Caecal Bacteria in Rabbits O.A.H. El-Ghalid, A.M. Abd El-Hady, G.M. El-Ashry, A.E. Kholif, O.H Matlo O.A.Olafadehan and A.M. El-Raffa	and 365
8.	A Nursery Feed for Indian Major Carps ( <i>Labeo rohita</i> ) and its Evaluation in Farme Field  Krushna Chandra Das, Kedar Nath Mohanta, Santosh Kumar Nay Snehalata Mohanty, Prabin Kumar Sahoo and Priyabrat Swain	377

9.	Effects of Feeding Graded Levels of Distillers Dried Grains with Soluble (DDGS) With or Without Supplementation of Multi-enzymes on Blood Bio-Chemical Constituents of Indigenous Chicken	382
	Ashim Kumar Saikia, Robin Bhuyan, Bibeka Nanda Saikia, Digendra Nath Sarma, Ranajit Roychaudhury, Arundhati Bora and Joga Dev Mahanta	
10.	Effect of Inclusion of Black Pepper Powder as Natural Feed Additive on the Performance of Japanese quail V. Sri Divya, D. Srinivas Kumarand E. Raghava Rao	388
11.	Chemical Evaluation and Nutrient Digestibility of Shrimp Waste Meal in Broilers N. Mounica, J.V. Ramana, D. Srinivasa Rao, J. Suresh and P. Kavitha	393
12.	Growth Performance, Carcass Traits and Economics of Kadaknath birds Rearing under Intensive Condition in Hot and Humid Climate P.K. Jena, B. Panigrahi, N. Panda, L.M. Mohapatra, B.K. Mallik, J. Bagh, P.K. Pati and A. Baliarsingh	399
13.	Effect of Dietary Supplementation of Carrot Meal on Survival, Growth and Pigmentation of Freshwater Ornamental Fish, Koi Carp, Cyprinus Carpio (L.)  Abhinka Jain, Vaneet Inder Kaur and Shathanagouda Admane Hollyappa	405
14.	Effect of Dietary Incorporation of Short Chain and Medium Chain Fatty Acid on Feed Intake and Serum metabolites in Broiler Chickens  Banani Gantayat, S.M. Durge, S.A. Amrutkar and V.B. Dongre	414
	<b>Short Communication</b>	
15.	Successful Management of Nitrite Poisoning in Crossbred Dairy Calves  E. Niyas, Shibu Simon, Surej Joseph Banglavan, Jith John Mathew, Ani S. Das, Reni John and S. Reshma	419
16.	In Sacco Degradability Kinetics of Cumbu Napier (COBN-5) Fodder at Two Different Stages of Harvest M. Madesh, K. Raja Kishore, D. Srinivas Kumar and A. Anitha	423
	<ul> <li>Contents (January-December 2019)</li> <li>Author Index</li> <li>Subject Index</li> <li>List of referees during 2019</li> </ul>	

Indian J. Anim. Nutr. 2019. 36 (4): 329-333 doi: 10.5958/2231-6744.2019.00054.9

## Effect of Feeding Straw Based Densified Complete Feed Blocks Containing Dhanwantharam Oil Residue and Tapioca Starch Waste on Body Weight, Dry Matter Intake, Feed Conversion Efficiency and Nutrient Digestibility in Crossbred Heifer Calves

H.S. Sunil Gowda<sup>1</sup>, Sajith Purushothaman<sup>2</sup>, K. Ally<sup>3</sup>, Deepa Ananth<sup>4</sup> and Shibu Simon<sup>5</sup>
Department of Animal Nutrition, College of veterinary and Animal Sciences,
Mannuthy-680651, Thrissur, Kerala, India

#### **ABSTRACT**

The study was carried out to evaluate the effect of feeding straw based densified complete feed blocks (DCFB) containing dhanwantharam oil residue and tapioca starch waste (TSW) on body weight (BW), dry matter intake (DMI), feed conversion efficiency (FCR) and nutrient digestibility in crossbred heifer calves. Eighteen healthy crossbred calves were divided into three groups of six each as uniformly as possible with regard to age, sex and body weight and each animal was allotted randomly to one of the three dietary treatments: T<sub>1</sub> (straw based densified complete feed block (DCFB), T<sub>2</sub> (DCFB containing 20% dhanwantharam oil residue) and T<sub>3</sub> (DCFB containing 20%TSW). Feeding trial lasted for 90 days. A digestion trial of five days duration was conducted to assess nutrient digestibility. Average daily gain (ADG), DMI and FCR did not differ in crossbred calves of different groups. The apparent digestibility of nutrients also did not differ among the groups. The cost of feeding in group fed with DCFB containing dhanwanthram oil residue was significantly reduced than control group fed with conventional feed blocks. Thus, feeding of straw based DCFB containing dhanwantharam oil residue and TSW are beneficial in terms of feed cost.

**Key words:** Densified complete feed blocks, Dhanwantharam oil residue, Nutrient digestibility, Tapioca starch waste

#### INDRODUCTION

The availability of fibrous crop residue especially that of cereal crop and coarse straw increased to 44.26 million tonnes in 2011-12 (NIANP, 2013). This provides a great opportunity to the livestock farmers to use these as source of roughage to feed their livestock. The storage and transportation of crop residues are highly unprofitable because of its low bulk density which prevents the usage of these residues. Hence the straws are left in the field or burnt which may cause environmental pollution (FAO, 2012). Densification of straws into blocks with concentrates and other nutrients increased the bulk density and nutritional quality (Preston and Leng 1984). Densified complete feed block (DCFB) comprised of proper ratio of roughage and concentrates along with other supplements like vitamins and minerals and made into a block. DCFB provides an opportunity for incorporating

deficient nutrients and unconventional feed such as agro-industrial and ayurvedic pharmaceutical industry byproducts. Ayurvedic pharmaceutical industry by-product dhanwanthram oil residue and agro industrial byproduct TSW were found to be useful in improving growth in crossbred calves (Purushothaman, 2018; Seethal, 2018). The work on the incorporation of agro-industrial and ayurvedic pharmaceutical industry by-product in DCFB are scarce; hence, the present study was taken to assess the effect of feeding straw based DCFB containing dhanwantharam oil residue and tapioca starch waste on the performance of crossbred heifer calves.

#### MATERIALS AND MATHODS

Three complete rations were formulated with 13-15 % crude protein (CP) and 60 % total digestible nutrients (TDN). Straw was chopped, the concentrate mixture was prepared and 5 % corn-steep liquor

<sup>1</sup>Corresponding author Email: sunilgowda17894@gmail.com Ph. No. 9916710813; M.V.Sc scholar; <sup>2&4</sup>Assistant professor; <sup>3</sup>Professor and head; <sup>5</sup>Assistant professor, Department of ARGO

(mixture containing water soluble extracts of corn soaked in water) was added which acted as a binder for block making. Then the chaffed straw and corn-steep liquor added, concentrate mixture were mixed thoroughly using horizontal mixer for about 5-10 minutes for uniform mixing. Finally, this mixture is fed to the block making machine (Real Tech Engineers, Coimbatore) and compressed to form DCFB at a pressure of 6000 psi for 30 sec. The prepared blocks were stored and used for feeding of experimental calves.

Eighteen healthy crossbred heifer calves of eight to twelve months of age were selected from University Livestock Farm and Fodder Research Development Scheme (ULF & FRDS), Mannuthy, Kerala, India. The calves were housed individually in well ventilated, clean and dry shed with facilities for feeding and watering. The calves were allocated into three groups of six each as uniformly as possible with regard to age, sex and body weight and each animal was allotted randomly to one of the three dietary treatments. The experimental design used was randomised block design. All the calves were dewormed before the commencement of the study. All the experimental calves were fed with straw based DCFB containing 60 % TDN and 13-15 % CP. Three experimental rations were T<sub>1</sub> - straw based DCFB (as control)  $\rm T_2$  - DCFB incorporating 20 % dhanwantharam oil residue and T<sub>2</sub> - DCFB incorporating 20 % TSW. Nutrients requirement of experimental animals were met by feeding as per the ICAR feeding standards (ICAR, 2013). Ad libium clean fresh drinking water was offered to all the animals. All the animals were maintained under identical conditions of feeding and management throughout the experimental period of 90 days.

Body weight of the animals was recorded at fortnightly intervals during the trial. Weighed quantities of DCFB were offered daily to all the experimental animals in the morning at 0900 h and afternoon at 1400 h. Residue left was collected manually and weighed, twice a day for analysing the moisture content and estimating daily DMI. A digestibility trial for a period of five days was carried out at the end of feeding trial. During digestion trial quantities of daily feed offered,

residue left and feces voided were recorded. DM content of the feed offered as well as residue left was determined daily. The dung voided by each animal was collected quantitatively. The entire quantity of dung voided by each animal during the 24 h were weighed separately at 0800 h every day and representative samples (@1% of the total quantity voided) were taken after mixing thoroughly and stored in double lined polythene bags. The samples collected each day were stored in deep freezer (-20°C) for further analysis. At the end of the trial, samples of dung collected for the five consecutive days from each animal were pooled, mixed and representative samples were taken after through mixing for chemical analysis. The moisture and CP content in faecal samples was estimated. The proximate composition of feed, fodder and dung were analyzed as per the standard procedure (AOAC, 2016).

Data obtained on the various parameters during the course of the experiment were analyzed statistically as per Snedecor and Cochran (1994) by analysis of variance (ANOVA) technique, using the software of Statistical Programme for Social Sciences (SPSS) version 24.0.

#### RESULTS AND DISCUSSION

Summarized data on mean initial and final BW, total weight gain and ADG for the heifer calves maintained on three treatments are illustrated in Table 1. Statistical analysis of the data revealed that there was no significance variation (P>0.05) in the total weight gain and ADG of heifer calves fed on various treatments.

Patil *et al.* (2017) also observed no significance variation in the body weight of crossbred calves fed with complete feed block (CFB) without mahua seed cake and CFB containing 5 % mahua seed cake and the values were 177.73 and 181.32 kg, respectively. The results obtained in the present study are similar to above observations. However, Sharma *et al.* (2010) reported that total body weight gain and ADG was insignificant in the crossbred female calves fed with complete feed in mash form, conventional ration and complete feed block form. On disagreement to present results,

Table 1. Performance of heifer fed on three different complete feed blocks

Parameters		P-value		
_	T <sub>1</sub>	$T_2$	$T_3$	
<b>Growth performance</b>				
Initial BW (kg)	135.33±8.49	135.63±13.05	132.47±18.28	0.984
Final body weight (kg)	196.50±9.36	201.30±18.25	191.77±23.26	$0.932^{\mathrm{ns}}$
Total weight gain (kg)	61.17±3.14	65.67±6.78	59.30±5.25	$0.687^{\rm ns}$
ADG (kg)	$0.68 \pm 0.03$	$0.73 \pm 0.08$	$0.66 \pm 0.06$	$0.687^{\mathrm{ns}}$
Feed intake				
Total DMI (kg/animal)	425.50±23.06	436.84±44.13	419.94±54.65	$0.960^{\rm ns}$
Average daily DMI (kg/animal)	4.65±0.25	4.72±0.46	4.56±0.58	$0.971^{ns}$
Average DMI/ 100kg BW	2.94±0.004	2.94±0.007	2.95±0.021	$0.833^{\mathrm{ns}}$
FCE	7.03±0.47	6.73±0.41	6.97±0.38	$0.872^{ns}$

Mean values are based on six replicates with SE, non significant at P>0.05

Kulathunga *et al.* (2015) detected an escalation of the live weight gain in crossbred heifers fed with CFB. Similarly, Somasiri *et al.* (2010) also found that lactating dairy cattle fed with gliricidia leaf meal block showed greater live weight gain.

Statistically, there was no significant variations (P>0.05) in daily DMI and DMI per kg metabolic body size of heifer calves maintained on the three treatment groups. The data on DMI per 100 kg body weight of heifer calves revealed that during third fortnight  $T_1$  and  $T_2$  showed significantly higher (P<0.01) DMI percentage as compared to  $T_3$ . During fourth and sixth fortnight.  $T_1$  showed significantly higher (P<0.01) DMI percentage as compared to treatment 2 and treatment 3.

Similar to the results of present study, Purushothaman (2018) also reported that the DMI was similar in the crossbred cattle maintained on TMRs containing dhanwantharam thailam residue and rape seed cake and the values obtained were 6.17and 5.92 kg, respectively. Yadav *et al.* (1990) reported that the daily DMI percentage was highest in adult buffalo bulls fed with ground paddy straw based feed block as compared to animals fed with raw wheat straw and paddy straw based feed block. Khan *et al.* (2017) also observed that DMI percentage was significantly higher in adult male goats fed with DCFB with condensed tannins as compared to DCFB without condensed tannins. On disagreement to this, Das *et al.* (2004) and Singh *et al.* (2007) observed higher level of DMI in crossbred calves

Table 2. Digestibility coefficients of nutrients (%) in heifer calves fed different complete feed blocks

Parameter		Dietary treatments			
	$T_{1}$	$T_2$	T <sub>3</sub>		
Dry matter	55.39±2.39	56.28±2.71	57.93±2.12	0.758 <sup>ns</sup>	
Crude protein	58.03±1.10	58.37±2.40	57.97±2.18	0.988 <sup>ns</sup>	
Crude fibre	50.88±2.39	52.17±2.88	56.87±1.69	$0.203^{ns}$	
Ether extract	74.26±1.59	77.42±1.41	73.13±2.08	0.221 <sup>ns</sup>	
Nitrogen free extract	59.98±2.76	58.20±3.01	62.20±2.52	$0.603^{ns}$	
Neutral detergent fibre	31.88±3.42	42.25±3.46	38.44±3.71	$0.144^{ns}$	
Acid detergent fibre	34.18±2.64	37.33±2.94	35.02±3.08	$0.732^{ns}$	

Mean values are based on six replicates with SE; ns- non significant at P>0.05

fed with CFB and an increase in the DMI was also reported by Kulathunga *et al.* (2015) in crossbred cows fed with DCFB.

The digestibilities of nutrients of complete feed blocks are presented in Table 3. Statistically there was no significant variation (P>0.05) in the digestibility of nutrients in DCFB fed to heifer calves maintained on different treatments. Das *et al.* (2004) reported that there were no significant differences in the digestibility of DM, CP, CF, EE, NFE and ADF in complete feed mash and CFB fed crossbred calves. Chacko (2015) observed a similar DM, OM, CP, CF, NFE and NDF digestibility for complete feed in the lactating dairy cows.

On contrary to present results, Lailer et al. (2010) reported that DM, CF and NFE digestibility were significantly higher in wheat straw based CFB and bajra straw based CFB fed to Murrah male calves fed with conventional feed. Dwivedi and Pathak (2011) reported that digestibility of DM, OM, EE, NFE, NDF, CP and total digestible nutrient were significantly greater in block fed male buffaloes as compared to mash fed animals. The mean FCE (Table 2) of experimental heifer calves fed on three complete feed blocks T<sub>1</sub>, T<sub>2</sub> and T<sub>3</sub> were 7.03, 6.73 and 6.97, respectively. Statistically, no significant difference was observed in the cumulative FCE of heifer calves maintained on various treatments. In accordance with present results, Sharma et al. (2010) reported that there was no significant difference in the FCE in crossbred calves fed with complete feed in mash (7.47), block (7.59) or conventional feed form (7.93). Similarly, Kulathunga et al. (2015) found that there was no significant difference in the feed conversion efficiency in heifers fed with CFB prepared from different agricultural wastes such as coconut poonac, wheat flour and hybrid Napier grass. On contrary to this, Kumar et al. (2015) found that FCE increased by 16% in buffalo calves fed with fungal zoospore containing CFB. The costs of CFB per kg were ₹ 17.25, 13.89 and 16.42 for T<sub>1</sub>, T<sub>2</sub> and T<sub>3</sub> respectively. The lower cost of CFB observed in T2 might be due to inclusion of ayurvedic pharmaceutical industry by-product dhanwantharm oil residue which was obtained at free of cost. In accordance with these results, Seethal (2018) also had reported that the cost of feed will decrease with increased supplementation level of dhanvanthram oil residue in the feed of calves.

#### **CONCLUSIONS**

The cost of feeding in group fed with DCFB containing dhanwantharam oil residue was significantly reduced than control group fed with conventional feed blocks and hence can be recommended as an alternate cost-effective feed.

#### REFERENCES

- AOAC. 2016. Official Methods of Analysis. (20th Ed.). Association of Official Analytical Chemists, Gaithersburg, Meryland, USA.
- Chacko, B. 2015. Evaluation of complete feed with varying levels of neutral detergent fibre for lactating dairy cows. Ph.D thesis, Kerala Veterinary and Animal Sciences University, Pookode.
- Das, M.M., Samantha, A.K., Singh, K.K., Kundu, S.S., Sharman, S.D. and Rai, S. 2004. Effect of different forms of complete feed on nutrient utilization in crossbred calves. *Indian. J. Anim. Sci.* 74: 969-972.
- Dwivedi, P.N. and Pathak, P.K. 2012. Nutritional evaluation of leaf meal added crop residue based complete bock for growing male buffaloes. *Indian J. Anim. Sci.* 82: 397-399.
- FAO. 2012. Crop residue based densified total mixed ration

   A user-friendly approach to utilize food crop
  by-productsfor ruminant production, [by T.K. Walli,
  M.R. Garg and Harinder P.S. Makkar]. FAO Animal
  Production and Health Paper No. 172. Rome, Italy.
- ICAR. 2013. *Nutrient Requirement of Cattle and Buffalo*. Indian Council of Agriculture and Research, New Delhi, 30-34p.
- Khan, M., Pathak, A.K. and Singh, S. 2017. Formulation and preparation of densified complete feed blocks with and without condensed tannins: impact on performance of *Haemonchus contortus* infected goats. *J. Anim. Res.* 7:431-439.
- Kulanthunga, K.M.W.H., Shantha, K.Y.H.D. and Nayananjalie, W.A.D. 2015. Preparation of cattle feed blocks using agricultural wastes. *Int. J. Multidisciplinary Stud.* 2:73-80.
- Kumar, S., Sehgal, J.P., Puniya, A.K. and Kumari, R. 2015. Growth performance and fibre utilization of Murrah male buffalo calves fed wheat straw based complete feed blocks incorporated with superior anaerobic

- fungal zoospores (*Neocallimastix* sp. GR-1). *Indian. J. Anim. Sci.* 85: 275-281.
- Lailer, P.C., Dahiya, S.S., Lal, M. and Lal, D., 2010. Effect of complete feed blocks on growth performance of Murrah male calves. *Indian J. Anim. Nutr.* 27: 220-223.
- NIANP- Annual report 2013-2014. National Institute of Animal Nutrition and Physiology. Bengaluru, Karnataka. 45p.
- Patil, A.K., Chaturvedi, V.B., Singh, P., Verma, A.K. and Das, A. 2017. Effect of feeding compressed complete feed block with or without deoiled mahua seed cake on methane production and energy utilization in crossbred calves. *Indian J. Anim. Sci.* 87: 1046-1048.
- Preston, T.R. and Leng, R.A. 1984. Supplementation of diets based on the fibrous residues and by-products. straws and other fibrous byproducts as feed. pp. 373-413. (Eds) Sundstol F and Owens E. Elsevier, Amsterdam, the Netherlands.
- Purushothaman, S. 2018. Evaluation of rumen fermentation pattern and nutrient utilization in Murrah buffalo, Vechur and crossbred cattle on different feeding regimen. Ph.D thesis, Kerala Veterinary and Animal Sciences University, Pookode.

- Seethal, C.C.R. 2018. Effect of dietary incorporation of Dhanwantharam thailam residue (DTR) on growth performance in calves. M.V.Sc thesis. Kerala Veterinary and Animal Sciences University, Pookode, 52-68p.
- Sharma, D., Tiwari, D.P. and Mondal, B.C. 2010. Performance of crossbred female calves fed complete ration as mash or block vis-à-vis conventional ration. *Indian J. Anim. Sci.* 80: 556-560.
- Somasiri, S.C., Premaratne, S., Gunathilake, H.A.J., Abeysoma, Dematawewa, C.M.B. and Satsara, J.H.M.N. 2010. Effect of Gliricida leaf meal blocks on intake, live weight gain and milk yield of dairy cows. *Trop. Agric. Res.* 22: 76-83.
- Sumayya, K.K. 2018. Evaluation of complete feeds in Malabari kids. M.V.Sc thesis. Kerala Veterinary and Animal Sciences University, Pookode.
- Snedecor, G.W. and Cochran, W.G. 1994. *Statistical Methods* (8<sup>th</sup> Ed.). The Iowa State University press, Ames, IA. 314p.
- Yadav, K.K., Rathee, C.S. and Lohan, O.P. 1990. Effect of compaction of roughage based complete feed on digestibility and rumen parameters. *Indian J. Anim. Nutr.* 7: 27-30.

Received on 28-08-2019 and accepted on 10-12-2019

Indian J. Anim. Nutr. 2019. 36 (4): 334-341 doi: 10.5958/2231-6744.2019.00055.0

# Nutrient Intake, Digestibility, Rumen Parameters and Blood Metabolites of Kacang Goats Fed Silage of Forage Mixture Produced from Intercropping of Sorghum Differing in Planting Space with Butterfly Pea (Clitoria ternatea)

E. Hartati, G.A.Y. Lestari, M.M. Kleden and I.G.N. Jelantik\*

Faculty of Animal Science, The University of Nusa Cendana Jln. Adisucipto, Penfui, Kupang 25001 Indonesia

#### **ABSTRACT**

An experiment was conducted to investigate the effect of feeding silage of sorghum-butterfly pea mixture (which was made of forage harvested from sorghum sown at differing planting space and intercropped with *Clitoria ternatea*) on intake, digestibility, rumen environment and blood metabolites of Kacang goats. The experiment was designed following a replicated latin square with three treatments and three periods. In each period, two goats were randomly fed one of the three silage mixture which was made of clitoria and sorghum mixture harvested from sorghum planting space of  $20\times20~{\rm cm^2}~({\rm S}_{20})$ ,  $40\times40~{\rm cm^2}~({\rm S}_{40})$  and  $60\times60~{\rm cm^2}~({\rm S}_{60})$ . Data on feed intake, digestibility, ruminal pH, ammonia (NH $_3$ ) and volatile fatty acid (VFA) concentrations as well as blood glucose and total plasma protein were collected during the last week of each period which lasted for three weeks. Results showed that crude protein (CP) content were increased with increasing sorghum planting space as the proportion of clitoria in the forage mixture increased. Meanwhile, crude fiber (CF) were comparable among different silage. Dry matter intake (DMI) and digestibility were not different (P>0.05) in Kacang goats fed different silage. However, rumen NH $_3$  and VFA concentrations were significantly (P<0.05) lower in S $_{20}$  compared to S $_{40}$  and S $_{60}$  groups. It is concluded that in order to produce good quality silage for Kacang goats there is no benefit of increasing sorghum planting space beyond  $40\times40~{\rm cm}^2$  when intercropped with *Clitoria ternatea*.

Key words: Clitoria ternatea, Intercropping, Kacang Goats, Silage, Sorghum

#### **INTRODUCTION**

There has been a growing interest of farmer in The Province of Nusa Tenggara Timur, Indonesia to utilize Sorghum (Sorghum bicolor L) fodder as ruminant feed to cope with feed scarcity during dry season. Compared to maize, sorghum is more drought tolerant capability and fodder production is also higher due to forage addition from second and third cut (Nullik, 2009). It is also palatable for ruminants when fed as fresh, hay or silage (Gupta et al., 2000). Sorghum is commonly harvested at early generative stage and given to animals either fresh or in ensiled form. The quality of sorghum silage, however, is only sufficient for moderate level of daily gain. CP content of sorghum silage varies from 5.2% to 7.4% (Mahanta and Pachauri, 2005) with digestibility of 55-62% when harvested at milk stage (Owen, 1967). For higher level of animal production, it is therefore necessary to improve silage

CP content by mixing with high-protein forages such as a variety of forage legumes during the ensiling process. Alternatively, sorghum can be sown inter-cropped with forage legumes, hence, it will produce a certain mixture of forage with higher quality of silage. In addition, forage legumes may provide additional soil nitrogen which improves forage production (Willey, 1990) and often nutritive quality of the companion plants (Mureithi and Thorpe, 1993). One of those forage legumes which have been reported to successfully thrive and producing appreciable amount of forage in intercropping system with grass, maize and sorghum in semi-arid area is butterfly pea (Clitoria ternatea). Mureithi and Thorpe (1993) reported that C. ternatea was the better legume to Napier grass inter-croppping compared to Macroptilium atropupureus and Stylosanthes guyanensin in coastal lowland of Kenya. This legume forage is also found to be well adapted to

the local environment (Nullik, 2009; Jelantik et al., 2015).

To obtain the expected silage quality from a clitoria-sorghum inter-cropping system, it is required to know the appropriate plant density for at least one of the species in the plant mixture. Different plant density obtained by planting certain forage species at certain planting space will determine the ratio of clitoria and sorghum biomass produced as it involves competition among the system component for light as well as for water and nutrients (Ali et al., 2013). This further affects silage nutritive value which consequently influences nutrient intake, digestibility and the silage utilisation by goats. Scientific evidence on the effect of feeding silage made from forage mixture harvested from sorghum-clitoria intercropping system to meat type goats in the tropics is limited. Therefore, this study was designed to investigate the intake and nutrient utilization by Kacang goats fed silage made from sorghum clitoria inter-cropping system where sorghum was planted at different space.

#### MATERIALS AND METHODS

Eighteen 2×2 m² plots were prepared in vertisol soil during March 2018 and 1 kg cows manure compost were applied during land preparation. Clitoria seeds (*Clitoria ternatea* cv Millagra) were sown at planting space of 20×20 cm² in April 2018. Seeds were directly broadcasted and immediately watered. Two weeks after clitoria was sown and already germinating, sorghum was planted at different planting space, *i.e.* 20×20 cm², 40×40 cm² and 60×60 cm² respectively at six plots for each planting space randomly. Nitrogen fertilizer was applied at 120 g/plot or 300 kg/ha 20 days after planting. All plots were watered every 2 days and weeds were removed every two weeks.

Forages were harvested at early flowering stage *i.e.* at 60 days after planting. The harvested forage mixture was then wilted for 6 hours and thereafter chopped at 2-4 cm length before put in a plastic mini silo (50 kg each) and ensiled for four weeks. No additive was added during the ensiling process.

Six mature Kacang goats weighing  $11.54 \pm 1.15$  kg were used in this experiment following a replicated  $3\times3$  latin square design with three treatments and three

periods of 21 days each. The animals were randomly allotted to one of the three experimental feeds, *i.e.* silage of forage produced form clitoria-sorghum inter-crops with different sorghum planting space:  $20\times20$  cm ( $S_{20}$ ),  $40\times40$  cm ( $S_{40}$ ) and  $60\times60$  cm ( $S_{60}$ ). All animals were also offered concentrate mixture containing ground maize, rice bran, coconut cake, coconut oil and premix. The nutrient content of concentrate mixture was 11.19% CP, 19.95% CF, 5.4% ether extract (EE), 57.74% nitrogen free extract (NFE) and 17.58 MJ gross energy (GE)/kg DM. Experimental goats had *ad libitum* access to clean water.

As much as 50 g of silage were sampled from every silo and about 10 g was sub-sampled for DM determination. The rest were sun-dried and pooled for each silage type every period. Silage was offered *ad libitum* to the animals twice a day at 0800 and 1600 h as soon as the concentrate mixture was completely consumed. Residues were collected before morning feeding, weighed and sampled for DM determination.

A digestion trial of five-day total fecal collection was conducted for determination of apparent digestibility of DM and nutrients at the last week of each period. Feces was collected before morning feeding. The collected feces were weighed and about 10% was sampled. Each fecal sample was sub-sampled for DM determination and the rest was sun-dried. The sun-dried fecal samples were then pooled at the end of the collection period for each animal and stored till further chemical analyses. Apparent dry matter digestibility was determined by the difference between dry matter or nutrients offered and dry matter or nutrients recovered in feces. Urine was also collected at the same period. The collected urine was weighed and sample was taken and frozen thereafter.

Rumen liquid was taken 2-4 hours after morning feeding by using a stomach tube from each animals at the last day of each period. The collected rumen liquid was strained with layers of nylon cloth followed by pH determination and thereafter quickly acidified using concentrated sulfuric acid to a pH below 3.0. The samples were then stored frozen. At the end of the experiment, the frozen rumen liquids were thawed

before analyses for ammonia and VFA concentration.

Blood samples were taken from jugular vein of each animals quickly after rumen fluid collection. Blood was collected using a venojeck connected to a vacuum glass bottle containing disodium EDTA as anticoagulant. Blood samples were then centrifuged at 3000 rpm for 15 minutes. Blood plasma was taken and stored frozen until analyses for glucose and total plasma protein.

Dry matter determination was conducted by drying in the ventilated oven at 105°C until constant weight. Proximate analyses were conducted following methodology of AOAC (2005). Ash content was determined by ashing feeds and fecal matter in a furnace at 550°C. CP content was determined by using Kjeldahl method and CF was determined after acid and alkali extractions. NH<sub>3</sub>-N concentration was analyzed by micro-diffusion technique (Conway, 1957). Total VFA concentration was determined by steam distillation technique.

All data were analyzed using Proc. GLM (SPSS version 21). Significant difference between means was assigned at P<0.05.

#### RESULTS AND DISCUSSION

Biomass proportion of sorghum and clitoria as well as nutrient content of silage made from forage which was produced from a clitoria-sorghum inter-crops with different sorghum planting space is presented in Table 1. As expected, the biomass proportion of *C*.

ternatea in the total fresh forage mix increased with higher sorghum planting space while sorghum biomass was reduced. Clitoria is known as a fast growing climbing forage legume (Gomez and Kalamani, 2003) that has been intercropped with maize (Jelantik et al., 2016), grass (Mureithi and Thorpe, 1993; Ali et al., 2013) and sorghum (Ayoub, 1985) with varying success. The biomass production of clitoria declines with narrower planting space of the companion plant due to the strong negative effect of shade on plant growth (Minson and Wilson, 1994). Ali et al. (2013) found that clitoria biomass production was suppressed when C. ternatea was grown intercropped with Setaria splendida. Similarly, Ayoub (1985) reported that biomass production of clitoria was reduced by 42% when grown inter-cropped with sorghum compared to monoculture. Some results, however, indicates that C. ternatea is one of shade-tolerant forage legumes (Congdon and Addison, 2003; Nicodemo et al., 2015) with negligible effect on its biomass production when intercropped with dense planting space. Jelantik et al. (2016) reported that when harvested at 60 days the total biomass production was not reduced when intercropped with maize at different planting space, i.e  $40\times40$  cm compared to  $180\times40$  cm.

Silage CP content in the present experiment increased with increasing sorghum planting space due to increasing proportion of clitoria in the silage. Clitoria contains higher CP compared to sorghum. Ratnawati *et* 

Table 1. Forage production and nutrient content of silage used in the experiment

Item		Sorghum planting space	
•	$S_{20}$	S <sub>40</sub>	S <sub>60</sub>
Forage biomass proportion	on before ensiled, kg DM	/kg	
Sorghum	0.652	0.590	0.462
Clitoria	0.348	0.410	0.538
<b>Nutrient content (% DM</b>	basis)		
OM	80.76	80,06	79,73
CP	16.83	19,22	18,36
CF	31.51	30,54	31,21
EE	3.44	2,70	5,75
Nitrogen NFE	28.98	27,80	24,41
GE (kcal)	2.75	2,71	2,84

al. (2013) reported CP content varying 17.85-18.73% in Clitoria harvested at 120 days after planting. Hartutik et al. (2012) reported that CP content of C. ternatea grown monoculture and harvested at 90 days after planting was 18.38%. Sorghum harvested at 50 days contained 9.3% CP and declined to 5.1 % at 100 day after sowing (Saini, 2012). Ayub et al. (2004) also reported that CP content of green sorghum fodder was 9.29%. Depending on the ratio of the mixture, CP content of the mixture varied from 13.23% to 16.16 % in rice bean dominating forage mixture (Ayub et al., 2004).

Unlike CP content, the silage CF content in the present experiment was comparable among different type of silage despite higher proportion of sorghum in the forage mixture with increasing planting space. Similar result was reported by Ayub *et al.* (2004) who found that CF content of sorghum rice-bean mixture varied between 30 to 34%, even when a larger

variation in CF content was noted for rice-bean and sorghum grown alone, *i.e.* 29.5-36.5%, respectively. This was possibly due to the increasing sorghum fiber content with increasing plant density while that of clitoria remain unchanged. Aydemir and Kizilsimsek (2018) found that NDF and ADF of sorghum were increased in lower plant density but this did not occur with soybean when they were grown as intercrops.

Nutrient intake and digestibility of silage of forage mixture produced from sorghum-clitoria inter-crops with different sorghum planting space in Kacang goats is presented in Table 2. DMI varied narrowly between 35.3 to 38.2 g LW<sup>0.75</sup> day<sup>-1</sup> or about 1.9-2.0% LW of Kacang goats weighing 10.06 to 13.45 kg. This level of DMI was lower than that reported earlier in goats consuming sorghum silage. Barros *et al.* (1986) reported DMI of goats consuming sorghum and clitoria silage were 65.9 and 77.2 g/kg LW<sup>0.75</sup>, respectively. This low level of intake was unexpected

Table 2. Nutrient intake and digestibility in goats consuming sorghum-C. ternatea silage

Parameter		Treatments	SEM	P-value	
	S <sub>20</sub>	S <sub>40</sub>	S <sub>60</sub>	•	
Dry matter intake (g/day)					
DMI (% of BW)	2.068	1.915	2.018	0.112	0.634
DMI (g/kg BW <sup>0.75</sup> )	38.12	35.30	37.18	2.132	0.651
Nutrient intake (g/day)					
OM	219.77	202.99	214.85	13.541	0.680
CP	40.98	40.95	41.82	2.34	0.981
CF	47.26	44.29	45.73	3.229	0.814
EE	13.70	10.27	14.75	1.091	0.046
NFE	106.74	96.44	101.63	6.65	0.572
GE (MJ/day)	37.40	33.81	36.83	2.357	0.277
Digestibility (%)					
DM	66.06	68.90	62.98	2.528	0.307
OM	68.42	70.69	64.79	2.360	0.262
CP	71.82	76.46	72.02	1.78	0.180
CF	35.73	46.60	35.53	4.596	0.211
EE	79.03a	70.89b	69.82b	2.127	0.025
NFE	75.84	74.14	69.58	2.741	0.156
Energy	73.87	74.84	70.27	1.960	0.277

Means within the same rows followed by a different alphabet differ significantly (P < 0.05)

considering the diets offered to the animals were relatively of good quality. All silage used in the present experiment contained high crude protein, *i.e.* much higher than 6.2% at which the intake of ruminant animals started to fall (Minson, 1990). Potchoiba (1990) observed DMI between 3.6 to 3.9% BW in goats fed good quality feeds high in CP content.

There are a number of factors that may cause the low silage intake by Kacang goats in the present experiment. High fiber content in sorghum-based silage is often reported to limit intake. Ngongoni *et al.* (2008) explained the observed lower intake of sorghum based silage compared to maize-based silage as sorghum silage contained higher NDF as compared to maize silage. Nichols *et al.* (1998) found that DMI of dairy cows were reduced with increasing NDF content of sorghum silage. Cooke *et al.* (2008) also found high correlation between DMI and forage NDF content. Legumes often do not ensile well due to moisture content and high buffering capacity which can cause high effluent loss and unstable silage of high pH and such silage has low palatability (Titterton, 1997).

Nevertheless, the increasing DMI with increasing sorghum planting space observed in this experiment was expected due to the increasing proportion of clitoria biomass in the silage. As discussed before, the increase of clitoria in the silage improved protein content of the silage by 14.2%. With this increase, it would be expected that DMI is increased. Negesse *et al.* (2001) reported a 36% increase of DMI when dietary CP level was increased from 8% to 15.5%.

Similarly, Jia et al. (1995) found increasing DMI with increasing dietary CP level. In addition, C. ternatea was reported to have higher intake compared to forage sorghum at any stage of maturity (Kawas et al., 1985). The results of the present experiment, however, showed that DMI and nutrient intakes were not different (P>0.05) between goats offered different types of silage. This finding is contrary to other research finding where legumes were included during sorghum ensiling. Ngongoni et al. (2008) reported that the inclusion of 40% lablab (Lab lab purpureus) in sorghum silage increased DMI in sheep. One explanation is that dietary CP content of the control diet was optimum that provided NH, and/or amino acid or peptida for microbial population and activities, thus further increase in CP content of the diet did not result in any further improvement in intake. This phenomenon was for example reported by Negesse et al. (2001) who observed a significant increase of intake of Saanen kids when dietary CP was improved from 80 to 105 g/kg but no further increase was noted at higher dietary CP content. In our experiment, dietary CP content was above that level. Previously, Lu and Potchoiba (1990) observed a sharp 35.0% increase in DMI when dietary CP was increased from 11.2 to 12.7% compared to only 9.0% increase when dietary CP was increased from 12.7 to 15.1%.

Since the level of intake in this experiment was low in all treatments, the difference of CP content of silage used in the present experiment was expected to have more profound effect on the DM digestibility.

Table 3. Rumen and blood attributes in goats consuming Sorgum-C. ternatea silage

		_	_	_	
Parameters	Treatment			SEM	P-value
	S <sub>20</sub>	S <sub>40</sub>	S <sub>60</sub>		
Rumen parameters					
pH	6.97	6.72	7.02	0.18	0.467
VFA (mmol/l)	$104.18^{a}$	114.58 <sup>b</sup>	121.81 <sup>b</sup>	2.15	0.001
NH <sub>3</sub> (mmol/l)	10.13 <sup>a</sup>	12.21 <sup>b</sup>	$11.07^{ab}$	0.39	0.017
<b>Blood metabolites</b>					
Total plasma protein (g/dl)	6.02	6.38	6.32	0.28	0.63
Glucose (mmol/dl)	115.29	113.18	112.86	3.789	0.835

Means within the same rows followed by a different alphabet differ significantly (P<0.05)

Results showed that the digestibility of nutrients did not differ (P>0.05) among treatments. This finding appears to corroborate well with some previous reports which indicate that C. ternatea had reasonably high digestibility due to its high CP content (Nasrullah et al., 2003). Hartutik et al. (2012) reported in vitro DM digestibility (IVDMD) and in vitro organic matter digestibility (IVOMD) of 65.95 and 66.28% for C. ternatea harvested at early flowering stage. Other researchers reported that IVDMD of C. ternatea varied between 59.9% (Ratnawati et al., 2013) to 74.15% (Nasrullah et al., 2003). Those recorded level of IVDMD was higher than that of sorghum. Hence, when there was an increase of biomass proportion of clitoria in the silage with higher sorghum planting space, it was rather expected that DM digestibility would be higher in  $S_{40}$  and  $S_{60}$  compared to  $S_{20}$ . The absence of increased digestibility with increasing clitoria proportion in the silage mixture may be caused by many different factors. One of these factors is the fermentative profile of the silage. Although CP content of silage with higher clitoria proportion due to higher sorghum planting space is higher, but most of the protein is degraded with improper ensiling process which ends up with high NH<sub>2</sub>-containing silage which has lower digestibility (Nussio et al., 2002). The other factor is that the content of water-soluble carbohydrates in sorghum was higher than in clitoria.

Rumen pH in this experiment varied between 6.72 and 7.02. This level is considered as favourable pH range which provides suitable environment for optimum ruminal fibre degradation (Van Soest, 1994). Similarly, NH $_3$  concentration in this experiment were well above the threshold level of 50 to 80 mg/l for optimal microbial growth and activities as suggested by Satter and Slyter (1974). Rumen pH did not differ (P>0.05) between goats consuming different type of silage, however, rumen liquid concentration of NH $_3$  and VFA were significantly lower (P<0.05) in S $_{20}$ , *i.e.* in goats receiving silage made of forage with narrower sorghum planting space. Silage made from higher sorghum planting space (S $_{40}$ ) and (S $_{60}$ ) were comparable. Rumen pH is the balance between the acid production rate and the buffering capacity from

saliva and to a lesser extent from weak base  $\mathrm{NH_3}$  produced in the rumen. It was therefore expected that when there was an increase VFA concentration in  $\mathrm{S_{40}}$  and  $\mathrm{S_{60}}$ , rumen pH would decline. However, since at the same time  $\mathrm{NH_3}$  concentration was also increased, it neutralized the effect.

Ruminal NH<sub>3</sub> concentration depends on CP content and its rate of degradation in the rumen. In this experiment, CP content of S<sub>60</sub> was higher than S<sub>20</sub>. However, the more determinant factors for ruminal NH<sub>3</sub> concentration is the extent of protein degradation in the rumen. In poor ensiling conditions, much of the protein is degraded into peptide, amino acids, NH<sub>3</sub> and amines. When consumed by the animals, these end products of ensilage will be quickly converted to NH<sub>3</sub> in the rumen which therefore gives rise of rumen NH<sub>3</sub> concentration. This probably explains for the increase of NH<sub>3</sub> concentration in goats consuming S<sub>60</sub>.

Rumen VFA concentration was lower (P<0.05) in goats consuming silage made of a mixture of forage produced from clitoria-sorghum inter-cropping with narrow sorghum planting space compared to sparser planting space. But rumen VFA concentration was comparable between  $\rm S_{40}$  and  $\rm S_{60}$ . The increase indicated that there was a higher extent and rate of fermentation of OM in the rumen in  $\rm S_{40}$  and  $\rm S_{60}$  compared to  $\rm S_{20}$ .

Nevertheless, rumen concentrations of both NH<sub>3</sub> and VFA were within the normal range reported earlier. Mc Donald *et al.* (2010) reported that rumen VFA concentration in ruminant typically varying between 70-150 mmol/l. Suryani *et al.* (2014) reported that goats consuming various forage and concentrate had VFA concentration of 40.45 mmol/l. Similarly, Naswantara (2006) noted a level of 71.81 mmol/l in goats consuming rice straw supplemented with different protein and energy sources. This indicates that silage of sorghum-clitoria mix in this experiment has reasonably high fermentation rate.

Blood glucose and total plasma protein were not different (P>0.05) in goats consuming different type of silage. However, the level of blood glucose was under normal level which is stated to be 50-80 mg/dl (Kaneko,

1997). Manu *et al.* (2007) reported blood glucose concentration varying between 51.00 to 94.80 mg/dl for Kacang goats in Timor savanas, Indonesia. The increase of glucose concentration in blood plasma in this experiment did not occur despite the increase of VFA concentration in the rumen fluid. In ruminant, glucose absorbed from carbohydrate digestion in the small intestine is generally low, only 3-8% of glucose synthesized in sheep (Preston and Leng, 1986). Hence, ruminants rely on the gluconeogenesis mechanism from propionate and amino acids to maintain blood glucose level. Major part (50-60%) of precursor for gluconeogenesis comes from propionate (Preston and Leng, 1986).

Total plasma protein in the present experiment. varied narrowly between 6.02 to 6.38 g/dl and no treatment effect (P>0.05) was found. This level was within the normal range reported by Mitruka and Rawnsley (1981), between 4.5-7.2 g/dl. Thus, it is apparent that the animal tends to maintain plasma protein concentration in a normal homeostatic level (Swenson and Reece, 1993).

#### **CONCLUSIONS**

It is concluded that CP content of the silage increased with increasing sorghum planting space as the proportion of clitoria in the mixture was increasing. Nutrient intake and digestibility, however, are not improved with increasing planting space although rumen NH<sub>3</sub> and VFA concentration was improved. It is therefore inferred that there is no benefit of increasing sorghum planting space beyond 40×40 cm<sup>2</sup> when sown together with *C. ternatea* to produce good quality silage for Kacang goats.

#### REFERENCES

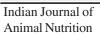
- Ali, A., Abdullah, L., Karti, P.D.M.H, Chozin, M.A. and Astuti, A.A. 2013. Production, competition indices, and nutritive value of Setaria splendida, Centrocema pubescens, and Clitoria ternatea in mixed cropping systems in peatland. *Media Peternakan* 12: 209-215.
- AOAC. (2005). Official Methods of Analysis. AOAC International. 18th ed. Airlington, USA: Association of Official Analytical Chemist.
- Ayoub, A.T. 1985. The potential contribution of some forage crops to the nitrogen budget and animal feed in the

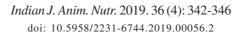
- Sudan Gezira farming system. In: I. Haque, S. Jutzi and P.J.H. Neate (Eds) *Potentials of Forage Legumes in Farming System of Sub-Sahara Africa*. Proc. Worskhop held at ILCA Addis Ababa, Ethiopia 16-19 September 1985. pp: 58-68.
- Ayub, M., Tanveer A., Nadeem, M.A., and Shah, S.M.A. 2004. Studies on the fodder yield and quality of sorghum grown alone and in mixture with ricebrean. *Pakistan J. Life Social Sci.* 2: 46-48.
- Aydemir, S.K. and Kizilsimsek, M. 2018. Assessing yield and feed quality of intercropped sorghum and soybean in different planting patters and in different ecologies. *Int. J. Environ. Sci. Technol.* 16: 5141-5146.
- Barros, N.N., Kawas, J.R., Freire, C.L, Aragjo-Filho, J.A., Shelton, J.M. and Johnson, W. L. 1986. Digestibility and intake of various native and introduced forages by goats and hair sheep in northeast Brazil. *Reuniao Techco-Cientifica Do Program De Apoio A Pesquisa Colaborativa De Pequenos Ruminantes, 1.*, 1986. Sobral. Anais. Sobral: EMBRAPA-CNPC: SR-CRSP, 1986. p. 219-233.
- Congdon, R.A. and Addison, H. 2003. Optimising nutrition for productive and sustainable farm forestry systems:

  Pasture legumes under shade. Report. Rural and Industries Research Tropical legumes in a eucalypt plantation 159.
- Cooke, K.M., Bernard, J.K. and West, J.W. 2008. Performance of dairy cows fed annual rygrass silage and corn silage with steam-flaked or ground corn. *J. Dairy Sci.* 91: 2417-2422.
- Gupta, P.C., Akbar, M.A. and Lodhi, G.P. 2000. Genetic variability in teh nutrition quality of sorghum fodder in India. *Indian J. Anim. Nutr.* 17: 70-72.
- Gomez, S.M. and Kalamani, A. 2003. Butterfly pea (*Clitoria ternatea*): a nutritive multipurpose forage legume for the tropic An overview. *Pakistan J. Nutr.* 2: 374-379.
- Hartutik, S., Fernandez, P.T. and Ratnawaty, S. 2012. Evaluation of legume herbs nutritive value as a ruminant feed and nitrogen supply on soil in West Timor, Indonesia. *Pakistan J. Agric. Res.* 25: 323-331.
- Jelantik, I.G.N., Nikolaus, T.T., Leo-Penu, C. and Jeremias, J. 2015. Herbage production and nutritive value of some forage legumes as calf supplement. Proc. The 3<sup>rd</sup> International Seminar on Animal Industry "Sustainable Animal Production for Better Human Welfare and Environment". 17-18 September 2015. Bogor, Indonesia. pp. 141-144.
- Jelantik, I.G.N., Nikolaus, T.T. and Leo-Penu, C. 2016. Forage production and nutritive value of *Clitoria ternatea* grown under different maize plant density. Proc. 3<sup>rd</sup>

- Animal Production International Seminar, 3<sup>rd</sup> ASEAN Regional Conference on Animal Production "Enhancing Synergistic Roles of Stakeholders for Development of Sustainable Livestock Production", Batu, Indonesia. pp. 374-377.
- Jia, Z.H., Sahlu, T., Fernandez, J.M., Hart, S.P. and Teh, T.H. 1995. Effects of dietary protein level on performance of Angora and cashmere-producing Spanish goats. *Small Rumin. Res.* 16: 113-119.
- Kaneko J.J. 1997. *Clinical Biochemistry of Domestic Animal*. 4<sup>th</sup> edition. Academic Press.
- Mahanta, S.K. and Pachauri, V.C. 2005. Nutritional evaluation of two promising varieties of forage sorghum in sheep fed as silage. *Asian-Australas. J. Anim. Sci.* 18: 1715-1720.
- Manu, A.E., Baliarti, E., Keman, S. and Datta, F.U. 2007. Effects of local feed supplementation on the performance of Bligon goats doe at the end of gestation reared in West Timor savannah. *Anim. Prod.* 9: 1-8.
- McDonald, P., Edwards, R.A., Greenhalgh, J.F.D., Morgan, C.A., Sinclair, L.A. and Wilkinson, R.G. 2010. *Animal Nutrition*. Seventh Addition. Prentice Hall, London.
- Mitruka, B.M. and Rawnsley, H.M. 1981. Clinical Biochemical and Hematological Reference Values in Normal Experimental Animals and Normal Humans. 2nd Ed. Year Book Medical Publisher Inc., Chicago.
- Mureithi, J.G. and Thorpe, W. 1993. The effects of herbaceous legume intercropping anf mulching on the productivity of Napier grass (Pennisetum purpureum) and total forage yield in coastal lowland Kenya. In: J. Ndikumana and P. de Leeuw (Eds). *Sustainable Feed Productionand Utilisation for Smallholder Livestock Enterprises in Sub-Saharan Africa*. Proc. 2<sup>nd</sup> African Feed Resources Network (AFRNET), Zimbabwe 6-10 December 1993. pp. 45-50.
- Nasrullah, Niimi, M., Akashi, R. and Kawamura, O. 2003. Nutritive evaluation of forage plants grown in South Sulawesi, Indonesia. *Asian-Australas. J. Anim Sci.* 16: 693-701.
- Negesse, T., Rodehutscord, M. and Pfeffer, E. 2001. The effect of dietary crude protein level on intake, growth, protein retention and utilization of growing male Saanen kids. *Small Rumin. Res.* 39: 243-251.
- Nichols, S.W, Froetschel, M.A., Amos, H.E. and Ely, L. O. 1998. Effect of fiber from tropical corn and forage sorghum silages on intake, digestion, and performance of lactating dairy cows. *J. Dairy Sci.* 81: 2383-2393.
- Nicodemo, M.L.F., de Souza, F.H. D., Pezzopane, J. R.M., Mendes, J.C.T., Junior, W. B., Tholon, P. and Santos,

- P.M. 2015. Performance of tropical legumes grown as understory of a eucalypt plantation in a seasonally dry area of Bazillian Cerrado. *Trop. Grasslands-Forrajes Tropicales*. 3: 151-160.
- Ngongoni, N.T., Mwale, M., Mapiye, C., Moyo, M.T., Hamudikuwanda, H. and Titterton, M. 2008. Research note: inclusion of lablab in maize and sorghum silages improves sheep performance. *Trop. Grasslands*. 42: 188-192.
- Nulik J. 2009. Clitoria ternatea as an alternative forage legume for cattle-maize integration system in Timor island. *Wartazoa*. 19: 43-51.
- Nussio, L.G., Paziani, S.F. and Nussio, C.M.B. 2002. Ensiling tropical grass. In: *Annual Meeting of the Brazilian Society of Animal Science*, pp. 60-83.
- Owen, F.G. 1967. Factors affecting nutritive value of corn and sorghum silage. *J. Dairy Sci.* 50: 404-416.3
- Preston, T.R. and Leng, R.A. 1987. *Matching Runinant Production System with Available Resources in The Tropic and Sub Tropic*. Penambul Book, Armidale.
- Ranawaty, S., Hartutik, Soebarinoto, and Chuzaeni, S. 2013. Production and nutritive value of shrub legumes in West Timor, East Nusa Tenggara Province, Indonesia. *J. Agric. Sci. Technol.* 3: 368-379.
- Saini, A. 2012. Forage quality of sorghum (Sorghum bicolor) as influenced by irrigation, nitrogen levels and harvesting stage. *Indian J. Sci. Res.* 3: 67-72.
- Satter, L.D. and Slyter, L.L. 1974. Effect of ammonia concentration on rumen microbial protein production *in vitro. Br. J. Nutr.* 32: 194-208.
- Suryani, N.N., Budiasa, I.K.M. and Astawa, I.P.A. 2014. Rumen fermentation and microbial protein synthesis of Ettawah Grade (PE) goat fed various composition of forage and different level of concentrate. *Majalah Ilmiah Peternakan*. 17: 54-60.
- Titterton, M. 1997. The feasibility the feasibility of using forage sorghum in drought years for silage in Zimbabwe. In: Kirk, R. (ed.) *Proceedings of the Annual General Meeting on Forage production for dairying in Zimbabwe*. Grasslands Society of Zimbabwe, August 1997.
- Van Soest, P.J. 1994. *Nutritional Ecology of the Ruminant*. 2<sup>nd</sup> ed. Cornell Univ. Press. Ithaca and London.
- Willey, R.W. 1990. Resource uses in intercropping system. *Agric. Water Manag.* 17: 215-231.
  - Received on 03-09-2019 and accepted on 25-11-2019







# Effect of Supplementing *Syzygium cumini* (Jamun) Fruit Shreds to Total Mixed Ration on the Performance of Lactating Crossbred Cows

A.H. Mayuresh, M. Wadhwa\*, J.S. Hundal, M.P.S. Bakshi, Amit Sharma, Simerjeet Kaur and B.K. Bansal

Department of Animal Nutrition, Guru Angad Dev Veterinary and Animal Science University, Ludhiana, Punjab 141 004 India

#### **ABSTRACT**

This study was taken up to assess the effect of supplementing *Syzygium cumini* (Jamun) fruit shreds on nutrient utilization, enteric methane (CH<sub>4</sub>) emission and performance of the lactating crossbred cows (CBCs). Twelve lactating CBCs were allocated into two groups (n=6) and were fed a total mixed ration (TMR) containing concentrate mixture, green Berseem fodder (*Trifolium alexandrium*) and wheat straw in 40: 42: 18 ratio on DM basis supplemented with or without *S. cumini* @ 2% for 180 days. A 14 days digestion trial was conducted by indicator method using chromic oxide green as an external marker, before the termination of feeding trial. The enteric CH<sub>4</sub> production was assessed by sulfur hexafluoride (SF<sub>6</sub>) tracer technique. *S. Cumini* fruit shreds are rich source of phenolics (21.78%), antioxidants (4.95%) and saponins (4.35%). Supplementation of diet with *S. cumini* fruit shreds did not affect intake and digestibility of nutrients. The CH<sub>4</sub> emission was mitigated (P<0.05) by 29% in CBCs fed on TMR supplemented with *S. cumini* fruit shreds. Supplementation of diet with *S. cumini* fruit shreds did not show any adverse effect on milk yield. It was concluded that CH<sub>4</sub> emission can be mitigated in lactating CBCs fed TMR supplemented with *S. cumini* fruit shreds without any adverse effect on the performance of lactating CBCs.

Keywords: Lactating crossbred cows, Methane, Milk yield, Nutrient utilization, Syzygium cumini fruit shreds

#### INTRODUCTION

India has the largest population of livestock in the world. Amongst the various livestock species, cattle population in India is 190.90 million and it contributes to around 37.28% of the total livestock population in the country (Anonymous, 2012). Being the highest milk producing nation, India contributes to 18.5% milk produced in the world (GOI, 2016). Despite the production potential of livestock, it also contributes directly or indirectly to climate change through the emissions of greenhouse gases such as carbon dioxide, CH<sub>4</sub> and nitrous oxide. Amongst various GHGs, CH<sub>4</sub> emission from enteric fermentation in ruminants contributes up to 63% (Herrero et al., 2009). The enteric CH<sub>4</sub> production can be reduced considerably by improving the efficiency of nutrient utilization and the energy lost as gaseous products of digestion can be diverted to improve the productivity of the animals.

Various strategies can be used to mitigate CH<sub>4</sub> production which includes: i) Dietary inclusion of feed

additives like organic acids, prebiotics, nitrates and sulphates, ii) Biological means by using yeast culture, bacteriocins and by enhancing acetogenesis, iii) Other strategies include animal breeding and selection and modification of rumen microbes genetically (Ingale et al., 2013). Number of studies have revealed that the in vitro CH<sub>4</sub> production can be reduced significantly by supplementing the ration with plant secondary metabolites (PSM) like essential oils (Hundal et al., 2016a), tannins (Hundal et al., 2016b) or with herbal feed additives (HFAs) containing saponins (Wadhwa et al., 2018; Singh et al., 2018a). Further, supplementing the ration with HFAs containing saponins improved performance of growing cattle (Singh et al., 2018b). Management strategies like genetic selection of efficient feed utilizers or high producing animals along with manipulation of diet (Bakshi and Wadhwa, 2009) can help in mitigating CH, emissions which would further help in increasing the efficiency in which animals use nutrients to produce milk or meat.

We hypothesized that supplementation of *S. cumini* in lactating CBCs would improve their performance by reducing enteric CH<sub>4</sub> production. Hence, this study was conducted to assess the effect of supplementing Syzygium cumini (Jamun) fruit shreds on nutrient utilization, enteric methane (CH4) emission and performance of the lactating crossbred cows.

#### MATERIALS AND METHODS

The present study was conducted in the Department of Animal Nutrition and Livestock Dairy Farm, Guru Angad Dev Veterinary and Animal Sciences University (GADVASU), Ludhiana, India. All experimental protocols were approved (GADVASU/2018/IAEC/14/09) and compliant with the guidelines established by Institutional Animal Ethics Committee constituted (IAEC) under Committee for the Purpose of Control and Supervision of Experiments on Animals (CPCSEA), New Delhi.

The S. cumini (Jamun) fruit shreds (JFS) were procured from the Unati Cooperative Marketingcum-Processing Society Ltd. Talwara, Hoshiarpur, Punjab and screened in triplicate for phenolics (Makkar et al., 1993), flavonoids (Balbaa et al., 1974) and saponins (Baccou et al., 1977). Twelve lactating CBCs were randomly divided into two groups of six animals each. In control group, cows were fed on basal TMR. In the experimental groups, the control TMR were supplemented with S. cumini fruit @ 2.0% of DM intake. Milk samples were collected at monthly interval. Impact of HFAs on the nutrient digestion were assessed by conducting a 14 days digestion trial by using chromic oxide green as an external marker @ 0.2% of daily DM intake. Faecal samples were collected from the rectum by grab method. Spot urine samples were also collected individually and was preserved in glass bottles, which were kept in refrigerator till analysed for nitrogen content (AOAC, 2007). The amount of chromic oxide (mg %) in faeces was also calculated (Hill and Anderson, 1958). The faeces, urine and orts were weighed daily at 9.00 a.m.

before feeding.

The dried and finely grounded feed, JFS, orts and faecal samples were analysed for DM, total ash, ether extract and nitrogen concentration (AOAC, 2000). Cell wall constituents were analysed by using methodology of Robertson et al. (1991). SF<sub>6</sub> trial was conducted in the 2<sup>nd</sup> and 5<sup>th</sup> month of feeding trial. CH<sub>4</sub> and SF<sub>6</sub> concentrations were determined by using gas chromatography. CH<sub>4</sub> emission rate was calculated as the product of the permeation tube emission rate and the ratio of CH<sub>4</sub> to SF<sub>6</sub> concentration in the sample. Samples were analyzed in duplicate. The gas chromatograph was fitted with a 3.3 m molecular sieve 5A column with inner diameter of 0.32 mm for SF<sub>6</sub> and a 1.5 m Porapak Q stainless steel column with inner diameter of 2 mm for CH<sub>4</sub>. The injector, detector and oven temperature was maintained at 40, 220 and 50°C for SF<sub>6</sub> and 40, 100 and 52°C for CH<sub>4</sub> estimation, respectively. Nitrogen was used as a carrier gas, with flow rate of 30 ml/min both for  $SF_6$  and  $CH_4$  estimation. Prepared standards were used to standardize the gas chromatograph for SF<sub>6</sub> (39.2 ppt and 101.7 ppt) and CH<sub>4</sub> (10.4 ppm and 101.9 ppm, Scott-Marrin Inc., Riverside, CA, USA). Background CH<sub>4</sub> [(CH<sub>4</sub>) b] was subtracted from CH<sub>4</sub> concentration in the canister [(CH<sub>4</sub>) y]. The concentration of CH<sub>4</sub> was then calculated by using the equation of Johnson et al. (1994). Enteric CH<sub>4</sub> emission rate was calculated by using following equation:

$$QCH_4 = QSF_6(CH_4)$$
 y-  $(CH_4)$  b /  $(SF_6)$ 

Where, QCH<sub>4</sub> is the CH<sub>4</sub> emission rate (g/min), QSF<sub>6</sub> is the known release rate of SF<sub>6</sub> from permeation tube (g/min), (CH<sub>4</sub>) b is the background CH<sub>4</sub>, (CH<sub>4</sub>) y is the CH<sub>4</sub> in sample in canister (g/m<sup>3</sup>) and SF<sub>6</sub> is the SF<sub>6</sub> concentration of collected sample in canister (g/m<sup>3</sup>).

Data were analyzed by using one way ANOVA and factorial design (Snedecor and Cochran 1994), by using SPSS (2009) version 16 and the significant differences in means were tested by Tukey's test.

Table 1. Chemical composition of TMR and S. cumini shreds (% DM basis)

Parameter	Ash	OM	CP	EE	NDF	ADF	Cellulose	ADL
TMR	7.60	92.40	14.82	3.10	47.20	27.00	10.85	5.30
S. cumini shreds	13.20	86.80	9.20	1.15	51.50	30.90	8.90	13.60

#### RESULTS AND DISCUSSION

The chemical composition of TMR and *S. cumini* shreds used during the experimental period is presented in Table 1. The active components present in *S. cumini* fruit shreds are presented in Table 2. Jamun seed contains various polyphenols such as ellagitannins, gallic acid, ellagic acid, etc., and phytosterol such as  $\beta$ -sitosterol. Being rich in phytochemicals, it is a potential source of phenolics, flavonids, tannins etc.

Supplementation of S. cumini shreds in the diet did not affect the daily DM intake and digestibility of nutrients (Table 3). No information is available regarding the effect of Jamun supplementation on the performance of animals. Similar to the findings of the present study, supplementation of pomegranate peel extract (PPE) upto 800 ml/cow/day to dairy cows did not exert any effect on the intake of DM and apparent nutrients digestibility (Abarghuei et al., 2013). Foiklang et al. (2016) studied the effect of grape pomace powder (GPP), mangosteen peel powder (MPP) and monensin on feed intake and nutrients digestibility in rumen fistulated steers. Results showed that GPP supplemented group had significantly higher (P<0.05) UTRS intake than those in other groups in terms of % BW and g/kg BW<sup>0.75</sup>. However, total DM intake was not significantly different among treatments (P>0.05).

Effect of supplementation of diet with *S. cumini* shreds on enteric  $CH_4$  emission was assessed by  $SF_6$  tracer technique. The results of the present study revealed that  $CH_4$  emission (g/d, g/DMI, g/kg DOMI) in *S. cumini* fruit shred supplemented group was decreased (P<0.05) by 27% (Table 4). The findings of the present study was in accordance with the results of Hundal *et al.* (2019), who observed that enteric  $CH_4$  emission (g/kg DMI) was reduced (P<0.05) by 25.3% with supplementation of diet with PPE and by 27.7%

when diet was supplemented with T. undulata extract in comparison to the animals fed control diet. CH<sub>4</sub> emission per liter of milk showed a decreasing trend but due to less number of observations, the effect was non-significant. The gross energy (GE) intake was same in animals fed control diet or diet supplemented with S. cumini. However, the loss of energy (MJ/d) from animals was less (P<0.05) by 27%, while loss of energy as percent of GE was less (P<0.05) by 28% in animals fed diet supplemented with S. cumini. CH<sub>4</sub> emission (g/ d) was observed to be higher when assessed after 5 months of feeding, however, CH<sub>4</sub> emission per liter of milk remained same during the experiment period. GE intake and loss of energy (MJ/d) was observed to be higher when estimated at the end of the experiment period (5 month), however loss of CH<sub>4</sub> % GE remained similar at 1 or 5 month of feeding trial.

Supplementation of diet with *S. cumini* shreds did not affect the milk yield of CBCs, irrespective of duration of feeding (Table 5). However, Anantasook *et al.* (2014) found that supplementation with RPM and RPO to diets of cows increased milk yield.

#### **CONCLUSIONS**

From the results it can be concluded that supplementation of *S. cumini* fruit shred reduced enteric CH<sub>4</sub> emission without impairing nutrient utilization and performance of lactating CBCs.

Table 2. Active components in S. cumini shreds

Active component	% DM basis
Total phenolics	21.78
Leucocyanidin (CT)	0.32
Saponins	4.35
Vitamin C	2.65
DPPH	2.00
Flavonoids	0.30

Table 3. Effect of S. cumini fruit shred supplementation on feed intake and nutrient digestibility

Parameters	Group		SEM	P-Value
	Control	S. cumini		
Dry matter intake, kg/d	14.83	14.88	0.113	0.134
Digestibility of nutrients, %				
Dry matter	66.08	71.66	1.514	0.660
Organic matter	70.98	71.99	1.408	0.547
Crude protein	78.98	78.76	1.065	0.601
Ether extract	73.86	72.12	1.351	0.326
Neutral detergent fibre	63.92	64.73	1.763	0.691
Acid detergent fibre	56.59	58.19	2.095	0.625
Cellulose	64.96	66.19	1.700	0.543

Table 4. Effect of supplementing S. cumini fruit shred on enteric CH<sub>4</sub> production

Parameter	Gr	oup	SEM	P-Value	Period, months		SEM	P-Value
	Control	S. cumini			1	5	-	
CH <sub>4</sub> , g/d	272.17ь	197.97a	13.44	0.008	215.29a	254.86 <sup>b</sup>	9.126	0.023
CH <sub>4</sub> , g/kg DMI	21.21 <sup>b</sup>	15.54 <sup>a</sup>	0.473	0.002	18.88	17.88	0.853	0.324
CH <sub>4</sub> ,g/kg DOMI	$32.68^{b}$	$23.68^{a}$	0.727	0.003	28.95	27.41	1.314	0.324
CH <sub>4</sub> ,g/l milk	21.79	16.41	2.735	0.214	14.79	23.41	1.485	0.100
GE intake, MJ/d	208.33	209.66	19.305	0.950	185.54a	232.45 <sup>b</sup>	6.597	0.006
Loss of energy, MJ/d	15.19 <sup>b</sup>	$11.04^{a}$	0.165	0.010	$12.02^{a}$	14.22 <sup>b</sup>	0.509	0.023
% Loss of GE	$7.33^{b}$	$5.26^{a}$	0.75	0.003	6.47	6.12	0.294	0.324

<sup>&</sup>lt;sup>a, b</sup>Mean with different superscripts in a row differ significantly (P<0.05)

Table 5. Effect of S. cumini fruit shred on milk yield

Parameters	Gr	oup	SEM Period, month			SEM			
	Control	S. cumini		1	2	3	4	5	
Milk yield, kg/d	14.04	14.09	0.18	14.16 <sup>c</sup>	12.97 <sup>b</sup>	16.22e	15.41 <sup>d</sup>	11.58a	0.28

a,b,c,dMean with different superscripts in a row differ significantly (P<0.01)

#### **ACKNOWLEDGEMENT**

Authors are highly grateful to the Indian Council of Agricultural Research (ICAR), New Delhi, India for providing the financial support to carry out the research work under the project entitled 'Estimation of Methane Emission under Different Feeding Systems and Development of Mitigation Strategies'.

#### **REFERENCES**

Abarghuei, M.J., Rouzbehan, Y., Salem, A.Z.M., and Zamiri, M.J. 2013. Nutrient digestion, ruminal fermentation and

performance of dairy cows fed pomegranate peel extract. *Livest. Sci.* 157: 452-461.

Anantasook, N., Wanapat, M., Cherdthong, A. and Gunun, P. 2014. Effect of tannins and saponins in Samanea saman on rumen environment, milk yield and milk composition in lactating dairy cows. *J. Anim. Physiol. Anim. Nutr.* 99: 335-344.

Anonymous. 2012. *All India Report*. Ministry of Agriculture. Retrieved fromhttp://dahd.nic.in/sites/default/files/Livestock 5.pdf

AOAC International, 2007. Official Methods of Analysis of AOAC International. 18th Edition. 2nd Revision.

- Gaithersburg, MD (USA).
- AOAC. 2000. Official Methods of Analysis, 17th edn., Association of Official Analytical Chemists, Washington DC: AOAC International.
- Baccou, J.C., Lambert, F. and Sanvaire, Y. 1977. Spectrophotometric method for the determination of total steroidal sapogenin. *Analyst* 102: 458-66.
- Bakshi, M.P.S. and Wadhwa, M. 2009. Dietary manipulation for mitigation of enteric methane emission. *Pakistan J. Zool.* 9: 887-893.
- Balbaa, S.I., Zaki, A.Y. and Elshamy, A.M. 1974. Total flavonoid and rutin content of the different organs of *Sophora japonica* L. *J. Assoc. Off. Anal. Chem.* 57: 752-55.
- Crompton, E.W. and Maynard, L.A. 1938. The relation of cellulose and lignin content to the nutritive value of animal feeds. *J. Nutr* 15: 987-993.
- Foiklang, S., Wanapat, M. and Norrapoke, T. 2016. Effect of grape pomace powder, mangosteen peel powder and monensin on nutrient digestibility, rumen fermentation, nitrogen balance and microbial protein synthesis in dairy steers. *Asian-Australas. J. Anim. Sci.* 29: 1416-1423.
- GOI. 2016. *Press Information Bureau*. Government of India. Ministry of Finance.
- Herrero, M., Thornton, P.K., Gerber, P. and Reid, R.S. 2009. Livestock, livelihoods and the environment: understanding the trade-offs. *Curr. Opin. Environ. Sustainability* 1: 111-120.
- Hill, F.W. and Anderson, D.L. 1958. Comparison of metabolizable energy and productive energy determinations with growing chicks. *J. Nutr.* 64: 587-603.
- Hundal, J.S., Singh, I., Wadhwa, M., Singh, C., Uppal, C. and Kaur, G 2019. Effect of Punica granatum and Tecomella undulata supplementation on nutrient utilization, enteric methane emission and growth performance of Murrah male buffaloes. *J. Anim. Feed Sci.* 28: 110-119.
- Hundal, J.S., Wadhwa, M. and Bakshi, M.P.S. 2016a. Effect of supplementing essential oils on the *in vitro* methane production and digestibility of wheat straw. *J. Anim. Nutr.* 1:14.
- Hundal, J.S., Wadhwa, M. and Bakshi, M.P.S. 2016b.
  Methane mitigation potential of tannins and their impact on digestibility of nutrients in-vitro. *Anim. Nutr. Feed Technol.* 16: 505-513.
- Ingale, S.L., Lokhande, A. and Zadbuke, S. 2013. Nutritional strategies to mitigate greenhouse gases emission from livestock agriculture: A review. *Livest Res. Int.* 1. 34-45.
- Johnson, K.A., Huyler, M., Westberg, H., Lamb, B. and

- Zimmerman, P. 1994. Measurement of methane emissions from ruminant livestock using a sulphur hexafluoride tracer technique. *Environ. Sci. Technol.* 28: 359-362.
- Makkar, H.P, Blümmel, M., Borowy, N.K. and Becker, K. 1993. Gravimetric determination of tannins and their correlations with chemical and protein precipitation methods. *J. Sci. Food Agric.* 61: 161-165.
- Robertson, J.B. and Van Soest, P.J. 1991. The detergent system of analysis and its application to human foods. In: *The Analysis of Dietary Fibre in Foods* (James W P T and Theander O). *Marcel Dekker*, *Inc. New York* Pp.123-58.
- Singh, A.S., Wadhwa, M. and Bakshi, M.P.S. 2018a. Effect of supplementing herbal feed additives containing saponins on nutrient utilization and *In vitro* methane production from total mixed ration with different roughage to concentrate ratios. In: Proc. *XVII Biennial Animal Nutrition Conference*, February 1-3, 2018, Junagadh, India, 466 pp.
- Singh, A.S., Wadhwa, M., Hundal, J.S., Uppal, C. and Bakshi, M.P.S. 2018b. Effect supplementing herbal feed additives containing saponins on the enteric methane production, nutrients utilization and performance of buffalo calves. In: Proc. XVII Biennial Animal Nutrition Conference, February 1-3, 2018, Junagadh, India.
- Snedecor, G.W. and Cochran, W.G. 1994. *Statistical Methods*.7 the edition. Oxford and IBH publications, New Delhi.
- Van Soest, P.J., Robertson, J.B. and Lewis, B.A., 1991. Methods for dietary fiber, neutral detergent fiber, and nonstarch polysaccharides in relation to animal nutrition. J. Dairy Sci. 74: 3583-3597.
- Wadhwa, M., Singh, A.S. and Bakshi, M.P.S. 2018. Effect of supplementing herbal feed additives (HFAs) containing saponins on the *in vitro* methane mitigation and fermentability of total mixed ration (TMR). In: *Proc. IX Asian Buffalo Congress* (ABS 197) p-104 held at Central Institute for Research on Buffaloes Hisar, India from 1-4 Feb, 2018.

Received on 26-07-2019 and accepted on 12-09-2019

# Comparative Evaluation of Oat Hay and Silage Based Rations on Nutrient Utilization and Methane Emissions in Murrah Buffaloes

U.B. Sontakke<sup>1</sup>, Sonali Prusty\*, S.S. Kundu<sup>2</sup> and Vijay Kumar Sharma<sup>3</sup>
Dairy Cattle Nutrition Division,
ICAR-National Dairy Research Institute, Karnal, Haryana-132001 India

#### **ABSTRACT**

In the present study oat hay and silage were compared for their methane ( $CH_4$ ) production and nutrient utilization potential in Murrah buffaloes. Sixteen dry buffaloes ( $566.5\pm12.5$  kg live weight) were selected from Livestock Research Centre of the National Dairy Research Institute, Karnal. Animals were distributed randomly into two groups ( $T_1$  and  $T_2$ ) based on body weight. Animals in group  $T_1$  and  $T_2$  were offered oat hay and oat silage, respectively along with supplementation of mineral mixture and salt. After feeding for a duration of 60 days, a digestibility trial was conducted followed by methane emission measurement by  $SF_6$  tracer technique. Average daily DM intake was similar in both the groups, whereas, fibre (NDF, ADF) intake was significantly (P<0.05) higher in hay fed group. Digestibility of nutrients (DM, OM, EE, NDF and ADF) except CP was not affected by hay or silage; CP digestibility was higher (60.59%) on silage feeding as compared to hay (58.28%) feeding. Feeding of hay resulted in more (P<0.05) enteric  $CH_4$  emissions (341.35 l/d) than silage feeding (317.86 l/d). Total  $CH_4$  production was reduced by 6.86% on oat silage than hay. Higher (P<0.05) percent of ME was lost in methane in group  $T_1$  (16.41) as compared to  $T_2$  (13.97). Thus, Oat could be stored as silage than hay for lowering methane emission from Murrah buffaloes.

Key words: Hay, Murrah buffaloes, Methane emission, Oat silage

#### **INTRODUCTION**

Scarcity of feed and fodder is the common problem faced by livestock owners of India. The problem is aggravated further by non-uniform availability of fodder throughout the year, as growing fodder availability is chiefly dependent upon the climate. There is surplus availability of green fodder during flush season, whereas, during extreme seasons majority of Indian farmers get less or even no green fodder. The excess fodder produced, stored either as hay or silage, fed to animals during scarcity period. The previous one is dehydrated product, whereas the latter one is an anaerobically fermented high moisture product. The above aspect may affect the rate of utilization of either of the stored forages in the rumen. The rate of utilization of the fodder can be best estimated from their intake level and nutrient digestibility. Utilizable crude protein (Zhao and Lebzien 2000) gives the idea of the protein entering the duodenum for digestion. Methane is a major greenhouse gas produced especially from fodder digestion in ruminant livestock which not only contribute to environmental pollution, but also results in substantial loss (2-12%) of gross energy intake which could have been otherwise utilized for productive purposes. Buffalo and cattle contribute about 39 and 53.1% of total livestock methane production in India (Singhal et al., 2005). Considering the fact that environmental issues can no longer be ignored while designing feeding regimes for large ruminants, it was hypothesized that comparative nutritive value and methane mitigating potential would vary according to methods of conservation of forage. The specific objective of this experiment was to compare nutrient utilization and methane emissions in Murrah buffaloes fed either oat hay or silage diets.

#### MATERIAL AND METHODS

The fodders were evaluated for their chemical composition and *in vitro* utilizable crude protein (Zhao

\*Corresponding author Email: world.sonalindri@gmail.com Assistant professor, Animal Nutrition, College of Veterinary Sc. and A.H., Anjora, Durg, Chhattisgarh; <sup>1</sup> Livestock Development Officer, Panchayat Samiti Etapalli, Gadchiroli, Maharashtra; <sup>2</sup> Former Head of Division, Dairy Cattle Nutrition, ICAR-National Dairy Research Institute, Karnal, Haryana; <sup>3</sup>Subject Matter Specialist, Krishi Vigyan Kendra, Kathua, SKUAST, Jammu

and Lebzien 2000) content before start of the animal experiment. The experiment was conducted in the experimental shed of the National Dairy Research Institute, Karnal, India (latitude 29.43°N & longitude 77.2°E). Environmental temperature during the study ranged from 27 to 31 °C, and humidity ranged from 16 to 18 %. All the experimental procedures were approved by Institutional Animal Ethics Committee of NDRI (IAEC/21/14 dated 04.01.2014).

Sixteen dry Murrah buffaloes ( $566.5 \pm 12.5 \text{ kg}$  body weight) were selected from Livestock Research Centre of National Dairy Research Institute, Karnal and distributed randomly into two groups based on body weight. Animals were housed in Dairy Cattle Nutrition experimental shed and given an acclimatization period of 10 days. Animals in group 1 were given hay and those of group 2 were fed silage for 60 days. In both the groups mineral mixture and salt were supplemented. A digestibility trial of 7 days duration was conducted to evaluate the nutrient digestibility. Methane emission was measured by  $SF_6$  tracer technique (Johnson *et al.*, 1994). A permeation tube containing  $SF_6$ , standardised

for steady release rate (0.0035 g/day) over a period of 6 weeks, was placed in the rumen 2 days prior to the experiment. The animals were fitted with halter with capillary tubing (SS Capillary tubing 0.0635" X 0.004") in such a way that the gas collection inlet was close to nose. The eructed gas from nose and mouth was collected in an evacuated canister. Five collections each of 24-h duration were made for each animal. Methane and SF<sub>6</sub> concentrations in the collected sample were quantified using a gas chromatograph (Nucon-5700) fitted with flame ionization and electron capture detectors (FID and ECD), respectively. The data obtained were subjected to the statistical analysis as per Snedecor and Cochran (1994).

#### RESULTS AND DISCUSSION

Detailed chemical composition of oat silage and oat hay is presented in Table 1. A relatively higher content of CP was observed in oat silage (11.97%) as compared to oat hay (9.20. The NDF (56.17 vs 43.16%) and ADF (47.40 vs 32.35%) content in oat hay were higher than the oat silage.

Average daily dry matter intake (DMI kg/day)

Table 1. Chemical composition of oat hay and oat silage fed to buffaloes

Parameter	Oat hay	Oat silage
DM (%)	87.00±0.72	26.72±0.80
	% DM	
OM	91.10±0.15	90.79±0.12
CP	9.21±0.22	11.97±0.09
EE	1.82±0.02	1.97±0.01
NDF	56.17±0.41	47.40±0.52
ADF	43.16±0.27	32.35±0.31
Ash	8.90±0.19	9.21±0.29
NDICP	1.3±0.07	2.1±0.09
ADICP	$0.6\pm0.03$	$1.0\pm0.06$
Hemicellulose	13.02±0.69	15.05±0.43
Cellulose	40.53±0.23	41.11±0.31
ADL	8.29±0.65	6.01±0.39
TDN %	54.61±0.61	56.83±0.72
DE (MJ/kg)	10.07±0.03	10.48±0.12
ME (MJ/kg)	8.29±0.05	8.71±0.01
uCP	10.01±0.49	10.32±0.97

Table 2. Effect of feeding oat hay or oat silage on intake and digestibility of nutrients in dry buffaloes

Parameter	Oat hay group (T <sub>1</sub> )	Oat silage group (T <sub>2</sub> )
BW, kg	566±21.29	567±29.65
DM intake, kg	8.30±0.19	8.34±0.20
DMI intake, (kg/100kg BW)	1.46±0.01	1.47±0.01
DMI intake, W <sup>0.75</sup>	71.07±0.66	71.55±0.67
CP intake, kg	$0.82^{b}\pm0.04$	$1.06^{a}\pm0.07$
CP intake, g/kgW <sup>0.75</sup>	$6.54^{b}\pm0.06$	$8.73^{a}\pm0.08$
EE intake, kg/d	$0.15 \pm 0.03$	0.16±0.01
NDF intake, kg/d	$4.66^{a}\pm0.04$	$3.95^{b}\pm0.21$
ADF intake, kg/d	$3.58^{a}\pm0.03$	$2.70^{b}\pm0.02$
Digestibility coefficient (%)		
DM	68.87±2.19	69.09±1.76
OM	70.44±3.33	69.51±1.88
CP	58.28b±1.78	$60.59^{a}\pm2.72$
EE	80.18±2.92	82.99±3.11
NDF	64.50±2.22	67.99±1.89
ADF	61.31±1.41	61.46±1.12

<sup>&</sup>lt;sup>a,b</sup>Means bearing different superscripts in a row differ significantly (\*P<0.05)

was similar (P>0.05) in both the groups ( $T_1$ ,8.30 and  $T_2$ , 8.34 kg/d). Neutral detergent fibre and acid detergent fiber intake was significantly (P<0.05) higher in  $T_1$  than  $T_2$ , while the CP intake was significantly (P<0.05) higher in  $T_2$  than  $T_1$  group. Wallsten *et al.* (2009) reported increasing intake of oat silage DM with increasing plant maturity when fed to heifers. The increase was from 1.6 kg/100 kg LW at early milk stage to 2.0 kg/100 kg LW at dough stage. In the present study silage DMI averaged 1.47 kg/100 kg BW for buffaloes.

y = 67.194x - 74.458445.00  $R^2 = 0.6119$ 420.00 405.00 358.00 365.00 345.00 325.00 305.00 285.00 265.00 6.00 6.50 5.00 5.50 7.00 NDF intake (kg/d)

Fig 1. Relationship between neutral detergent fibre intake (NDFI) and methane emissions in buffaloes

Data pertaining to the nutrient digestibility of buffaloes fed on oat hay or silage are presented in Table 2. The CP digestibility (%) was significantly (P<0.05) higher in  $T_2$  than  $T_1$ , whereas digestibility coefficients of all other nutrients were similar between the groups. The DM and CP digestibility coefficients were 68.87 and 69.09; 58.28 and 60.59% in  $T_1$  and  $T_2$ , respectively. Silage protein usually serve as rumen degradable protein (RDP) as most of the proteins are broken down during fermentation to non-protein

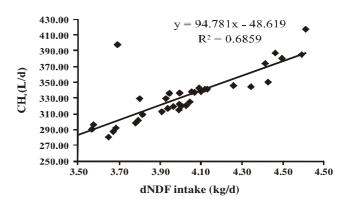


Fig 2. Relationship between digestible neutral detergent fibre intake (dNDFI) and methane emissions in buffaloes

nitrogen, like ammonia nitrogen, (Papadopoulos and Mckersie, 1983; Wilkinson, 2005). In the present study *In vitro* utilizable CP were found to be similar in both the groups.

The energy intake and loss of energy in the form of methane from buffaloes fed on oat hay or silage is presented in Table 3. The energy intake in terms of gross energy (GE) and digestible energy (DE) was similar between the groups. The overall methane production was significantly lower (P<0.05) in oat silage fed group than oat hay fed group. The higher methane emission (g/kg DM, MJ/kg DM and g/kg NDF intake) was in T<sub>1</sub> (24.36, 1.36 and 35.68) as compared to  $T_2(21.79, 1.21)$ and 34.52). Percentage of DE and ME lost in methane was 13.51, 16.41% and 11.61, 13.97% in  $T_1$  and  $T_2$ , respectively. The difference in energy loss was analysed to be significant (P<0.05). Methane loss (MJ/d) was 6.86% lower in dry buffaloes fed on oat silage instead of oat hay-based diets. Methane release in buffalo in relation to NDF intake and digestible NDF intake is depicted in fig. 1 and 2, respectively. Significant correlation was observed between methane emissions and NDF intake ( $R^2 = 0.61$ , P<0.05). Sundstol (1981) observed a reduction in loss of gross energy in the form of methane when forages were ensiled than when dried.

This could be due to reduced rate of carbohydrate fermentation in the rumen with silage as extensive fermentation already occurred during ensiling. Similar findings to present study were reported on comparison of alfalfa hay and silage (Benchaar et al., 2001). They used a mechanistic model approach to assess a 33% depression in total CH<sub>4</sub> production (Mcal/d) on alfalfa silage rather than alfalfa hay. Kirkpatrick and Steen (1999) observed no differences in gross energy loss (%) as methane when forages were conserved either as silage or by freezing directly after harvesting. Silage based diets when fed to Merino lambs recorded reduced loss of energy via methane, urine and faeces (Beukes, 2013). Martin et al. (2008) observed less methane production on DM intake basis on corn silage based diet as compared to hay diet in two successive experiments. Martin et al. (2010) reviewed methane mitigation strategies to uncover that methane production tend to be lowered when forages were ensiled than when they were dried. Many studies suggest that methane yield in cattle respond in a quadratic manner when pasture forage was substituted with maize silage (Blaxter and Wainman 1964; Arndt et al., 2010). Blaxter and Clapperton (1965) found that both feed intake and digestibility were important factors affecting CH, yield

Table 3. Methane emissions and energy loss in buffaloes fed oat hay or oat silage

Parameter	Oat hay group (T <sub>1</sub> )	Oat silage group (T <sub>2</sub> )
DMI, kg/d	9.00±0.26	9.37±0.19
GEI, MJ/d	162.00±1.68	168.66±2.79
DEI, MJ/d	90.63±1.57	98.20±4.85
MEI, MJ/d	74.61 <sup>b</sup> ±1.31	81.61°±1.57
CH <sub>4</sub> , 1/d	341.35°±3.44	317.86 <sup>b</sup> ±6.28
CH <sub>4</sub> , g/d	219.27°±2.21	204.18b±4.03
CH <sub>4</sub> , g/kg DMI	24.36°±0.88	21.79b±0.82
CH <sub>4</sub> , g/kg NDFI	35.68°±0.22	34.52 <sup>b</sup> ±0.37
CH <sub>4</sub> , MJ/d	12.24°±0.12	11.40 <sup>b</sup> ±0.23
CH <sub>4</sub> , MJ/kg DM	$1.36^{a}\pm0.05$	$1.21^{b}\pm0.02$
CH <sub>4</sub> , MJ/kg DDM	$1.77^{a}\pm0.13$	$1.65^{b} \pm 0.03$
CH <sub>4</sub> loss as %GE	$7.56^{a}\pm0.05$	$6.76^{\text{b}} \pm 0.08$
CH <sub>4</sub> loss as % DE	13.51°±0.07	11.61 <sup>b</sup> ±0.14
CH <sub>4</sub> loss as %ME	16.41°±0.10	13.97 <sup>b</sup> ±0.19

<sup>&</sup>lt;sup>a,b</sup>Means bearing different superscripts in a row differ significantly (\*P<0.05)

(% of GEI) and that at the maintenance level of feeding CH<sub>4</sub> yield increased with higher digestibility of the diet. McCaughey et al. (1997, 1999) reported that feed intake rather than digestibility was the major determinant of CH<sub>4</sub> production. In the present study, where buffaloes were fed close to maintenance, neither feed intake nor feed digestibility was related to CH<sub>4</sub> yield (% of GEI), whereas, CH<sub>4</sub> production (g/d) was directly related to NDF intake and digestible NDF intake. A similar relationship was found in small ruminants (sheep and alpaca) fed forages ad libitum indoors or grazed (Pinares-Patiño et al., 2003). Moe and Tyrrell (1980) also advocated the concept of CH<sub>4</sub> production as a function of the amount of cell walls digested in the rumen. Singh et al. (2011) attributed the variation in CH<sub>4</sub> production from dry roughages to significant difference in the NDF, ADF, carbohydrate and protein fractions. Woodward et al. (2001) found CH<sub>4</sub> production of 26.9 and 35.1 g/kg DMI in lactating dairy cows fed on silages made from Lotus corniculatus and pasture, respectively.

#### **CONCLUSIONS**

Enteric  $\mathrm{CH_4}$  emissions (I/d) was significantly higher (P<0.05) in oat hay fed group (341.35) than oat silage fed group (317.86). Total  $\mathrm{CH_4}$  production was reduced by 6.86% in dry buffaloes fed on oat silage instead of oat hay, whilst nutrient intake and digestibility values remained unaltered. Thus, oat fodder could be better preserved as silage than hay for lowering methane emission from Murrah buffaloes.

#### REFERENCES

- AOAC. 2005. Official Methods of Analysis, 18<sup>th</sup> edn. Association of Official Analytical Chemists. Arlington, Verginia, USA.
- Arndt, C., Wattiaux, M.A., Powell, J. M. and Aguerre, M.J. 2010. The effect of dietary alfalfa silage to corn silage ratio on lactating cow performance and methane emission. Dairy Forage and Agricultural Research, USDA, Madison, Wisconsin, USA, https://www.ars.usda.gov/research/publications/publications.htm?seq\_no\_115 = 257454.
- Benchaar, C., Pomar, C. and Chiquette, J. 2001. Evaluation strategies to reduce methane production in ruminants: a modelling approach. *Canadian J. Anim. Sci.* 81:

- 563-574.
- Beukes, J. 2013. Maize silage based diets for feedlot finishing of merino lambs. Master's thesis: Stellenbosch University, South Africa.
- Blaxter, K.L. and Clapperton, J.L. 1965. Prediction of the amount of methane produced by ruminants. *Br. J. Nutr.* 19:511-522.
- Blaxter, K.L. and Wainman, F. W. 1964. The utilization of the energy of different rations by sheep and cattle for maintenance and for fattening. *J. Agric. Sci.* 63: 113-128
- Johnson, K.A., Huyler, M.T., Westberg, H.H., Lamb, B.K. and Zimmerman, P. 1994. Measurement of methane emissions from ruminants livestock using a SF<sub>6</sub> tracer technique. *Environ. Sci. Technol.* 28: 359-362.
- Kirkpatrick, D.E. and Steen, R.W.J. 1999. The effect of method of conservation of grass and supplementation on energy and nitrogen utilization by lambs. *J. Agric. Sci.* 133: 409-417.
- Martin, C., Rouel, J., Jouany, J. P., Doreau, M. and Chilliard, Y. 2008. Methane output and diet digestibility in response to feeding dairy cows crude linseed, extruded linseed, or linseed oil. *J. Anim. Sci.* 86: 2642-2650.
- Martin, D., Morgavi, P. and Doreau, M. 2010. Methane mitigation in ruminants: from microbe to the farm scale. *Animal*. 4: 351-365.
- McCaughey, W.P., Wittenberg, K. and Corrigan, D. 1999. Impact of pasture type on methane production by lactating beef cows. *Canadian J. Anim. Sci.* 79: 221-226.
- McCaughey, W. P., Wittenberg, K. and Corrigan, D. 1997. Methane production by steers on pasture. *Canadian J. Anim. Sci.* 77: 519-524.
- Moe, P.W. and Tyrrell, H.F. 1980. *Methane Production in Dairy Cows. In Energy Metabolism*, ed. L.E. Mount Publication.
- NRC. 2001. *Nutrient Requirements of Dairy Cattle*: Seventh Rev. Edn. National Research Council, National Academy of Sciences. Washington, D.C.
- Papadopoulos, Y.A. and Mckersie, B.D. 1983. A comparison of protein degradation during wilting and ensiling of six forage species. *Canadian J. Plant Sci.* 63: 903-912.
- Pinares-Patiño, C.S., Ulyatt, M.J.G., Waghorn, C., Lassey, K.R., Barry, T.N., Holmes, C.W. and Johanson, D.E. 2003. Methane emission by alpaca and sheep fed on Lucerne hay or grazed on pastures of perennial ryegrass/white clover or birdfoot trefoil. *J. Agric. Sci.* 140: 215-226.

- Singh, S, Kushwaha, B. P., Nag, S. K., Mishra, A.K., Bhattacharya, S., Gupta, P.K. and Singh, A. 2011. *In vitro* methane emission from Indian dry roughages in relation to chemical composition. *Curr. Sci.* 101: 57-65.
- Singhal, K.K., Mohini, M., Jha, A.K. and Gupta, P.K. 2005. Methane emission estimates from enteric fermentation in Indian livestock: Dry matter intake approach. *Curr. Sci.* 88: 119-127.
- Snedecor G.W. and Cochran, W.G. 1994. *Statistical Methods*. 8<sup>th</sup> edn. IOWA State University Press, Ames, Iowa, USA.
- Sundsol, F. 1981. *Methods for Treatment of Low Quality Roughages*. Pages 61-80 in Kategile J A, Sand A N and Sundstol F. edn. Utilization of low quality roughages in Africa. Agricultural University of Norway, Norway.

- Wallsten, J, Nadeau, E., Bertilsson, J. and Martinsson, K. 2009.
  Voluntary intake and diet selection by dairy heifers fed ensiled whole-crop barley and oats harvested at different stages of maturity. *Livest. Sci.* 122: 94-98.
- Wilkinson, J.M. 2005. *Silage*. Chalcombe Publications, Painshall, Welton, Lincoln, United Kingdom.
- Woodward, S.L., Waghorn, G.C., Ulyatt, M.J. and Lassey, K.R. 2001. Early indications that feeding Lotus will reduce methane emissions from ruminants. Proc. N. Z. Soc. *Anim. Prod.* 61: 23-26.
- Zhao, G.Y. and Lebzien, P. 2000. Development of an *in vitro* incubation technique for the estimation of the utilizable crude protein (uCP) in feeds for cattle. *Arch. Anim. Nutr.* 53: 293-302.

Received on 19-09-2019 and accepted on 26-09-2019



### Effect of Depotash Vinasse on Rumen Fermentation Kinetics in vitro

# Gaurav Pratap Singh, Vandana Kumari Leitanthem, Amit N. Sharma, Madhu Mohini, Naresh Arora¹ and Goutam Mondal\*

Animal Nutrition Division, ICAR-National Dairy Research Institute, Karnal-132001, Haryana, India

#### **ABSTRACT**

Vinasse is a by-product of distilleries which may be used in animal feeding otherwise it is a pollutant of water and soil. An *in vitro* experiment was conducted to observe the effect of molasses and depotash vinasse on rumen fermentation parameters such as *in vitro* dry matter digestibility (DMD), organic matter digestibility (OMD), ammonia nitrogen (NH<sub>3</sub>-N), partition factor (PF) and microbial biomass production (MBP). Concentrate mixture, pellet with molasses (Pellet M) and pellet with depotash vinasse (pellet DPV) were prepared with same ingredients while pellet M contained 8% molasses and pellet DPV contained 8% depotash vinasse (as at this level binding was optimum) with little change of ingredients, chemical composition was similar. The substrate for the *in vitro* experiment comprised of concentrate mixture/ pellet M/, pellet DPV individually and TMRs with concentrate/ pellets, oat green and wheat straw in different proportions (40:40:20). The results revealed no noticeable changes due to addition of depotash vinasse and molasses pellets (upto 8%) on rumen fermentation parameters. DM, OM and crude protein (CP) digestibility was similar in all the groups. MBP and NH<sub>3</sub>-N was also similar in all the groups. It was concluded that depotash vinasse can be used in pellet making and subsequent feeding to animals without any harmful effect on rumen fermentation parameters.

Keywords: Composition and In vitro digestibility, Depotash vinasse, Rumen fermentation

#### INTRODUCTION

Vinasse is a by-product from industrial production of yeasts, alcohol, citric acid or other substances by fermentation of molasses (Fernandez et al., 2009). Vinasse is produced mostly from raw materials viz, corn, wheat, rice, potatoes, sugar beets, sugarcane and sweet sorghum. It is also called as condensed molasses solubles (CMS). It is a light-brown to dark brown liquid which contains water (93%), organic solids and minerals (7%) that is condensed to 60% (on wet basis) and is variable depending on source, batch and location etc. though, for production of each liter of ethanol, 12-15 liter of water is used which cannot be recycled (Christofoletti et al., 2013). The vinasse have generally been disposed directly to the flowing water that affects water quality, subsequently aquatic life, thus, a threat to ecosystem. As an alternate to control the water pollution, significant proportion of vinasse have been disposed to agriculture land (Nataraj et al., 2006). Vinasse has less energy, high ash content. It is especially rich in sulphur (S) and potassium (K), than molasses. Vinasse may be used in feeding of cattle

and sheep because its crude protein consists mainly of non-protein nitrogen (NPN) compounds such as free amino acids and betaine. In cattle and poultry feed, vinasse may be used as binder and dust reducer rather than as a protein replacer. Use of vinasse beyond 5% level as a replacement for molasses in ruminant diet results reduced voluntary feed intake, weight gain and feed conversion efficiency (Campos et al., 2014). Possible reason for poor performance of animal on vinasse containing diet is due to its high level of S. High S content in vinasse reduces selenium (Se) and copper (Cu) absorption and responsible for excess production of hydrogen sulphide (H2S) in rumen. High K reduced magnesium (Mg) absorption which in turn leads to hypomagnesaemia. However, depotash vinasse, (after removal of maximum potassium) kind of new product with minerals, low CP with binding properties, though not exploited yet in India or elsewhere. Moreover, number of rumen protozoa and the amount of ruminal ammonia nitrogen in dairy cattle and sheep were also reported to be increased due to vinasse in diet (Sugoro et al., 2016). Under these circumstances, depotash

\*Corresponding author Email: gmondal1075@gmondal1075@gmail.com; ¹Sr Technical Officer, CSSRI, Karnal-132001, Haryana, India

vinasse have been used for preparing feed pellet and total mixed ration and tested for *in vitro* rumen fermentation and nutrient digestibility.

#### MATERIALS AND METHODS

Samples of molasses were collected from Sagbara, Gujarat and depotash vinasse was collected from Demo plant at Kemrej, Gujarat. Physical characteristics of depotash vinasse and molasses were observed visually and pH by digital pH meter. The cattle feed pellets were manufactured at Nand Gopala Feeds, Sagbara, Gujarat. Ingredients used for pellets preparation were maize grain, mustard cake, ground nut cake, wheat bran, mineral mixture and salt. Molasses and depotash vinasse were added at the level of 8% in each formulation by little changing the composition (Table 1). Eight percent level of molasses and depotash vinasse was selected because at this level binding and pelleting were optimum. The chemical composition viz., moisture, CP, total ash, acid insoluble ash (AIA) of depotash vinasse, molasses, concentrate mix and pellets were analyzed by following protocol of AOAC (2007) and mineral composition viz., calcium (Ca), Mg, K, sodium (Na), Cu, cobalt (Co), zinc (Zn) and manganese (Mn) were analyzed by using Atomic Absorption Spectrophotometer (AAS; Hitachi Z-5000, Polarized Zeeman) (Table 2).

TMR was prepared by mixing of concentrate mixture, and pellets with green oats and wheat straw in 40:40:20 ratios. A total of 6 groups were there *i.e.* concentrate mixture, pellet with molasses, pellet with depotash vinasse individually and TMR with

concentrate mixture: green oats: wheat straw (40:40:20), TMR with molasses containing pellet: green oats: wheat straw (40:40:20) and TMR with depotash vinasse containing pellet: green oats: wheat straw (40:40:20). Samples of TMRs in triplicate were used to observe the gas production and nutrient digestibility (Menke and Steingass, 1988).

The generated data were analyzed by one way ANOVA using IBM SPSS version 20 to observe the treatment effect on fermentation kinetics.

#### RESULTS AND DISCUSSION

Colour of the molasses was light brown while that of depotash vinasse was dark brown. Odour of the molasses was sweetish while it was mild sweetish for depotash vinasse. pH of the samples varied between 4.5-5.1. These parameters indicated that depotash vinasse was visually similar to the molasses. Chemical composition of depotash vinasse and molasses is depicted in Table 2. DM of depotash vinasse was around 76.36% and in molasses it was 60.85%. The OM varied from 85-90% in both the samples. The CP content was higher in depotash vinasse than molasses (7.70 vs. 5.36%). Total ash content was higher in depotash vinasse than molasses due to higher mineral content. Acid insoluble ash was 1.5 to 2.01% in both the samples. Calcium level was higher in DPV and low in the molasses varied from 0.4-0.8% in DPV and 0.2–0.4% in molasses. The concentration of potassium low in depotash vinasse as well as in molasses (3.3-3.9%). The concentration of sodium was higher in DPV, varied from 0.1-0.3% and low in molasses at

Table 1. Ingredient composition of concentrate mixture and pellets (% DM basis)

Ingredient	Concentrate mix	Molasses pellets	Vinasse pellets
Maize	40	35	35
Ground nut cake	10	12	12
Mustard oil cake	20	21	21
Wheat bran	27	21	21
Molasses	-	8	-
De-potash vinasse	-	-	8
Mineral mixture	2	2	2
Common salt	1	1	1

Table 2. Chemical composition of depotash vinasse and molasses (% DM basis or as mentioned)

Parameter	Depotash vinasse	Molasses
DM	76.36	60.85
CP	7.70	5.36
Total ash	16.49	8.66
AIA	2.01	1.52
S	2.05	0.67
Ca	1.20	0.33
Mg	2.80	0.19
K	1.80	3.26
Na	0.26	0.07
Cu (ppm)	45.26	35.08
Co (ppm)	19.21	17.00
Zn (ppm)	8.62	9.16
Mn (ppm)	55.51	35.21

around 0.1%. The concentration of zinc varied from 8.2–9.6 ppm in DPV and 9.6–10.4 ppm in molasses, whereas spent wash had high zinc concentration around 112 ppm. The iron concentration was high in DPV,

varied from 92–585 ppm and low in molasses around 33–154 ppm. The value of manganese was 55 and 35 ppm which was higher in DPV followed by in molasses group, respectively. The finding of present study was

Table 3. Chemical Composition of concentrate mixture and pellets samples

Parameter	Concentrate	Pellet (M)*	Pellet (V)**
Chemical composition (%	% DM)		
DM	90.4±0.27	89.3±0.18	88.7±0.15
OM	89.5±0.18	90.6±0.11	89.5±0.05
CP	20.6±0.15	19.3±0.12	21.7±0.17
EE	4.47±0.19	4.83±0.02	4.75±0.05
NDF	24.1±0.17	26.7±0.05	25.7±0.09
ADF	12.1±0.25	13.7±0.05	12.3±0.05
CF	6.15±0.24	$5.60\pm0.02$	4.99±0.05
AIA	2.15±0.05	$2.35 \pm 0.05$	1.07±0.01
	10.5±0.16	$9.40\pm0.03$	10.6±0.05
Mineral composition (%	or as mentioned)		
Ca	0.61±0.01	$0.71 \pm 0.01$	$0.67 \pm 0.04$
K	2.12±0.17	$2.35 \pm 0.05$	2.35±0.05
Na	$1.84 \pm 0.24$	$0.99 \pm 0.02$	$1.04 \pm 0.02$
Mg	$0.81 \pm 4.42$	$0.84 \pm 6.12$	$0.79 \pm 6.22$
Cu (ppm)	29.42±2.74	28.5±3.52	27.08±2.11
Zn (ppm)	42.1±8.32	41.55±1.54	45.3±1.22
Fe (ppm)	210.1±8.41	221.5±14.5	206.2±6.58
Mn (ppm)	69.5±2.52	66.3±1.45	63.55±2.09
Durability Index (%)	-	95.71%	96.74%

Table 4. Effect of molasses and depotash vinasse on fermentation kinetics

Parameter	Group						P-Value
	Concentrate	Pellets (M)	Pellets (V)	TMR Conc	TMR (M)	TMR (V)	'
Net gas(ml/24h)	35.07±6.40	34.47±1.56	36.63±1.79	33.88±1.35	35.93±1.09	32.81±0.23	0.935
Gas (ml/kg	175.35±32	172.34±7.79	183.13±8.94	169.41±6.77	179.67±5.47	164.04±1.16	0.935
DM digested)							
IVDMD (%)	61.52±.66	62.19±2.0	62.32±0.64	62.89±2.0	62.79±2.4	60.65±0.60	0.907
TOMD (%)	65.16±2.21	65.68±0.04	68.69±1.15	63.36±2.3	66.10±5.9	61.84±0.62	0.643
PF	$3.49 \pm 0.52$	$3.45 \pm 0.15$	$3.37 \pm 0.12$	$3.48 \pm 0.04$	3.36±0.21	$3.45 \pm 0.08$	0.997
MBP (mg)	39.91±10.3	40.98±3.40	40.57±2.0	41.10±0.86	39.38±8.69	38.97±1.92	1
NH <sub>3</sub> -N mg/dl)	6.40±0.50	$6.00\pm0.20$	5.50±1.5	$6.00 \pm 0.10$	$5.50 \pm 0.30$	5.10±0.20	0.774

Pellets (M), - molasses pellets; Pellets (V), - depotash vinasse pellets; TMR Conc, total mixed ration with concentrate mixture; TMR (M), - total mixed ration with molasses pellets; TMR (V), - total mixed ration with depotash vinasse pellets.

similar to that of Khan *et al.* (2006), they found similar chemical composition of cane sugar molasses. Brito *et al.* (2015) reported that molasses had a similar DM, CP, and Ca. The present findings are in line with the findings of Maneerat *et al.* (2015) and Zali *et al.* (2017). The difference in the chemical composition of molasses and vinasse may be due to nature of raw material used for alcohol making, their inclusion level, and the type of channels they pass through, and batch to batch variation, application of fertilizer during crop production.

Chemical composition of concentrate mixture and pellets prepared from molasses (PM) and depotash vinasse (PV) have been presented in Table 3. The value of DM, OM, CP, ether extract and NDF level were alike in concentrate mixture and both the pellets. Ca level was 0.6 to 0.7% in the feeds. Mineral concentration, including the trace minerals was comparable in all the groups.

Results of the *in vitro* experiment have been shown in Table 4. The net gas production and total gas production were similar in concentrate, pellets of molasses, pellets of vinasse and TMR with concentrate, with molasses pellets and with depotash vinasse pellets. It was similar statistically, though there were little variations among the groups. The *in vitro* dry matter digestibility (IVDMD) was similar with the average value around 62%. The total OM digestibility (TOMD) was also similar in all the groups with the average value of

66%. PF and MBP were not different among the samples of different groups. NH<sub>3</sub>-N was also similar among the samples, but higher with concentrate/ pellets individually as it was not utilized in optimum quantity by the microbes.

The results are similar with the findings of Mavimbela and Ryssen (2001) as they observed that the NH<sub>3</sub>-N concentrations in ruminal fluid did not differ between the treatment groups. The results are conflicting with Iranmehr *et al.* (2011) who found that the ruminal NH<sub>3</sub>-N and population of protozoa in rumen fluids were increased in all treatment groups. In the present experiment no such result observed.

#### **CONCLUSIONS**

Depotash vinasse, a by-product of distillery industry was evaluated for its chemical and mineral composition and found comparable to molasses. Its incorporation in pellet and incubation with rumen liquor individually and with TMR indicated no adverse effect on rumen fermentation characteristics and digestibility of nutrients, hence, can be used in cattle feed at 8% in concentrate mixture. Though, before incorporation in the dairy ration, it must be validated by *in vivo* experiments.

#### **ACKNOWLEDGEMENTS**

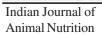
The authors are thankful to the Director, NDRI, Karnal for providing necessary facility and infrastructures. The authors are also thankful to Chem Process Systems Pvt. Ltd. Sannand, Gujarat for financial assistance.

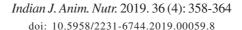
#### REFERENCES

- AOAC. Association of Official Analytical Chemist. 2007. Official Analytical Method. Washington, DC.
- Brito, A.F., Petit, H.V., Pereira, A.B.D., Soder, K.J. and Ross, S. 2015. Interactions of corn meal or molasses with a soybean-sunflower meal mix or flaxseed meal on production, milk fatty acid composition, and nutrient utilization in dairy cows fed grass hay-based diets. *J Dairy Sci.* 98: 443-457.
- Campos, C.R., Mesquita, V.A., Silva, C.F. and Schwan, R.F. 2014. Efficiency of physicochemical and biological treatments of vinasse and their influence on indigenous microbiota for disposal into the environment. *Waste Manage*. 34: 2036-2046.
- Christofoletti, C.A., Escher, J.P., Correia, J.E., Marinho, J.F.U. and Fontanetti, C.S. 2013. Sugarcane vinasse: environmental implications of its use. *Waste Manage*. 33: 2752-2761.
- Fernández, B., Bodas, R., López-Campos, O., Andrés, S., Mantecón, A.R. and Giráldez, F.J. 2009. Vinasse added to dried sugar beet pulp: Preference rate, voluntary intake, and digestive utilization in sheep. *J. Anim. Sci.* 87: 2055-2063.

- Iranmehr, M., Khadem, A.A., Rezaeian, M., Afzalzadeh, A. and Pourabedin, M. 2011. Nutritional value of vinasse as ruminant feed. *Krmiva*. 53: 3-8.
- Khan, M.A., Sarwar, M., Nisa, M., Iqbal, Z., Khan, M.S., Lee, W.S. and Kim, H.S. 2006. Chemical composition, in situ digestion kinetics and feeding value of Oat grass (*Avena sativa*) ensiled with molasses for Nili-Ravi buffaloes. *Asian-Australas. J. Anim. Sci.* 19: 1127-1133.
- Maneerat, W., Prasanpanich, S., Tumwasorn, S., Laudadio, V. and Tufarelli, V. 2015. Evaluating agro-industrial by-products as dietary roughage source on growth performance of fattening steers. *Saudi. J. Biol. Sci.* 22: 580-584.
- Mavimbela, D.T. and Van Ryssen, J.B.J. 2001. Effect of dietary molasses on the site and extent of digestion of nutrients in sheep fed broiler litter. *South Afr. J. Anim. Sci.* 31: 33-40.
- Menke, K.H. and Steingass, H. 1988. Estimation of the energetic feed value obtained from chemical analysis and *in vitro* gas production using rumen fluid. *Anim. Res. Dev.* 28: 7-55.
- Nataraj, S.K., Hosamani, K.M. and Aminabhavi, T.M. 2006. Distillery wastewater treatment by the membrane-based nanofiltration and reverse osmosis processes. *Water Res.* 40: 2349-2356.
- Zali, A., Eftekhari, M., Fatehi, F. and Ganjkhanlou, M. 2017.
  Effect of vinasse (condensed molasses solubles) on performance and meat chemical composition of Holstein male calves. *Ital. J. Anim. Sci.* 16: 515-520.

Received on 31-07-2019 and accepted on 03-12-2019







# The Effects of Dietary ω -3 and ω -6 Fatty Acids on Nutrient Utilization and Growth Performance in Sahiwal Heifers

Sushil Kumar<sup>1</sup>, Sajjan Sihag<sup>1</sup>, Zile Singh Sihag<sup>1</sup>, C.S. Patil<sup>2</sup>, Surender Singh Dhaka<sup>2</sup> and Anand Kumar Pandey<sup>3</sup>

Department of Animal Nutrition, Lala Lajpat Rai University of Veterinary and Animal Sciences, Hissar-125001, Haryana, India

#### **ABSTRACT**

This study was conducted to evaluate the effect of dietary supplementation with  $\omega$ -3 and  $\omega$ -6 fatty acid rich oil on nutrient utilization, growth performance and pubertal age in Sahiwal heifers. Eighteen healthy Sahiwal heifers of average 18.33±1.14 month's age and 194±4.16 kg body weights were randomly assigned to 3 treatment groups viz T<sub>1</sub>, T<sub>2</sub> and T<sub>3</sub> with 6 animals in each group. Heifers were individually fed diets of chopped wheat straw, green fodder and concentrate mixture to meet nutrient requirement as per ICAR (2013). The concentrate mixtures fed to the heifers under treatment group T, and T, were supplemented with  $\omega$ -6 fatty acid rich soybean oil and ω-3 fatty acid rich linseed oils @ 3.5 percent, respectively. To make the rations iso-caloric, palm oil @ 3.5 percent was supplemented in the concentrate mixture of heifers in control group (T<sub>1</sub>). All the concentrate mixtures were isonitrogenous. Average daily DMI was significantly (P<0.05) higher in heifers fed linseed oil supplemented concentrate mixture as compared to others. Total body weight gain and ADG was significantly (P<0.05) higher T, than T, and control heifers. The OMD was significantly (P<0.05) improved in groups fed ω-6 and ω-3 fatty acid rich oils as compared to control. The CP, DCP and TDN percent; and TDN intake did not differ among different dietary treatments. Feeding of linseed oil resulted in significant (P<0.05) reduction in the estrous age of Sahiwal heifers. The total dry matter intake up to first estrous was reduced by 32.12 and 12.77 percent in T<sub>3</sub> and T<sub>2</sub>, respectively, as compared to control (T<sub>1</sub>). Similarly, total concentrate mixture intake up to attainment of heifers' pubertal age in  $\omega$ -3 rich linseed oil and  $\omega$ -6 rich soybean oil fed group reduced by 38.82 and 16.11 percent, respectively, over control diet. The study deduced that ω-3 fatty acid supplementation reduced pubertal age of Sahiwal heifers.

Keywords: ω-3 and ω-6 fatty acids, Growth, Nutrient utilization, Pubertal age, Sahiwal heifers

#### INTRODUCTION

Livestock rearing is an important source of employment, income and livelihood of rural population of India. Demand of milk is ever increasing over time. For maximum output from commercial dairy farming sound rearing approach of heifers so as to replace the older and unproductive animals through culling play a significant role. However, heifer production is considered to be the most expensive part of dairy farm operation because it needs more inputs for a longer period with no immediate returns. Proper planning for replacement heifers affects success and longevity of a dairy herd. For successful dairy farming, a complete package of heifer rearing is imperative to exploit the performance potential at minimum expense. As reported by Razzaque *et al.* (2010) the input in dairy industry in

term of feed cost represents around 84% of the total input costs for heifer production. Heifers must reach 30 to 40% of their mature body weight in order to reach puberty. Adequate nutrient supply to the animal is paramount to development of heifers. Age at puberty is an important production trait; and hormones, nutrition, and genotype play key roles in attainment of puberty (Bellows and Hall, 1996).

High levels of grains in the concentrate mixture can adversely affect DMI, rumen fermentation and fiber digestibility. High level of dietary energy intake influences reproductive performance of animals, as reported by Barth *et al.* (2008) in young bulls through direct effect on the age at puberty and growth of scrotal circumference. Fat supplements can be used @ 20 to 30 g/kg DMI (Palmquist, 1988) without

Corresponding author Email: chahar53@gmail.com; <sup>1</sup>Department of Animal Nutrition, <sup>2</sup>Department of Animal Genetics and Breeding, <sup>3</sup>Teaching Veterinary Clinical Complex (Department of Veterinary Gynaecology and Obstetrics), LUVAs, Hissar

negatively affecting fibre digestibility.

The  $\omega$ -6 and  $\omega$ -3 FAs as well as their different dietary ratios can affect fermentation and nutrient digestibility in rumen (Zhang et al. 2008). Traditionally, lipids were added to the diet of bovine to increase the energy density and alleviate the negative energy balance during early lactation. But, recent studies have unveiled many beneficial effects of lipids other than providing energy, especially the one rich in polyunsaturated fatty acids. Some groups of FA are considered essential to mammals because mammalian cells are unable to synthesize them. Juchem (2007) indicated that between 70 and 85% of the PUFAs get biohydrogenated in the rumen when cows are fed with unprotected oils. Despite extensive biohydrogenation of unsaturated FA, feeding increasing amounts of  $\omega$ -6 and  $\omega$ -3 FA alters tissue composition and influences cellular function and animal performance and reproductive functions (Silvestre et al., 2011). Therefore, it is clear that despite limitations in delivery of specific amounts of polyunsaturated fatty acid for absorption, altering the FA composition of the diet is capable of influencing animal performance. Hence, this experiment was conducted to study effect feeding  $\omega$ -3 and  $\omega$ -6 fatty acid rich oil on nutrient digestibility and growth performance of sahiwal heifers.

# MATERIALS AND METHODS

The experiment was performed after taking due permission from Institutional Animal Ethics Committee, CPCSEA, New Delhi. The experiment was conducted at cattle farm, Department of Animal Genetics and Breeding, LUVAS, Hisar, using 18 Sahiwal heifers with no overt clinical sickness. The heifers averaged 194±4.16 kg in body weight and 18.33±1.14 months in age. The animals were assigned to 3 groups viz. T<sub>1</sub>, T<sub>2</sub> and T<sub>3</sub> of 6 animals each following completely randomized design. Animals were dewormed against internal parasites and dipped against ectoparasites before the start of the experiment following standard protocol. Routine vaccination against Hemorrhagic Septicemia and Foot and Mouth disease was done. All experimental animals were housed in feeding stalls having arrangements for individual

feeding and watering. The animals under different dietary treatments were maintained under similar husbandry practices. All the animals were fed isonitrogenous and isoenergetic diets containing weighed quantity of green fodder and concentrate mixture; and wheat straw was fed free of choice. The concentrate mixture fed to heifers in groups T2 and T3 was supplemented with soybean oil as ω-6 PUFA source and linseed oil as ω-3 PUFA source, respectively @ 3.5% (3.63 kg per 100 kg). To make the diets isocaloric, palm oil was added in the concentrate mixture of control (T<sub>1</sub>). The diets were fed twice daily at 8:00 AM and 4:00 PM. The proximate compositions of feed ingredients, fodder, wheat straw and the different concentrate mixtures were analyzed according to methods of AOAC (2005) and cell wall fractions were determined as per the procedure of Van Soest et al. (1991).

Amount of feed offered, residues left and body weight were measured at fortnightly interval for two consecutive days. A digestion trial of 7 days collection period was conducted during which quantitative collection of total feces voided on 24 hourly bases was made to determine the nutrients digestibility of nutrients. Feed cost was calculated according to the price of different feedstuff during the experiment at local market. Data obtained were subjected to statistical analysis as per Snedecor and Cochran (1994) following procedures for Completely Randomized Design (CRD).

# RESULTS AND DISCUSSION

Dry matter and crude protein content of  $T_1$ ,  $T_2$  and  $T_3$  were 93.16, 92.92 and 93.24 percent; and 21.25, 20.98 and 21.14 percent, respectively, indicating that concentrate mixtures were iso-nitrogenous. Ether extract and nitrogen free extract content were 5.32, 5.18 and 5.57; and 56.55, 55.22 and 55.03 percent in concentrate mixture of groups  $T_1$ ,  $T_2$  and  $T_3$ , respectively which revealed that all the concentrate mixtures were isocaloric. Accordingly, TDN intake and TDN content of the diets were also similar. The crude protein content of wheat straw and green fodder were 2.45 and 10.45 percent, respectively. Neutral detergent

fibre (NDF) and acid detergent fibre (ADF) contents in concentrate mixture of  $T_1$ ,  $T_2$  and  $T_3$  were 20.20, 20.86 and 19.63 percent; and 14.30, 14.50 and 14.35, respectively. The NDF and ADF content of wheat straw and green fodder were 75.82 and 50.37 percent; and 43.5 and 33.96 percent, respectively.

Data pertaining to intake and digestibility of nutrients are presented in Table 2. Intake of DM, CP, EE, CF, NFE, NDF and ADF was similar among the groups. The lack of significant effects on intake of these nutrients agrees with results of Muller *et al.* (2004) who showed that  $\omega$ -3 and  $\omega$ -6 fatty acids supplementation in crossbred heifers resulted in no differences (P>0.05) in intake of DM, CP, and NDF and ADF. Fiorentini *et al.* (2013) also found no effect

of adding soybean oil to the diets of crossbred heifers on nutrient intakes.

In the present investigation **PUFA** supplementation increased (P<0.05) the organic matter digestibility in T2, and T3 groups by 4.01 and 6.20 percent over that of control group. This corroborates well with the earlier reports by Kumar and Thakur (2007). Linseed oil supplementation increased the intake of digestible OM significantly (P<0.05) in T<sub>3</sub> (3160.35g). Linseed as well as soybean oil, both increased the digestible EE intake significantly (P<0.05) which might be due to the numerically higher d igestibility coefficient of EE in PUFA fed heifers. In the same way, Len et al. (2016) reported that dietary supplementation of fat sources rich in PUFA has

Table 1.Ingredient (kg/100 kg) and chemical composition (% DM basis) of different concentrate mixtures

Attributes	Co	ncentrate mixtu	ires	Rougl	nages
	T <sub>1</sub>	$T_2$	T <sub>3</sub>	Maize fodder	Wheat straw
<b>Ingredient Composition</b>					
Maize	30	30	30	-	-
Groundnut cake	25	25	25	-	-
Mustard cake	10	10	10	-	-
Wheat bran	32	32	32	-	-
Mineral mixture	2	2	2	-	-
Common salt	1	1	1	-	-
Total, kg	100	100	100	-	-
Palm oil, kg	3.63	-	-	-	-
Soybean oil, kg	-	3.63	-	-	-
Linseed oil, kg	-	-	3.63	-	-
<b>Chemical composition</b>					
DM (%)	93.16	92.92	93.24	25.11	94.43
OM	91.42	91.17	91.03	89.63	90.73
Crude protein	21.25	20.98	21.14	10.45	2.45
Crude fibre	8.30	9.79	9.29	26.40	36.79
Ether extract	5.32	5.18	5.57	4.31	2.87
NFE	56.55	55.22	55.03	48.47	47.62
Total ash	8.58	8.83	8.97	10.37	9.27
NDF	20.20	20.86	19.63	70.69	80.01
ADF	14.30	14.50	14.35	34.92	52.46

positive effects in the improvement of digestibility of EE and OM in Murrah buffalo calves. The CP% and TDN% of the different TMR (Table 3) were not affected due to PUFA addition. Similarly, DCP content was also statistically similar in all the rations. No noteworthy changes were seen in intake of CP, DCP and TDN among the different groups. CP intake varied (P>0.05) from 674.76 g/day in control to 685.40 g/day in T<sub>3</sub>. DCP intake was 480.46, 491.81 and 507.44 g/day in T<sub>1</sub>, T<sub>2</sub> and T<sub>3</sub>, respectively. Dietary PUFA supplementation increased the daily TDN intake from 3.251 kg/day in control to 3..352 kg/d in soybean and 3.491 kg/day in linseed oil fed heifers.

The overall mean values of daily total dry matter intake were 6.01, 6.25 and 6.67 kg in treatment groups  $T_1$  (palm oil),  $T_2$  (soybean oil) and  $T_3$  (linseed oil), respectively (Table 4). The statistical analysis of the data revealed that daily dry matter intake was significantly (P<0.05) higher in heifers fed linseed oil supplemented concentrate mixture as compared to palm oil or soybean oil supplemented groups. The higher value in linseed oil fed group might be due to comparatively more body weight gain in that group. Similar to this study, Childs *et al.* (2008c) also reported higher overall feed intake in fish oil supplemented crossbred heifers as source of  $\omega$ -3 fatty acids compared to control.

Table 2. Intake and apparent digestibility of nutrients in heifers under different dietary treatment groups

	Attribute		<b>Treatments</b>	
		T	$T_{2}$	$T_3$
DM	DM Intake, kg/d	5.00±0.03	4.97±0.18	5.14±0.06
	% digestibility	66.58±0.85	67.59±1.74	67.51±2.55
	DDM Intake, kg/d	3.33±0.04	3.38±0.20	3.47±0.14
OM	OM Intake, g/d	4434.84±66.31	4441.13±179.12	4419.25±148.62
	% digestibility	65.38 <sup>b</sup> ±0.67	69.39°±1.22	71.58°±0.60
	DOM Intake, g/d	2898.15b±33.04	$3071.52^{ab} \pm 75.32$	3160.35°±91.20
CP	CP Intake, g/d	674.76±1.27	674.00±6.68	685.40±1.93
	% digestibility	71.20±0.65	72.91±1.31	74.04±1.96
	DCP Intake, g/d	480.46±4.88	491.81±13.23	507.44±13.52
EE	EE Intake, g/d	244.76±5.83	246.32±0.12	248.83±0.62
	% digestibility	71.00±1.88	74.57±0.83	74.85±1.70
	DEE Intake, g/d	173.32 <sup>b</sup> ±2.44	183.67°±2.06	186.26°±4.44
CF	CF Intake, g/d	1101.02±15.60	1106.59±53.07	1194.16±4.53
	% digestibility	58.28±2.23	58.53±2.39	57.43±3.15
	DCF Intake, g/dd	641.76±26.31	653.94±56.57	686.33±39.28
NFE	NFE Intake, g/d	2468.22±21.95	2505.14±89.31	2609.70±10.72
	% digestibility	70.41±1.68	71.34±1.28	71.93±2.75
	DNFE Intake, g/d	1738.28±48.66	1792.40±93.52	1878.45±79.31
NDF	NDF Intake, g/d	3059.06±18.63	3121.16±128.03	3107.97±87.51
	% digestibility	60.32±1.93	59.51±2.27	58.93±2.45
	DNDF Intake, g/d	1846.22±65.46	1871.60±145.41	1828.63±82.05
ADF	ADF Intake, g/d	1639.13±26.65	1600.62±70.32	1691.70±11.55
	% digestibility	53.25±2.69	52.58±2.87	51.77±4.34
	DADF Intake, g/d	874.99±54.66	851.15±81.67	877.06±77.09

The mean values bearing different superscripts in a row differ significantly (P<0.05)

Table 3. Mean nutrient intake and nutritive value of experimental ration under different dietary treatments

	Attributes		<b>Treatments</b>	
			$T_2$	$T_3$
CP	CP%	11.65±0.02	11.64±0.12	11.84±0.03
	Intake, g/day	674.76±1.27	674.00±6.68	685.40±1.93
DCP	DCP%	8.30±0.08	8.49±0.23	8.76±0.23
	Intake, g/day	480.46±4.88	491.81±13.23	507.44±13.52
TDN	TDN%	55.01±1.53	56.70±3.26	59.49±2.63
	Intake, kg/day	3.251±0.08	3.352±0.16	3.491±0.13

There was no significant effect on DMI per 100 kg body weight by supplementing palm oil or soybean oil or linseed oil in concentrate mixture of Sahiwal

heifers. Similar trend was observed when dry matter intake was calculated in terms of dry matter intake per kg metabolic body size, which did not differ significantly

Table 4. Mean dry matter intake, growth performance and economics of feeding different diets (kg/day, DM basis) in experimental heifers

Attributes		<b>Treatments</b>	
_	$T_{1}$	$T_2$	$T_3$
DMI, kg/d	6.01b±0.12	6.25b±0.11	6.67°±0.14
DMI, %BW	$2.54 \pm 0.02$	2.53±0.03	$2.54 \pm 0.02$
DMI, g/kg W <sup>0.75</sup>	98.99±1.19	100.58±1.29	102.53±1.17
Total DMI, kg	901.53b±18.17	937.62 <sup>b</sup> ±16.66	999.98a±20.89
Av. roughage intake, kg	3.31b±0.09	$3.56^{b} \pm 0.08$	$3.96^{a}\pm0.12$
Av. CM intake, kg	$2.70\pm0.07$	$2.69 \pm 0.07$	$2.70\pm0.07$
Total roughage intake, kg	496.28b±14.19	533.42 <sup>b</sup> ±12.02	594.39a±17.27
Total CM intake, kg	405.25±10.61	404.20±10.58	405.59±10.62
Initial BW	185.67±9.30	196.17±4.38	201.16±6.79
Final BW	268.33b±7.11	283.00b±6.42	310.33°±7.79
Total gain, kg	82.67b±3.21	86.83 <sup>b</sup> ±6.75	109.17°±7.26
ADG, g	551 <sup>b</sup> ±21	579 <sup>b</sup> ±45	$728^{a}\pm48$
FCE	$0.09 \pm 0.01$	$0.09 \pm 0.01$	0.11±0.01
Days taken to 1st estrous	131.33 a±10.31	$110.17^{ab} \pm 13.79$	80.33b±13.66
Pubertal age, days	713.00°±29	665.33ab±26	623.67b±20
DMI till 1st estrous, kg	789.31±61.95	688.54±86.16	535.82±91.09
		(-12.77%)	(-32.12%)
Roughage intake till	434.71±34.12	392.19±49.07	318.12±54.08
1st estrous, kg			
CM intake till	354.67 a±27.83	297.50 ab ±37.14	217.00b±36.86
1st estrous, kg		(-16.11)	(-38.82)
CM cost, ₹/kg	24.53	25.51	28.13
Cost of CM feeding	8698.34±682.68	7587.95±949.47	6101.40±1037.26
till 1 <sup>st</sup> estrous, ₹		(-1110.39, -12.77%)	(-2596.94, -29.86%)

The mean values bearing different superscripts in a row differ significantly (P<0.05)

in different treatment groups. The results of the study are in agreement of earlier finding of Dirandeh *et al.* (2014) who reported that average dry matter intake was not affected by dietary inclusion of roasted soybean and linseed as a source of (P<0.05)  $\omega$  -6 and  $\omega$  -3 fatty acid, respectively, in the diets of lactating cattle.

Statistical analysis of the data revealed that total body weight gain during experimental period was significantly (P<0.05) higher in heifers fed linseed oil (ω-3 rich oil) supplemented concentrate mixture as compared to soybean oil ( $\omega$ -6 rich oil) and palm oil supplemented heifers(Table 4). Similar response with respect to average daily gain (ADG) was also noted. The results corroborate with findings of Childs et al. (2008) who found that on feeding of graded level of fish oil as source of ω-3 fatty acids significantly increased (P<0.05) the daily body weight gain of crossbred heifers as compared to non-supplemental groups of heifers. Kumar and Thakur (2007) also confirmed that bypass fat supplementation with varying degree of unsaturation increased growth rate in female buffalo calves. The present findings are also in agreement with the report Hill et al. (2011) who reported, ADG and efficiency of calves were improved by feeding ω-3 fatty acid rich feed as compared to control diets.

The results of the study revealed that numerical values of FCE improved upon supplementation of  $\omega$ -3 rich FA oil, however, the difference was statistically non-significant. In earlier report, Garcia *et al.* (2014) revealed that increasing mean intake of linoleic acid from approximately 4.6 to 11.0 g/day during the first 60 day of life increased average daily gain (0.50 vs. 0.45 kg/d) without a change in dry matter intake, thus improving feed efficiency.

The cost of feeding during experimental period and up to attainment of first estrous in Sahiwal heifers under different dietary treatments have been depicted in Table 4. The results of the study revealed that daily dry matter intake and total dry matter intake during 150 days of experimental period was significantly (P<0.05) higher in the animals fed concentrate mixture containing  $\omega$ -3 fatty acid rich oil (T<sub>3</sub>) as compared to  $\omega$ -6 (T<sub>2</sub>) and saturated fatty acid (T<sub>1</sub>) rich oils. However,

the experimental heifers fed soybean oil added concentrate mixture did not show any significant effect of dry matter intake over control group. The mean number of days taken from initiation of experiment to first estrous sign was 131.33, 110.17 and 80.33 days in  $T_1$ ,  $T_2$  and  $T_3$ , respectively, indicating that supplementation of linseed oil has significantly (P<0.05) reduced the days for first sign of heat over soybean and saturated fatty acid oils.

The findings of present investigation revealed that supplementation of  $\omega$ -3 rich linseed oil in the concentrate mixture of Sahiwal heifers resulted in significant (P<0.05) reduction in the pubertal age of heifers (623.67 days) as compared to  $\omega$ -6 rich soybean oil supplemented group (665.33 days) and control group (713 days) despite the fact that all the diets were isocalric and iso-nitrogenous. Tran et al. (2016) reported that feeding of ω-3 PUFA enriched diet reduced pubertal age and improved both fresh and post-thawing semen quality in male buffalo. The total dry matter intake up to first estrous was reduced by 32.12 and 12.77 percent in  $T_3$  and  $T_2$ , respectively, as compared to control  $(T_1)$ . Similarly, total concentrate mixture intake up to attainment of heifers' pubertal age in ω-3 rich linseed oil and ω-6 rich soybean oil fed group was reduced by 38.82 and 16.11 percent, respectively, over control diet. Hence, the perusal of data on feed cost demonstrated that there was a net saving of ₹2596.94 and 1110.39 in Sahiwal heifers by feeding PUFA rich linseed and soybean oil, respectively, from the age 18.33 months to pubertal age.

# **CONCLUSION**

The results of the present study inferred that feeding  $\omega$ -3 and  $\omega$ -6 fatty acid rich oil to Sahiwal heifers leads to improved organic matter digestibility and increased intake of digestible ether extract. In addition to that, a diet enriched with source of  $\omega$ -3 FA such as linseed oil might be a candidate diet to be fed to pre-pubertal heifers to increase the average daily gain; and decrease pubertal age and the cost of feeding.

#### **ACKNOWLEDGMENTS**

The authors are indebted to the Vice Chancellor, Lala Lajpat Rai University of Veterinary and Animal Sciences, Hisar, Haryana for providing the infrastructure facilities that enabled the successful completion of the research.

#### **REFERENCES**

- AOAC 2005. Association of Official Analytical Chemists. *Official Methods of Analysis* (20<sup>th</sup> ed.). Gaitherburg, Madison, USA.
- Barth, A.D., Brito, L.F.C. and Kastelic, J.P. 2008. The effect of nutrition on sexual development of bulls. *Theriogenol*, 70: 485-494.
- Bellows, R. A. and Hall, J.B. 1996. Physiology and management of the replacement heifer. In: L. M. Rode (ed.) Animal Science Research and Development, Meeting Future Challenges. *Proc. Canadian Soc. Anim. Sci. Lethbridge Res. Centre*, Lethbridge, Alberta. pp 217-234.
- Childs, S., Hennessy, A.A., Sreenan, J.M., Wathes, D.C., Cheng, Z., Stanton, C., Diskin M.G. and Kenny, D.A. 2008. Effect of level of dietary n-3 polyunsaturated fatty acid supplementation on systemic and tissue fatty acid concentrations and on selected reproductive variables in cattle. *Theriogenol*. 70: 595-611.
- Dirandeh, E., Towhidi, A., Zeinoaldini, S., Ganjkhanlou, M., Ansari, P.Z. and Fouladi-Nashta, A. 2014. Effects of different polyunsaturated fatty acid supplementations during the postpartum periods of early lactating dairy cows on milk yield, metabolic responses, and reproductive performances. *J. Anim. Sci.* 91:713-721.
- Fiorentini, G., Messana, J.D., Dian, P.H.M., Reis, R.A., Canesin, R.C., Pires, A.V. and Berchielli, T.T. 2013. Digestibility, fermentation and rumen microbiota of crossbred heifers fed diets with different soybean oil availabilities in the rumen. *Anim. Feed Sci. Technol.* 181: 26-34.
- Garcia, M., Greco, L.F., Favoreto, M.G., Marsola, R.S., Wang, D., Shin, J.H., Block, E., Thatcher, W.W., Santos, J.E. and Staples, C.R. 2014. Effect of supplementing essential fatty acids to pregnant nonlactating Holstein cows and their preweaned calves on calf performance, immune response, and health. *J. Dairy Sci.* 97: 5045-64.
- Hill, T.M., Bateman, H.G. Aldrich, J.M. and Schlotterbeck, R.L. 2011. Impact of various fatty acids on dairy calf performance. *Prof. Anim. Sci.* 27:167-175.
- ICAR. 2013). *Nutrient Requirements of Cattle and Buffalo*. 3<sup>rd</sup> edn. Indian Council of Agricultural Research. New Delhi, India.
- Juchem, S. O. 2007. Lipid digestion and metabolism in dairy cows: Effects on production, reproduction and health. PhD thesis. Univ. of California, Davis.

- Kumar, B. and Thakur, S.S. 2007. Effect of supplementing bypass fat on the performance of buffalo calves. *Indian. J. Anim. Nutr.* 24: 233-236.
- Len, T.V., Malla, B.A., Sharma, A.N., Kumar, S., Tyagi, N. and Tyagi, A.K. 2016. Effect of Different Bypass Fat Sources on Nutrient Utilization, Blood Parameters and Growth in Male Murrah Buffalo Calves. *Indian J. Anim. Nutr.* 33(4): 376-382.
- Müller, M., Prado, I.N., Lobo Jr., A.R., Capovilla, L.C.T. and Rigolon, L.P. 2004. Fat sources of ω-3 and ω-6 on the apparent digestibility of heifers confined. *Acta Sci. Anim. Sci.* 26:393-398.
- Palmquist, D.L. 1988. The feeding value of fats. In: Feed science (ed. ER Orskov), Elsevier Science Publisher, Amsterdam, Netherlands. pp. 293-311.
- Razzaque, M. A., Mohammed, S.A. and Al-Mutawa, T. 2010. Dairy heifers rearing in hot arid zone: An economi assessment. *Am. J. Appl. Sci.* 7: 466-472.
- Silvestre, F.T., Carvalho, T.S., Francisco, N., Santos, J.E., Staples, C.R. and Jenkins, T. 2011. Effects of differential supplementation of fatty acids during the peripartum and breeding periods of Holstein cows: I. Uterine and metabolic responses, reproduction, and lactation. *J. Dairy Sci.* 94:189-204.
- Snedecor, G.W. and Cochran, W.G. 1994. *Statistical Methods*. 8<sup>th</sup> edn., Iowa State University Press, Ames, Iowa, USA-50010.
- Tran, L.V., Malla, B.A., Sharma, A.N., Kumar, S., Tyagi, N. and Tyagi, A.K. 2016. Effect of ω-3 and ω-6 polyunsaturated fatty acidenriched diet on plasma IGF-1 and testosterone concentration, puberty and semen quality in male buffalo. *Anim. Reprod. Sci.* 173: 63-72.
- Van Soest, P.J., Robertson, J.B. and Lewis, B.A. 1991. Methods for dietary fiber, neutral detergent fiber, and non-starch polysaccharides in relation to animal nutrition. *J. Dairy Sci.*, 74: 3583-3597.
- Zhnag, C.M., Guo, Y.Q., Yuan, Z.P., Wu, Y.M., Wang, J.K., Liu, J.X. and Zhu, W.Y. 2008. Effect of octadeca carbon fatty acids on microbial fermentation, methanogenesis and microbial flora *in vitro*. *Anim. Feed Sci. Technol*. 146: 259-269.
  - Received on 19-10-2019 and accepted on 31-12-2019

# A Newly Developed Mixture of Herbal Plants and Spices Enriched with Special Extracts and Essential Oils Enhances Feed Utilisation, Growth Performance and Lowers Harmful Caecal Bacteria in Rabbits

O.A.H. El-Ghalid\*, A.M. Abd El-Hady\*, G.M. El-Ashry†, A.E. Kholif‡, O.H Matloup‡, O.A.Olafadehan# and A.M. El-Raffa\*

Poultry Production Department, Faculty of Agriculture, Alexandria University, 21545, Alexandria, Egypt

#### **ABSTRACT**

This study was conducted to evaluate the effect of feeding a mixture of herbal plants and spices enriched with special extracts and essential oils (phytogenic additives mixture) on feed intake, feed utilisation, blood chemistry, slaughter characteristics and caecal bacteria in rabbits. Ninety weaned unsexed V-Line rabbits, weighing 668.7±8.2 g and aged 30±1 day, were randomly allocated to three groups of 30 rabbits each. Animal were fed either a basal diet without additives (Control) or supplemented with 0.5 ml (LA) or 1 ml (HA) of the additives mixture per litre of drinking water. Additives did not affect feed intake; however, the LA rabbits showed better (P<0.001) feed conversion ratio as compared to the other treatments. The LA treatment increased (P<0.01) final body weight and average daily gain as compared to the control and HA treatments. Both the LA and HA treatments decreased (P<0.05) blood cholesterol, triglycerides and low-density lipoproteins; however, the LA treatment increased (P<0.05) high-density lipoproteins and total antioxidant capacity and decreased malondialdehyde (P<0.05) relative to the control. Treatments did not affect (P>0.05) carcass characteristics. The LA treatment followed by HA treatment increased (P<0.001) beneficial Lactobacillus spp. bacteria and decreased the concentrations of coliform bacteria and E. coli compared with the control. It is concluded that phytogenic additives mixture enhanced feed conversion, daily gain and positively altered caecal bacteria profile best response being observed at the lower dose of the phytogenic additives mixture as compared to the higher dose.

**Key words:** Blood metabolic profile, Carcass characteristics, Pathogenic bacteria, Phytogenic additives mixture, Rabbits, Weight gain

#### INTRODUCTION

The acute shortage of meat supply compels livestock farmers to improve feed utilization, health status and meat production from their animals. This shortage can be bridged by the farming of highly prolific animals with short production cycles, such as rabbits. Rabbits have high fertility rates and fast growth rates, making them good sources of meat. Rabbit meat is highly digestible, tasty, low in fat and cholesterol contents, rich polyunsaturated fatty acids content with higher nutritional advantage ( $\omega$ -6/ $\omega$ -3 ratio =5.9) and low in calorie content as compared to other meats (Combes, 2004; Dalle Zotte, 2002).

Feed additives such as herbal plants and seeds have been used in ruminants (Kholif *et al.*, 2017; Morsy

et al., 2018) and recently in the diets of rabbits (Cervantes-Valencia et al., 2015; Gado et al., 2016; Hafsa et al., 2016) to enhance feed conversion and health. Essential oils, spices and herbs have been successfully used in animal feed to increase the oxidative stability of meat and reduced total mesophilic aerobes, Pseudomonas spp. and Enterobacteriaceae (Fortier et al., 2012; Koné et al., 2016; Soultos et al., 2009). Combining herbal plants, spices, special extracts and essential oils (phytogenic additives mixture) has attracted an increased attention from rabbit producers due to its notable antioxidative, antibacterial and antifungal activities and ability to enhance feed flavour, palatability, voluntary feed intake and weight gain (Zeng et al., 2015). Studies on single components of herbal

\*Corresponding author: Dr. Ghada Mostafa el Ashry; E-mail: ghadaashry@yahoo.com \*Poultry Production Department, Faculty of Agriculture, Alexandria University, 21545, Alexandria, Egypt; †Regional Centre for Food Feed, Agriculture Research Centre, Ministry of Agriculture, Dokki, Egypt; †Dairy Science Department, National Research Centre, 33 Bohouth St. Dokki, Giza, Egypt; \*Department of Animal Science, University of Abuja, P.M.B. 117, Abuja, Nigeria;

plants, spices, special extracts or essential oils have showed positive effects on feed utilization, animal health and live performance of rabbits (Celia *et al.*, 2016). Benlemlih *et al.* (2014) reported that mortality was reduced in growing rabbits fed diet supplemented with 0.5 g fennel and thyme essential oil/kg diet. Furthermore, a diet supplemented with 0.5% fennel seed increased the digestibility of organic matter, crude fibre and ether extract, and final weight and body weight gain (Omer *et al.*, 2013).

Plant extracts, essential oils and herbal plants possess antimicrobial (Lambert et al., 2001) and antioxidant (Sarac et al., 2007) activities, and had been evaluated in broiler chickens (Lewis et al. 2003) and rabbits (Celia et al., 2016; Placha et al., 2013), with mixed results on performance and animal health. During the period around weaning, young rabbits are threatened by frequently-occurring enteric diseases. In order to control such problem there is extensive use of antibiotics (Bovera et al., 2010b, 2010a). Considering the harmful effects of antibiotics, there is an increasing interest among animal nutritionist to explore new and safe antimicrobial agents that can suitably replace antibiotics and effectively prevent and/or overcome infections, and improve welfare and health of the animals. Keeping these points in mind, the present experiment was conducted to study the effect of different doses of a phytogenic feed additive mixture (herbal plants, spices enriched with special extracts and essential oils) in the drinking water on feed conversion, blood metabolites, carcass characteristics and caecal bacterial profile of growing rabbits.

# MATERIALS AND METHODS

This experiment was conducted at the Nucleus Rabbit Breeding Unit of the Poultry Research Centre, Poultry Production Department, Faculty of Agriculture, Alexandria University, Egypt during spring season. The area has a climate with winter rains and an annual average rainfall of 22 mm and mean annual temperature ranges between 14 and 32°C. Rabbits care and procedures were conducted under established approved standards of the Animal and Fish Production

Department, Faculty of Agriculture, Alexandria University (Egypt).

A newly developed mixture (Aromix® 2336, Masa Egypt Company, Alexandria, Egypt) of natural finely ground herbs and spices enriched with special extracts and essential oils, was evaluated that contained (per litre):2.5 g carvacol, 0.75 g thymol, 17.8 g menthol and 50 g propylene glycol dissolved in 11 of distilled water.

Ninety weaned unsexed V-Line rabbits, aged 30±1 day and with a bodyweight of  $668.7 \pm 7.7$  g, were used in the study The total duration of the experiment was 63 days. During the first 2 weeks of the experiment, rabbits were housed in 5 galvanized wire pens (6 rabbits per pen) provided with feeders and automatic waterer system. They were there after housed in 10 galvanized wire pens (3 rabbits per pen) during the rest of the experiment. Each pen was 35 cm high × 40 cm width  $\times$  50 length. Pens were the experimental units for the intake and performance studies. All rabbits were kept under the same managerial, hygienic and environmental conditions in rooms with standard air conditioning maintaining 20-25°C temperature, 55-65% relative humidity and 14-16 h of lighting. Rabbits were treated subcutaneously with a coccidiostat (Sulphadimidine sodium BP solution Dimi-Vet® injection; Square Pharmaceuticals, Dhaka, Bangladesh) once at the beginning of the experiment at 1 ml/rabbit.

Rabbits of the control treatment were fed on a basal diet as a Control diet (Table 1). Other rabbits were supplemented with 0.5 ml (LA) or 1 ml of phytogenic additives mixture (HA) per litre drinking water. Feed was provided in two equal meals at 08:00 and 17:00 h daily. Droppings from the cages on the floor were collected and removed daily. Body weight and feed intake of rabbits were recorded weekly, while mortality rate was recorded daily. Feed conversion was calculated as the ratio of the feed intake (kg) to weight gain (kg).

At the end of the experiment, 10 rabbits were sampled and about 3 ml blood from each rabbit was taken at 08:00-09:00 h from the marginal ear vein into non-heparinized tube. Blood samples were centrifuged for 15 minutes on  $4000 \times g$ , serum harvested and stored

in a deep freezer at -20°C until biochemical analysis using specific kits (Stanbio Laboratory, Boerne, TX, USA) according to manufacturers' recommendations. Serum samples were analysed for total proteins, albumin, total lipids, triglyceride, total cholesterol, high density lipoprotein (HDL), low density lipoprotein (LDL), aspartate amino transferase (AST), alanine amino transferase (ALT), alkaline phosphate (ALP), uric acid, creatinine, calcium, inorganic phosphorus, immunoglobulin (IgG and IgM), glucose, triiodothyronine ( $T_3$ ), thyroxin ( $T_4$ ), total antioxidant capacity (TAC) and activity of malondialdehyde (MDA).

At the end of the experimental period (9 weeks of age), ten rabbits from each group (one from each pen) were randomly taken, fasted for 12 hours, weighed individually, slaughtered followed by exsanguinations (Cheeke, 1987). After complete exsanguinations, the carcass was weighed again and the difference between the pre-slaughter weight and weight after exsanguinations was considered as the blood weight. The slaughtered rabbits were skinned, the skin weight recorded, and the carcass eviscerated. The stomach, intestine, liver, heart, lung, and periscapular and perirenal fat were separated and weighed. The remaining part was considered as hot carcass. The following carcass traits were determined: dressing percentage (weight of hot eviscerated carcass including liver, heart, abdominal fat and head divided by the live body weight); organs weight as a percentage of live body weight (liver, heart, lung, pancreas, kidney, thyroid and adrenal glands, abdominal and shoulder fat, full stomach, full intestine and caecum). Slaughtering and dissection were carried out according to World Rabbit Science Association (WRSA) recommendations (Blasco and Ouhayoun, 1993). Hot carcasses were chilled at 4°C for 24 h. Intestine, caecum and carcass lengths were also measured.

Total number of caecal bacteria (total gut bacilli and lactobacilli populations) was counted according to American Public Health Association (APHA, 1960). The isolation of coliforms and *Escherichia coli* from the caecum was completed using the method described by the Food and Drug Administration (1984). Dilution

factor is a reciprocal value of dilution exponent. Such value is expressed as CFU/g (Colony Forming Units), (Barnes and Impey, 1970). Incubation was done at 30°C for 2-7 days. Microbiological contents of the caeca from 6 rabbits/ treatment were collected. The numbers of anaerobic bacteria, lactose entero-bacteria and coliform bacteria, were counted on appropriate selective and non-selective agar plates. The colony counts were expressed in log<sub>10</sub> CFU related to 1 g of sample. The pH of caecal content was determined with a glass electrode pH meter.

Except for bacterial counts and mortality data, data collected were analysed using the GLM procedure (SAS Inst. Inc. Cary, NC, USA) for a completely randomized design. The treatment (Control, LA or HA) was the only source of variation (fixed effect). For the intake and performance studies, pen within each treatment was the experimental unit, whereas for the carcass characteristics and caecal bacterial count study, rabbit within each treatment was the experimental unit. Measurements (body weight changes, feed intake and conversion, and mortality) recorded daily or weekly were analysed as a repeated measure. Bacterial count values were log transformed and analysed using the PROCGLIMMIX of SAS with log of bacterial counts as the response variables and diet (Control, LA or HA) as the factor explanatory variable, assuming that random residual variance followed a Normal distribution. When treatment effects were significant, the means were compared using Duncan's Multiple Range Test. Polynomial (linear and quadratic) contrasts (adjusted for the equal spacing of treatments) were used to examine dose responses to increasing doses of additives mixture and for comparison between control vs. the average of additives mixture treatments. Significance was declared when P<0.05.

# RESULTS AND DISCUSSION

Treatment LA showed greater (P=0.001) final body weight and average daily gain (P<0.001) compared with HA and control (Table 2). With no difference for feed intake (P>0.05), the LA rabbits showed better feed conversion ratio (P<0.001) compared with the other

treatments. The treatments LA and HA decreased mortality rate as compared to control. LA and HA treatments decreased serum cholesterol (P=0.002), triglycerides (P=0.023) and LDL cholesterol (P=0.032) (Table 3). The LA treatment increased HDL cholesterol (P=0.047) and TAC (P=0.049) and decreased MDA (P=0.041).

Additive doses did not affect pH of caecal content, carcass weight, relative organ weights, cecum length and carcass length (Table 4). The LA treatment showed greatest (linear and quadratic effects, P<0.001) beneficial effect with respect to *Lactobacillus* spp. count followed by the HA treatment while the control

Table 1. Ingredient and chemical composition of the basal diet fed to rabbits

Ingredient	g/kg DM
Alfalfa hay	240
Wheat bran	217
Soybean meal	180
Ground barley	160
Wheat straw	80
Yellow corn	70
Molasses	30
Di-calcium phosphate	12
Salt	4
Vitamins and minerals mixture <sup>1</sup>	3
Limestone	2
Methionine	1.5
Lysine	0.5
Chemical analysis	(g/kg DM)
Dry matter	917
Organic matter	902
Crude protein	177
Crude fibre	144
Ether extract	27
Nitrogen free extract	554
Digestible energy (kcal/kg) <sup>2</sup>	2584

<sup>1</sup>Each 1 kg contains: 13340 IU vitamin A, 2680 IU vitamin D<sub>3</sub>, 10 IU vitamin E, 2.68 mg vitamin K, 10.68 mg calcium pantothenate, 0.022 mg vitamin B<sub>12</sub>, 0.668 mg folic acid, 400 mg choline chloride, 26.68 mg chlorotetracycline, manganese 133.34 mg, 66.68 mg iron, 53.34 mg zinc, 3.2 mg copper, 1.86 mg iodine, 0.268 mg cobalt, 0.108 mg selenium; <sup>2</sup>DE Calculated according to Schneider and Flatt (1975).

had the least count (Table 5). Conversely, coliform bacteria and *E. coli* counts were lowest for the LA, followed by the HA and highest for the control (linear and quadratic effects, P<0.001).

The present study aimed to explore the effects of two levels of a phytogenic feed additive in drinking water of rabbits on feed intake and conversion, growth performance, carcass characteristics and some cecum microflora. Due to scarcity of available data on effect of phytogenic additives mixture on rabbits, comparison was done with studies that used different essential oils, spices and herbs either in rabbits or broiler chickens.

The insignificant differences among the treatments for the initial body weights reveal effective random distribution and homogeneity of the experimental rabbits at the beginning of the experiment. The low and high doses of the phytogenic additives mixture increased daily gain by about 13 and 6%, respectively, compared with the control. The increased daily gain and final body weight reveal enhanced feed utilization which may be partly explained by the increased nutrient absorption (Zeng et al., 2015) and improved secretions of saliva, bile, mucus and enzyme activity (Platel and Srinivasan, 1996, 2000; Jang et al., 2004; Jamroz et al., 2005; Muhl and Liebert, 2007). In addition, the observed decreased numbers of pathogenic bacteria in the cecum may improve the ability of epithelial cells to regenerate villus and thus enhance intestinal absorptive capacity (Mourão et al., 2006), resulting in improved daily gains. Similarly, the improved population of the beneficial lactic acid bacteria (Lactobacillus sp) possibly enhanced the gut function and health status of the additives mixture administered rabbits which might have improved feed utilisation and weight gains as a result of the substantial reduction in digestive disorders of the rabbits (Celia et al., 2016). Simitzis (2017) reported that phenolic compounds in additives mixture renders antimicrobial activity against several microorganisms. Due to their hydrophobic characteristics they alter the permeability of the cytoplasmic membrane to hydrogen ions and potassium, and deplete the intracellular ATP pool leading to the disruption of essential cellular processes (Costa et al., 2013). These results corroborate well with

the caecal microbial profile discussed later. Stein and Kil (2006) showed that the hydrophobic constituents of essential oils disintegrate the outer membrane of pathogenic bacteria (e.g., *E.coli*).

Phytogenic additives mixture did not affect feed intake; however, the low dose of the additives mixture increased feed conversion, revealing unaffected palatability and enhanced feed efficiency of the diets with the additives. The unchanged feed intake was not expected since essential oils, herbs and spices are often used to improve flavour and, indirectly, palatability of feeds (Zeng et al., 2015). Supplementation of phytogenic additives (essential oils) and herb extracts improved feed conversion ratio (Jamroz et al., 2005). In consonance with the current findings, Celia et al. (2016) observed unaffected feed intake and enhanced feed conversion ratio with the feeding of herbal formulation containing a mixture of essential oils, herbs, spices and extracts. In addition, Hong et al. (2012) reported that the addition of essential oil improved feed efficiency in broilers. In the present experiment, the lower dose of the phytogenic additives mixture was more effective than the higher dose, suggesting that 0.5 ml additives mixture as the optimum level for the young rabbits. It appears that the higher dose additives mixture contained relatively high amount of phenolic compounds which depressed performance relative to the lower dose. Nevertheless, the phenolic compounds concentration was not as high as to induce toxicity or negatively affect the performance of rabbits. This conjecture was confirmed by the superior performance of the HA rabbits relative to the control rabbits. Zeng *et al.* (2015) earlier attributed relatively low performance of animals on high dose of phytogenic additives to the concentration of phenolic compounds.

The LA and HA treatments decreased mortality rate by 60 and 50%, respectively. This effect was expected since the health status of the rabbits supplemented with the additives mixture was affected positively. As discussed later, the additives increased the count of the beneficial *Lactobacillus* spp. with a concurrent decrease in the number of the pathogenic bacterial. The presence of pathogenic bacteria causes digestive disturbances that are often responsible for high morbidity and mortality of young rabbits after weaning and economic losses in rabbit farms (Bovera et al., 2010a; Licois, 2004). The higher mortality of rabbits on high dose of additives mixture relative to the low dose further confirms the efficacy of the low dose. It appears that the high dose increased the concentration of active substances which mildly affected the rabbits but the effect was not pronounced as to raise mortality rate above that of the control rabbits. As previously stated, the low additive dose was more effective than the high dose in enhancing rabbit performance and reducing mortality. Nevertheless, the HA was superior to the control (no additive supplementation). The better gain and feed utilisation efficiency and lower mortality

Table 2. Growth performance, feed intake and feed efficiency of rabbits fed different treatments (Mean± SEM)

		Treatment <sup>1</sup>				P-Values		
	Control	LA	HA	T	W	T×W	L	Q
Initial BW (g)	672±14	669±14	665±14	0.938	-	-	0.723	0.966
Final BW (g)	1840°±25.9	1985 <sup>b</sup> ±23.7	1908 <sup>b</sup> ±23.7	0.001	-	-	0.059	0.001
ADG, $(g/d)$	$33.4^{\circ}\pm0.70$	$37.7^{a}\pm0.68$	35.5 <sup>b</sup> ±0.69	< 0.001	< 0.001	< 0.001	0.039	0.001
FI (g/d)	112.2±1.60	111.3±1.55	109.9±1.57	0.603	< 0.001	< 0.001	0.318	0.918
FCR	$3.42^{a}\pm0.05$	$3.00^{b} \pm 0.051$	$3.38^{a}\pm0.052$	< 0.001	< 0.001	< 0.001	0.639	< 0.001
Mortality (%)	$16.7^{a}\pm0.33$	$6.7^{\circ} \pm 0.21$	$10.0^{b} \pm 0.31$	< 0.001	< 0.001	< 0.001	0.043	0.004

Means in the same row with different superscripts differ, P<0.05; <sup>1</sup>Rabbits were fed either a basal diet without additives (Control) or supplemented with 0.5 ml (LA) or 1 ml (HA) of the additives mixture per litre of drinking water; T, treatment; W, week; T×W, treatment x week interaction; L, linear effect; Q, quadratic effect; ADG, average daily gain; FI, feed intake; FCR, feed conversion ratio

of HA than the control indicate that the concentrations of the phenolic compounds and other active substances in this treatment were below intoxicating levels.

Blood metabolic profiles have been used for diagnosis and prognosis of diseases in animals, and are also useful to assess the welfare condition of animals or to understand if some changes in a diet can affect animal physiology (Olafadehan, 2011). The concentrations of total protein, albumin, globulin, creatinine and uric acid were within the ranges for healthy rabbits (Ajayi and Raji, 2012). Abd-El-Hady (2005) noted unchanged serum total protein, albumin

and globulin in rabbits fed on a diet supplemented with a mixture of natural finely ground herbs and spices enriched with special extracts and essential oils at 300 and 400 g/ton feed. In addition, the non-significant change in concentrations of immunoglobulins of the LA and HA rabbits are indicative unaffected immune defences. The additives mixture did not affect glucose,  $T_3$  and  $T_4$  hormones concentrations; however, Abd-El-Hady (2005) observed that feeding of phytogenic additives mixture increased blood glucose and  $T_3$  hormone concentrations. Glucose concentrations observed in this study were within the range indicated

Table 3. Blood measurements of rabbits fed different doses of additives mixture

	,	Treatment <sup>1</sup>	1	SEM		P-Values	
	Control	LA	HA	-	T	L	Q
Total protein (mg/dl)	6.54	6.85	6.76	0.191	0.493	0.408	0.395
Albumin (mg/dl)	3.59	3.74	3.93	0.149	0.288	0.119	0.875
Globulin (mg/dl)	2.94	3.12	2.83	0.099	0.136	0.426	0.067
Glucose (mg/dl)	128	138	130	10.1	0.749	0.846	0.467
Total lipids (mg/dl)	229	184	182	11.8	0.013	0.008	0.160
Cholesterol (mg/dl)	107 <sup>a</sup>	$88.8^{b}$	$93.7^{b}$	3.31	0.002	0.008	0.008
Triglycerides (mg/dl)	137ª	111 <sup>b</sup>	$117^{\rm b}$	6.5	0.023	0.045	0.048
HDL (mg/dl)	51.9 <sup>b</sup>	65.7a	$57.4^{ab}$	3.7	0.047	0.314	0.023
LDL (mg/dl)	$35.4^{a}$	$30.5^{b}$	29.6 <sup>b</sup>	1.59	0.032	0.015	0.303
AST (U/l)	23.8	19.5	21.1	1.83	0.265	0.303	0.205
ALT (U/l)	35.0	33.1	32.4	2.28	0.706	0.426	0.822
ALP (mg/dl)	97.7	106.4	104.4	4.06	0.304	0.256	0.295
Creatinine (mg/dl)	0.94	0.70	0.75	0.052	0.006	0.012	0.027
Uric acid (mg/dl)	3.43	3.09	3.27	0.317	0.757	0.728	0.514
Calcium (mg/dl)	9.82	11.0	10.6	0.46	0.217	0.219	0.209
Phosphorus (mg/dl)	4.04	4.28	4.11	0.250	0.791	0.843	0.516
Immunoglobulin G (mg/dl)	57.2	61.3	59.7	4.79	0.832	0.710	0.636
Immunoglobulin M (mg/dl)	72.4	78.4	77.7	3.25	0.374	0.261	0.403
Triiodothyronine (T <sub>3</sub> ; ng/ml)	1.22	1.54	1.44	0.105	0.116	0.162	0.118
Thyroxine (T <sub>4</sub> ; ng/ml)	33.1	36.8	35.9	1.51	0.210	0.197	0.222
$T_{4}/T_{3}$	27.8	24.5	25.9	1.31	0.222	0.312	0.157
Malondialdehyde (nmol/ml)	137ª	129 <sup>b</sup>	$131^{ab}$	2.51	0.041	0.087	0.093
TAC (imol/l)	1.49 <sup>b</sup>	$1.89^{a}$	$1.76^{ab}$	0.121	0.049	0.127	0.089

Means in the same row with different superscripts differ, P<0.05; ¹The rabbits were fed either a basal diet without additives (Control) or supplemented with 0.5 ml (LA) or 1 ml (HA) of the additives mixture per litre of drinking water. T, treatment effect; L, linear effect, Q, quadratic effect; HDL, high-density lipoproteins; LDL, low-density lipoproteins; AST, aspartate aminotransferase; ALT, alanine aminotransferase, ALP; alkaline phosphatase; TAC, total antioxidant capacity

for healthy rabbits (Özkan *et al.*, 2012). Liver enzymes (AST, ALT and ALP) are conventionally used for diagnosing domestic animal hepatic damage. The unchanged values suggest that no damage occurred in the liver. The reported concentrations fell within reference ranges for healthy rabbits (Ajayi and Raji, 2012).

Both of LA and HA treatments decreased cholesterol, triglycerides and LDL cholesterol, suggesting beneficial modulatory influence of phytogenic feed additive on cholesterol metabolism and turnover. These effects support the cardiovascular protective influence of the phytogenic feed additives supplementation. The active components and phenolics in the additives have the ability to affect lipid

metabolism in animal tissues by increasing the antioxidative enzymes activity and preventing the production of reactive oxygen species and off-flavours derived from the peroxidation of polyunsaturated fatty acids (Miguel, 2010). Moreover, lowering of the cholesterol and LDL cholesterol concentrations could possibly be associated with cellular cholesterol biosynthesis (Fuhrman *et al.*, 2000) and decrease in intestinal absorption of cholesterol resulting in an increase in faecal excretion of neutral lipids (Purohit and Vyas, 2006). Phytogenics (polyphenolics and flavonoids) have the ability to inhibit the hepatic 3-hydroxy-3-methylglutaryl coenzyme A reductase activity, a key regulatory enzyme in cholesterol synthesis, (Lee *et al.*, 2004). Elshater *et al.* (2009) reported that reduction of

Table 4. Carcass characteristics of rabbits fed different doses of additives mixture

		Treatment <sup>1</sup>	l	SEM		<b>P-Values</b>	
-	Control	LA	HA	<b>-</b> Т	L	Q	
Caecal pH value	6.33	6.29	6.32	0.020	0.469	0.731	0.244
Carcass weight (g)	1073	1122	1085	23.8	0.350	0.733	0.165
Relative carcass weight	57.3	58.5	57.5	0.95	0.668	0.927	0.380
Dressing out percentage	63.4	64.8	64.1	1.03	0.631	0.639	0.409
Liver percentage	3.88	4.19	4.14	0.242	0.638	0.468	0.551
Heart percentage	0.33	0.35	0.41	0.025	0.091	0.038	0.498
Lung percentage	0.72	0.70	0.76	0.055	0.680	0.567	0.512
Spleen percentage	0.12	0.10	0.13	0.013	0.510	0.624	0.298
Pancreas percentage	1.03	1.04	1.21	0.059	0.081	0.047	0.280
Kidney percentage	1.03	1.04	1.21	0.059	0.081	0.047	0.280
Thymus gland percentage	0.10	0.10	0.08	0.009	0.337	0.416	0.220
Thyroid gland percentage	0.009	0.008	0.008	0.0006	0.994	0.921	0.973
Adrenal gland percentage	0.008	0.009	0.009	0.0010	0.554	0.415	0.481
Abdominal fat percentage	0.90	0.75	0.74	0.099	0.457	0.276	0.550
Shoulder fat percentage	0.31	0.28	0.25	0.048	0.683	0.391	0.955
Full stomach percentage	4.89	4.67	4.80	0.260	0.837	0.807	0.593
Full intestine percentage	7.29	8.03	7.63	0.331	0.316	0.470	0.183
Full cecum percentage	5.88	6.76	6.68	0.509	0.418	0.281	0.450
Intestine length (Cm)	379	386	388	6.2	0.601	0.348	0.739
Cecum length (Cm)	46.8	48.0	48.7	1.32	0.619	0.341	0.879
Carcass length (Cm)	33.7	33.5	32.8	0.55	0.518	0.276	0.762

Means in the same row with different superscripts differ, P<0.05; T, treatment effect; L, linear effect, Q, quadratic effect;  $^1$ The rabbits were fed either a basal diet without additives (Control) or supplemented with  $0.5 \, \text{ml}$  (LA) or  $^1$  ml (HA) of the additives mixture per litre of drinking water

cellular cholesterol biosynthesis is associated with increased activity of the LDL receptor, leading to enhanced removal of LDL from blood and thus causing a decreased serum or plasma cholesterol concentration.

The LA treatment increased HDL cholesterol and TAC, and decreased MDA because the additives mixture contains phenolic compounds that elicit antioxidant action by scavenging reactive oxygen species, enhancing the cellular antioxidant enzyme (e.g., superoxide dismutase, catalase and glutathione peroxidase) and increasing glutathione in the cells (Borek, 2001). The MDA is an indicator of cell membrane injury and its concentration depends on lipid peroxidation (El-Gogary et al., 2018). Increasing HDL cholesterol reveals reduced atherogenic risk by virtue of increased reverse cholesterol transport from peripheral organs to liver (Hermansen et al., 2003). The antioxidative activity of some phenolics components in feed additives mixture has been attributed to scavenging of superoxide anion and hydroxyl radicals (Elkirdasy et al., 2015). Although the phytogenic additives mixture altered some blood metabolic profiles, the occurrence of all the blood parameters within the normal physiological ranges ruled out the possibility of any intoxication, especially with the high dose. Therefore, the results indicate the safety of the additives mixture doses used in the current study.

The weak effects of additives on carcass weight, relative organs weights, cecum length, carcass length are in agreement with previous studies in which feeding of phytogenic additives did not affect carcass characteristics and relative organ weights (Cho *et al.*,

2014; Hong *et al.*, 2012; Kirkpinar *et al.*, 2011) or meat pH value (Cho *et al.*, 2014). Abdel-Wareth *et al.* (2018) observed that supplementation of rabbits with thyme essential oil did not affect the percentage of head, liver, heart, lungs and kidneys of the chilled carcass as well as the percentage of fore, mid and hind parts.

The LA and HA treatments increased the beneficial (Lactobacillus spp.) bacteria counts by about 12 and 6%, respectively. It is well documented that essential oil, herbs and phytogenic extracts suppress harmful microorganisms and stimulate beneficial microbes such as *Lactobacillus* spp. (Simitzis, 2017). Lactobacillus spp. is a normal member of the rabbit intestinal microflora, with a positive effect on regular gut function (Carabaño et al., 2010; Celia et al., 2016). Therefore, increasing the number of *Lactobacillus* spp. could play a role in preventing the mortality of the rabbits. Lactobacilli bacteria activate the intestinal immune system and increase the resistance to diseases through the release of low-molecular weight peptides which induce immune activation (Muir et al., 2000). Moreover, increasing the count of Lactobacilli bacteria contributes to the colonization resistance against pathogenic microbes by modifying the receptors used by them (Adil and Magray, 2012; Rinttila and Apajalahti, 2013).

As compared to control, the LA and HA treatments decreased coliform bacteria and *Escherichia coli* counts by about 49 and 33%, and 28 and 23%, respectively. Placha *et al.* (2013) observed decreased number of coliforms in the caecum of rabbits fed a diet supplemented with 0.5 g/kg DM of thyme essential oil. Celia *et al.* (2016) observed reduced

Table 5. Caecal bacteria of rabbits (log<sub>10</sub> cfu/g) fed different doses of additives mixture

	ŗ	Treatment <sup>1</sup>	I	SEM		P-Values	
	Control	LA	HA	T	L	Q	
Lactobacillus spp.	7.80°	8.74 <sup>a</sup>	8.25 <sup>b</sup>	0.067	< 0.001	< 0.001	< 0.001
Coliform bacteria count	$7.54^{a}$	$3.88^{c}$	$5.02^{b}$	0.099	< 0.001	< 0.001	< 0.001
Escherichia coli	6.41a	4.61°	4.91 <sup>b</sup>	0.082	< 0.001	< 0.001	< 0.001

Means in the same row with different superscripts differ, P<0.05; T, treatment effect; L, linear effect, Q, quadratic effect;  $^{1}$ The rabbits were fed either a basal diet without additives (Control) or supplemented with 0.5 ml (LA) or 1 ml (HA) of the additives mixture per litre of drinking water

bacterial diversity in the caecum of rabbits fed a diet supplemented with herbal formulation containing a mixture of essential oils, herbs, spices and extracts. A reduction in the number of pathogenic bacteria changes the microbial ecology in favour of beneficial species in the intestine (Michiels et al., 2009), resulting in improved ability of epithelial cells to regenerate villus and thus enhances intestinal absorptive capacity (Mourão et al., 2006). Such effects reveal the antimicrobial activity of additives mixture to control pathogenic bacteria. Phenolic compounds in essential oils, herbs and extracts display antimicrobial action against Escherichia coli (Jang et al., 2007) and many other pathogens, and prevent their adhesion, colonization and proliferation in the gut of broilers (Simitzis, 2017). Essential oils, phytogenic extracts and aromatic plants are well known to exert antibacterial, antifungal and antiviral activity. The pathogenic bacteria are normal inhabitants of the intestinal tract of rabbits and can inflict digestive disturbances that are often responsible for high morbidity and mortality of young rabbits after weaning, causing economic losses in rabbit farms.

Decreasing pathogens such as E. coli contributes to healthy microbial metabolites, improves intestinal integrity and ensures protection against enteric disease (Baker et al., 2012; Oviedo-Rondón et al., 2006; Placha et al., 2013; Tiihonen et al., 2010). Phenolic compounds of essential oils, extracts and herbal plants cause structural and functional damage to cytoplasmic membranes of harmful bacteria (Soultos et al., 2009). Bozin et al. (2006) showed the in vitro antibacterial activity of carvacol and thymol (the main components of oregano essential oil) on multi-resistant strains of E. coli. The mechanisms of reduction of such pathogenic bacteria in the intestinal tract after administration of phytogenic additives have not been rigorously studied up till date. However, the hypothesis that plant extracts/essential oils inhibit target cells in the membrane and deplete the transmembrane potential and/ or the pH gradient which result in the leakage of cellular materials and destruction of bacterial cells has been confirmed (Cleveland et al., 2001; Szabóová et al., 2012).

#### **CONCLUSIONS**

The results of the present experiment demonstrate that supplementation of phytogenic additive mixture enhanced feed conversion, daily gain and positively altered caecal bacterial profile of rabbit. Better response was obtained at lower dose (0.5 ml/l drinking water) as compared compared to dose (1 ml/l drinking water) of supplementation. Thus, it was concluded that supplementation of phytogenic feed additive at 1 ml/l drinking water would be beneficial to improve performance and gut health of rabbits. Additional studies, involving *in vitro* and *in vivo* evaluations, are recommended to investigate different levels of the additives mixture on the performance of rabbits at different stages of growth.

#### REFERENCES

- Abd-El-Hady, A.M. 2005. Performance, physiological parameters and slaughter characteristics in growing rabbits as affected by a herbal feed additives (Digestarom®). *J. Int. Sci. Publ. Agric. Food* 2: 353-365.
- Abdel-Wareth, A.A.A., Taha, E.M.M., Südekum, K.-H.H. and Lohakare, J. 2018. Thyme oil inclusion levels in a rabbit ration: Evaluation of productive performance, carcass criteria and meat quality under hot environmental conditions. *Anim. Nutr.* 4: 410-416.
- Adil, S. and Magray, S.N. 2012. Impact and manipulation of gut microflora in poultry: A review. *J. Anim. Vet. Adv.* 11:873-877.
- Ajayi, A.F. and Raji, Y. 2012. Haematological and serum biochemical indices of pre-pubertal male rabbits fed with graded level of blood-wild sunflower forage meal mixture. *African J. Biotechnol*. 11: 8730-8734.
- Baker, J., Brown, K., Rajendiran, E., Yip, A., DeCoffe, D., Dai,
  C., Molcan, E., Chittick, S.A., Ghosh, S., Mahmoud,
  S.and Gibson, D.L. 2012. Medicinal lavender modulates the enteric microbiota to protect against
  Citrobacter rodentium-induced colitis. AJP Gastrointest. Liver Physiol. 303: 825-836.
- Barnes, E.M. and Impey, C.S. 1970. The isolation and properties of the predominant anaerobic bacteria in the caeca of chickens and turkeys. *Br. J. Poult. Sci.* 11: 467-481.
- Benlemlih, M., Aarab, A., Bakkali, M., Arakrak, A. and Laglaoui, A. 2014. Effect of dietary fennel and thyme essential oil supplementation on zootechnical parameters and

- caecal microflora of growing rabbit. *Rev. Microbiol. Ind. San. Environ.* 8: 16-25.
- Blasco, A.and Ouhayoun, J. 1993. Harmonization of criteria and terminology in rabbit meat research. Revised proposal. *World Rabbit Sci.* 4: 93-99.
- Borek, C. 2001. Antioxidant health effects of aged garlic extract. *J. Nutr.* 131:1010-1015.
- Bovera, F., Marono, S., Di Meo, C., Piccolo, G., Iannaccone, F. and Nizza, A. 2010a. Effect of mannanoligosaccharides supplementation on caecal microbial activity of rabbits. *Animal*, 4: 1522-1527.
- Bovera, F., Nizza, S., Marono, S., Mallardo, K., Piccolo, G., Tudisco, R., De Martino, L.and Nizza, A. 2010b. Effect of mannan oligosaccharides on rabbit performance, digestibility and rectal bacterial anaerobic populations during an episode of epizootic rabbit enteropathy. *World Rabbit Sci.* 18: 9-16.
- Bozin, B., Mimica-Dukic, N., Simin, N., and Anackov, G. 2006. Characterization of the volatile composition of essential oils of some Lamiaceae spices and the antimicrobial and antioxidant activities of the entire oils. *J. Agric. Food Chem.* 54:1822-1828.
- Carabaño, R., Piquer, J., Menoyo, D.and Badiola, I. 2010. The digestive system of the rabbit, in: de Blas, C., Wiseman, J., Allain, D., Blas, E., Carabano, R. (Eds.), Nutrition of the Rabbit. CABI Publishing, Wallingford/; Cambridge, MA, pp. 9-27.
- Celia, C., Cullere, M., Gerencsér, Z.S., Matics, Z.S., Giaccone, V., KOvács, M., Bónai, A., Szendro, Z.S. and Dalle Zotte, A. 2016. Dietary supplementation of digestarom® herbal formulation: Effect on apparent digestibility, faecal and caecal microbial counts and live performance of growing rabbits. *World Rabbit Sci.* 24: 95-105.
- Cervantes-Valencia, M.E., Alcala-Canto, Y., Salem, A.Z.M., Kholif, A.E., Ducoing-Watty, A.M., Bernad-Bernad, M.J. and Gutiérrez-Olvera, C. 2015. Influence of curcumin (Curcuma longa) as a natural anticoccidial alternative in adult rabbits: First results. *Ital. J. Anim. Sci.* 14. https://doi.org/10.4081/ijas.2015.3838
- Cheeke, P.R. 1987. *Rabbit Feeding and Nutrition*. Academic Press Inc. Ed., Orlando, USA.
- Cho, J.H., Kim, H.J.and Kim, I.H. 2014. Effects of phytogenic feed additive on growth performance, digestibility, blood metabolites, intestinal microbiota, meat color and relative organ weight after oral challenge with clostridium perfringens in broilers. *Livest. Sci.* 160: 82-88.
- Cleveland, J., Montville, T.J., Nes, I.F. and Chikindas, M.L. 2001. Bacteriocins: safe, natural antimicrobials for food

- preservation. Int. J. Food Microbiol. 71: 1-20.
- Combes, S., 2004. Valeur nutritive de la viande de lapin. INRA Prod. Animal. 17: 373-383.
- Costa, L.B., Luciano, F.B., Miyada, V.S. and Gois, F.D. 2013. Herbal extract and organic acids as natural feed additives in pig diets. *S. Afr. J. Anim. Sci.* 43: 181-193.
- Dalle Zotte, A. 2002. Perception of rabbit meat quality and major factors influencing the rabbit carcass and meat quality. *Livest. Prod. Sci.* 75: 11-32.
- El-Gogary, M.R., El-Said, E.A. and Mansour, A.M. 2018. Physiological and Immunological Effects of Rosemary Essential Oil in Growing Rabbit Diets. *J. Agric. Sci.* 10:485. https://doi.org/10.5539/jas.v10n7p485
- Elkirdasy, A., Shousha, S., Alrohaimi, A.H. and Arshad, M.F. 2015. Hematological and immunobiochemical study of green tea and ginger extracts in experimentally induced diabetic rabbits. *Acta Pol. Pharm. Drug Res.* 72: 497-506.
- Elshater, A.E., Muhammad, M.A.S. and Mahrous, M.A.M. 2009. Effect of ginger extracts consumption on levels of blood glucose, lipid profile and kidney functions in alloxane induced-diabetic rats. *Egypt. Acad. J. Biol. Sci.* 2: 153-162.
- Fortier, M.P., Saucier, L. and Guay, F. 2012. Effects on microbial quality of fresh pork loin during storage from oregano oil and cranberry pulp diet supplementation in pigs. *Can. J. Anim. Sci.* 92: 465-471.
- Fuhrman, B., Rosenblat, M., Hayek, T., Coleman, R. and Aviram, M. 2000. Ginger extract consumption reduces plasma cholesterol, inhibits LDL oxidation and attenuates development of atherosclerosis in atherosclerotic, apolipoprotein E-deficient mice. *J. Nutr.* 130: 1124-1131.
- Gado, H.M., Kholif, A.E., Salem, A.Z.M., Elghandour, M.M.Y., Olafadehan, O.A., Martinez, M.A. and Al-Momani, A.Q. 2016. Fertility, mortality, milk output, and body thermoregulation of growing Hy-Plus rabbits fed on diets supplemented with multi-enzymes preparation. Trop. Anim. Health Prod. 48: 1375-1380.
- Hafsa, A.S.H., Salem, A.Z.M., Hassan, A.A., Kholif, A.E., Elghandour, M.M.Y., Barbabosa, A., Lopez, S., Abu Hafsa, S.H., Salem, A.Z.M., Hassan, A.A., Kholif, A.E., Elghandour, M.M.Y., Barbabosa, A. and Lopez, S. 2016. Digestion, growth performance and caecal fermentation in growing rabbits fed diets containing foliage of browse trees. *World Rabbit Sci.* 24:283. https://doi.org/10.4995/wrs.2016.4359
- Hermansen, K., Dinesen, B., Hoie, L.H., Morgenstern, E. and Gruenwald, J. 2003. Effects of soy and other natural products on LDL: HDL ratio and other lipid

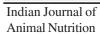
- parameters: a literature review. Adv. Ther. 20: 50-78.
- Hong, J.C., Steiner, T., Aufy, A. and Lien, T.F. 2012. Effects of supplemental essential oil on growth performance, lipid metabolites and immunity, intestinal characteristics, microbiota and carcass traits in broilers. *Livest. Sci.* 144:253-262.
- Jamroz, D., Wiliczkiewicz, A., Wertelecki, T., Orda, J. and Skorupiñska, J. 2005. Use of active substances of plant origin in chicken diets based on maize and locally grown cereals. *Br. Poult. Sci.* 46: 485-493.
- Jang, I.S., Ko, Y.H., Kang, S.Y and Lee, C.Y. 2007. Effect of a commercial essential oil on growth performance, digestive. Anim. Feed Sci. Technol. 134: 304-315.
- Jang, I.S., Ko, Y.H., Yang, H.Y., Ha, J.S., Kim, J.Y., Kim, J.Y., Kang, S.Y., Yoo, D.H., Nam, D.S., Kim, D.H. and Lee, C.Y. 2004. Influence of essential oil components on growth performance and the functional activity of the pancreas and small intestine in broiler chickens. *Asian-Australas. J. Anim. Sci.* 17: 394-400.
- Kholif, A.E., Abdo, M.M., Anele, U.Y., El-Sayed, M.M. and Morsy, T.A. 2017. Saccharomyces cerevisiae does not work synergistically with exogenous enzymes to enhance feed utilization, ruminal fermentation and lactational performance of Nubian goats. *Livest. Sci.* 206:17-23.
- Kirkpinar, F., Unlu, H.B. and Ozdemir, G. 2011. Effects of oregano and garlic essential oils on performance, carcass, organ and blood characteristics and intestinal microflora of broilers. *Livest. Sci.* 17: 219-225.
- Koné, A.P., Cinq-Mars, D., Desjardins, Y., Guay, F., Gosselin, A. and Saucier, L. 2016. Effects of plant extracts and essential oils as feed supplements on quality and microbial traits of rabbit meat. World Rabbit Sci. 24: 107-119.
- Lambert, R.J.W., Skandamis, P.N., Coote, P.J. and Nychas, G.J.E. 2001. A study of the minimum inhibitory concentration and mode of action of oregano essential oil, thymol and carvacrol. *J. Appl. Microbiol*. 91:453-462.
- Lee, K.W., Everts, H.and Beynen, A.C. 2004. Essential Oils in Broiler *Nutrition*. *Int. J. Poult. Sci.* 3, 738-752.
- Licois, D. 2004. Domestic rabbit enteropathies, In: The Eighth World Rabbit Congress. Puebla, Mexico, pp. 385-403.
- Michiels, J., Missotten, J.A.M., Fremaut, D., De Smet, S.and Dierick, N.A. 2009. *In vitro* characterisation of the antimicrobial activity of selected essential oil components and binary combinations against the pig gut flora. *Anim. Feed Sci. Technol.* 151: 111-127.
- Miguel, M.G. 2010. Antioxidant and anti-inflammatory activities of essential oils: a short review. Molecules,

- 15:9252-9287.
- Morsy, T.A., Kholif, A.E., Matloup, O.H., Abu Elella, A., Anele, U.Y., Caton, J.S., Elella, A.A., Anele, U.Y. and Caton, J.S. 2018. Mustard and cumin seeds improve feed utilisation, milk production and milk fatty acids of Damascus goats. *J. Dairy Res.* 85: 1-10.
- Mourão, J.L., Pinheiro, V., Alves, A., Guedes, C.M., Pinto, L., Saavedra, M.J., Spring, P. and Kocher, A. 2006. Effect of mannan oligosaccharides on the performance, intestinal morphology and cecal fermentation of fattening rabbits. *Anim. Feed Sci. Technol.* 126: 107-120.
- Muhl, A. and Liebert, F. 2007. No impact of a phytogenic feed additive on digestion and unspecific immune reaction in piglets. *J. Anim. Physiol. Anim. Nutr. (Berl)*. 91: 426-431.
- Muir, W.I., Bryden, W.L. and Husband, A.J. 2000. Immunity, vaccination and the avian intestinal tract. *Dev. Comp. Immunol*. 24: 325-342.
- Olafadehan, O.A. 2011. Changes in haematological and biochemical diagnostic parameters of Red Sokoto goats fed tannin-rich Pterocarpus erinaceus forage diets. *Vet. Arch.* 81: 471-483.
- Oviedo-Rondón, E.O., Hume, M.E., Hernández, C. and Clemente-Hernández, S. 2006. Intestinal microbial ecology of broilers vaccinated and challenged with mixed Eimeria species, and supplemented with essential oil blends. *Poult. Sci.* 85: 854-860.
- Özkan, C., Kaya, A. and Akgül, Y. 2012. Normal values of haematological and some biochemical parameters in serum and urine of New Zealand white rabbits. *World Rabbit Sci.* 20: 253-259.
- Placha, I., Chrastinova, L., Laukova, A., Cobanova, K., Takacova, J., Strompfova, V., Chrenkova, M., Formelova, Z. and Faix, S. 2013. Effect of thyme oil on small intestine integrity and antioxidant status, phagocytic activity and gastrointestinal microbiota in rabbits. *Acta Vet. Hung*. 61: 197-208.
- Platel, K.and Srinivasan, K. 2000. Stimulatory influence of select spices on bile secretion in rats. *Nutr. Res.* 20: 1493-1503.
- Platel, K. and Srinivasan, K. 1996. Influence of dietary spices or their active principles on digestive enzymes of SI mucosa in rats. Wiley Online Libr. 44: 55-59.
- Purohit, A. and Vyas, K.B. 2006. Antiatherosclerotic effect of Capparis decidua fruit extract in cholesterol-fed rabbits. *Pharm. Biol.* 44: 172-177.
- Rinttila, T. and Apajalahti, J. 2013. Intestinal microbiota and metabolites—Implications for broiler chicken health and performance. *J. Appl. Poult. Res.* 22: 647-658.

- Sarac, N., Ugur, A., Duru, M.E. and Varol, O. 2007. Antimicrobial activity, antioxidant activity and chemical composition of Origanum onites L. and Origanum vulgare L. ssp. hirtum (Link) ietswaart from Mugla (Turkey). Int. Med. Aromat. Plants Conf. Culin. Herbs 826: 397-404.
- Schneider, B.H. and Flatt, W.P., 1975. *The Evaluation of Feed through Digestibility Experiments*. University of Georgia Press, Athens, Georgia, USA.
- Simitzis, P.E. 2017. Enrichment of animal diets with essential oils—a great perspective on improving animal performance and quality characteristics of the derived products. Medicines, 4:35. https://doi.org/10.3390/medicines4020035
- Soultos, N., Tzikas, Z., Christaki, E., Papageorgiou, K. and Steris, V. 2009. The effect of dietary oregano essential oil on microbial growth of rabbit carcasses during refrigerated storage. *Meat Sci.* 81: 474-478.

- Stein, H.H. and Kil, D.Y. 2006. Reduced use of antibiotic growth promoters in diets fed to weanling pigs: Dietary tools, part 2. *Anim. Biotechnol.* 17: 217-231.
- Szabóová, R., Lauková, A., Chrastinová, L., Strompfová, V., Simonová, M.P., Plachá, I., Vasilková, Z., Chrenková, M. and Faix, Š., 2012. Benefficial effect of plant extracts in rabbit husbandry. *Acta Vet. Brno* 81:245-250.
- Tiihonen, K., Kettunen, H., Bento, M.H.L., Saarinen, M., Lahtinen, S., Ouwehand, A.C., Schulze, H. and Rautonen, N. 2010. The effect of feeding essential oils on broiler performance and gut microbiota. *Br. Poult. Sci.* 51: 381-392.
- Zeng, Z., Zhang, S., Wang, H.and Piao, X. 2015. Essential oil and aromatic plants as feed additives in non-ruminant nutrition: A review. *J. Anim. Sci. Biotechnol.* 6: 1-10.

Received on 03-12-2019 and accepted on 29-12-2019





Indian J. Anim. Nutr. 2019. 36 (4): 377-381 doi: 10.5958/2231-6744.2019.00061.6

# A Nursery Feed for Indian Major Carps (*Labeo rohita*) and its Evaluation in Farmer's Field

Krushna Chandra Das\*, Kedar Nath Mohanta, Santosh Kumar Nayak, Snehalata Mohanty, Prabin Kumar Sahoo and Priyabrat Swain

Fish Nutrition and Physiology Division,

ICAR- Central Institute of Freshwater Aquaculture, Kausalyaganga, Bhubaneswar 751 002, India

#### **ABSTRACT**

Realizing the importance of carp feed for its nursery phase, the Fish Nutrition and Physiology Division of ICAR- Central Institute of Freshwater Aquaculture (CIFA), Bhubaneswar has been working since last three years and has developed a nursery feed for carp which ensured better survival, growth and palatability. The feed was prepared from maize, soybean meal, rice bran, groundnut oil cake, fish meal, oil, minerals and vitamin mixture by extrusion technology and then crumbled into suitable size for feeding the carp fry. The prepared feed was evaluated through demonstration experiments in different locations of Odisha in Government fish farm and farmer's ponds following standard protocols. The evaluation was done in terms of growth, feed efficiency and survivability with existing feed commonly used by the farmers of the region. Result of this experiment showed that, the fish fed with developed nursery feed had 1.5 to 2 times higher growth performance, 25% higher survivability and better feed efficiency compared to the feed which is used by most of the farmers of this region.

Keywords: Carps, Extrusion, FCR, Growth performance, Nursery feed

# **INTRODUCTION**

Nursery phase is one of most critical phase in the production of stockable seed materials in carp culture. Generally, it is observed that the recovery of spawn to fry and fry to fingerling is around 25-30% and 40-50%, respectively in farmers' practice. Nutrition plays a vital role in growth and survival of the carp in nursery rearing. Non-availability of quality feed is one of the major reasons for the poor survival and growth making the nursery rearing of carp less remunerative. Realizing the importance of carp feed for its nursery phase, the Fish Nutrition and Physiology Division of ICAR-CIFA, Bhubaneswar has been working since last three years and has developed a nursery feed (Carp starter) for carp which ensures better survival and growth.

Extrusion technology is gaining importance for production of fish feed for Indian Major Carp. Extrusion process improves nutrient digestibility, palatability, pellet durability, water stability and pellet storage life (Barrows and Hardy, 2000). It also increases *in vivo* digestibility of dry matter (DM) and energy of feed in rainbow trout *Oncorhynchus mykiss* (Cheng and Hardy, 2003). Good quality feed containing til oil

cake could be produced through extrusion technology by maintaining extrusion temp of 130°C and moisture of 20% (Das et al., 2018). The nursery feed for carp (Carp starter) was produced from maize, soybean meal, rice bran, groundnut oil cake, fish meal, oil, minerals and vitamin mixture by extrusion and then crumbled into suitable size for feeding the fry. This feed is nutritious and highly palatable. It has been tested at indoor facilities and demonstrated multi-locationally in Government fish farm and farmer's pond. In this experiment, Carp starter produced through extrusion technology was evaluated in terms of growth, feed efficiency and survivability with existing feed commonly used by the farmers of the region.

# MATERIALS AND METHODS

A fish feed for carp fry was formulated using high quality locally available feed ingredients *i.e.* maize, soyabean meal, groundnut oil cake (GNOC), fish meal and de-oiled rice bran (DORB) with supplementation of minerals and vitamin mixture. The ingredients were pulverised, mixed, extruded and then dried. The extruded and dried feed was then ground into suitable size, packaged and stored for feeding the fish (Fig. 1). The

 $^*Corresponding \ author \ Email: kcdasicar@gmail.com$ 





Fig. 1. Carp Starter after preparation for carp fry

feeds were produced in the feed mill facilities of ICAR-CIFA, Bhubaneswar, India with extrusion temperatures of 130°C and moisture of 20% maintaining constant pressure (10 kg/cm²). After production of extruded feed, it was packaged for easy transportation to the experimental site. The feed was then analysed for chemical characteristics to ascertain the quality of the feed (Table 1). Developed nursery feed was then compared with a feed (control feed) which is normally used by most of the farmers of this region. A control feed was also prepared after mixing GNOC and rice bran in 1: 1 ratio and then grinded properly into powder form as it is the standard practices of feeding carp fry in many places of India including Odisha.

The chemical parameters *i.e.* DM, crude protein (CP), ether extract (EE), crude fibre (CF), nitrogen free extract (NFE) and total ash were analysed for quality evaluation (AOAC, 2012). DM was estimated by oven drying the samples at 105°C till a constant weight and CP content was calculated by estimating nitrogen content by micro-Kjeldahl method and multiplying with a factor 6.25. EE was determined by solvent extraction with petroleum ether, boiling point 40–60°C, for 10–12 h. Total ash content was determined by incinerating the sample at 650 °C for 6 h and CF was determined by acid digestion (1.25%) followed by alkali digestion (1.25%).

Demonstration experiment was simultaneously conducted in five location of Odisha *i.e.* state fishery farm, Keonjhar; College of Fisheries, Rangailunda (OUAT), Berhampur; KVK, OUAT, Ranital, Bhadrak; ICAR-CIFA, Bhubaneswar and Sahu Fish Farm, Subarnapur, Gop, Puri.

Pond preparation was done in all the five places for demonstration experiment. Before stocking the fish, all the ponds were dried and bleaching powder (30%)

Table 1. Chemical characteristics of nursery feed (Carp Starter)

Parameters	% on DM basis
Moisture	7.5
Crude Protein (CP)	35.58
Crude Fibre (CF)	4.62
Total Ash	10.74
Ether Extract (EE)	6.10
Nitrogen Free Extract (NFE)	42.96
Energy (kcal/g)	3.61

active chlorine) was applied at 300 kg ha<sup>-1</sup> to eradicate the unwanted predatory and weed fishes. Lime was applied at 1000 kg ha<sup>-1</sup>. After 7 days of lime application, the water was filled up to 1.0 m depth and ponds were fertilized with raw cow dung, urea and bleaching powder at 3.0 ton ha<sup>-1</sup>, 40.0 kg ha<sup>-1</sup> and 60.0 kg ha<sup>-1</sup>, respectively (ICAR 2009).

Rohu fry (average weight 0.5 gm) were either procured from CIFA Farm, Bhubaneswar, India or purchased from the local market for stocking in the respective pond sites. Carp fry were stocked randomly into triplicate ponds (each 130×60 ft) for both control and treatment following a completely randomized design and were stocked @ 1 lakh/ acre. The fish of control ponds were fed with GNOC and rice bran (1:1 ratio) as it is the standard practices of rearing fry in many places of India including Odisha. The fish of treatment ponds were fed with carp starter to compare the performance with fish of control ponds. Both



Fig. 2. Rohu fry after feeding with Carp Starter for one month

treatment and control fish were fed at the rate of 10% of body weight (BW) for 2 weeks followed by 5 % of BW for another 2 weeks. Sampling of fish was done in fortnightly interval to regulate the feeding requirement and to estimate the average BW and biomass of fish in each pond (Fig. 2). The experiment was continued for one month.

The data of the experiment were statistically analyzed by using computer software (GraphPad PRISM 2007).

# RESULTS AND DISCUSSION

The chemical characteristics of nursery feed (Carp Starter) used in the experiment is presented in Table 1. The feed contained 35.58% protein, 6.10% fat and 4.62 % CF and size of feed particles varied from 0.4 to 0.5 mm. The chemical characteristics showed that, it is a very good feed for carp fry.

The growth performance of *L. rohita* fry fed with different test diets in different locations of Odisha like Keonjhar, Bhadrak, Bhubaneswar, Berhampore and Gop

Table 2. Growth performance of L. rohita fry fed with different diets in Odisha

Locality	Parameters	Control	Carp Starter	P value	SEM
Keonjhar	Initial weight (g)	0.30	0.30	-	-
	Final weight (g)	3.36ª	4.72 b	0.0001	0.3069
	Weight gain (g)	3.06 a	4.42 b	0.0001	0.3069
	SGR (% day-1)	8.77 a	12.27 b	< 0.0001	0.7877
	FCR (Apparent)	2.46 a	1.26 b	0.0002	0.2717
	Survival (%)	63 a	87 <sup>b</sup>	0.0114	5.888
Bhadrak	Initial weight (g/fish)	0.75	0.75	-	-
	Final weight (g/fish)	2.53 a	6.00 b	0.0042	0.8188
	Weight gain (g/fish)	1.78 a	5.25 b	0.0042	0.8188
	Survival (%)	53.0 a	82.0 b	0.0056	6.913
	FCR (Apparent)	2.10 a	1.69 b	0.0371	0.1094
Bhubaneswar	Initial weight (g/fish)	0.93	0.93	-	-
	Final weight (g/fish)	5.07 a	6.12 b	0.0135	0.2598
	Weight gain (g/fish)	4.14 a	5.19 <sup>b</sup>	0.0135	0.2598
	SGR (% day -1)	4.8 a	5.3 b	0.0376	0.1335
	Survival (%)	80°	95 <sup>b</sup>	0.0086	2.422
	FCR (Apparent)	1.57 a	1.24 b	0.0014	0.076
Berhampore	Initial weight (g/fish)	0.7	0.7	-	-
	Final weight (g/fish)	1.45 a	3.68 b	0.0001	0.5008
	Weight gain (g/fish)	0.75 a	2.98 b	0.0001	0.5008
	SGR (% day -1)	2.43	5.53	0.0376	0.1335
	Survival (%)	75 a	94 b	0.0004	4.394
	FCR (Apparent)	3.43 a	1.5 b	0.0001	0.4349
Gop	Initial weight (g/fish)	0.5	0.5	-	-
	Final weight (g/fish)	3.37 a	5.60 <sup>b</sup>	0.0001	0.5016
	Weight gain (g/fish)	2.87 a	5.10 <sup>b</sup>	0.0001	0.5016
	SGR (% day <sup>-1</sup> )	6.36	8.05	0.0376	0.1335
	Survival (%)	71ª	95 <sup>b</sup>	0.0001	5.241
	FCR (Apparent)	2.45 a	1.5 b	0.0004	0.2045

Means with different superscripts in a row differ significantly (P<0.01).

is presented in table 2. The initial average weight of fry was 0.63 g (varied from 0.3 g to 0.93 g) in both control and treatment groups and attained 3.15 g (varied from 1.45 g to 5.07 g) in control group and 5.22 g (varied from 3.68 g to 6.12 g) in treatment groups after 30 days of feeding trial. The weight gain in all the feeding experiments were significantly higher in treatment groups compared to control groups. The specific growth ratio (SGR) were also significantly higher in treatment group compared to control group of fish and is indicated in table. Similarly, feed conversion ratio (FCR) of fish fed on carp starter was significantly superior (P<0.05) compared to fish fed on control feed. The growth performance of fish during the experiment showed that the growth rate of fish in Keonjhar, Bhadrak, Bhubaneswar, Berhampore and Gop was higher than control group of fish fed with ground nut oilcake and rice bran. There was an average increase of 1.5 to 2 times of weight gain fed with carp starter compared to control feed.

Nursery feed (carp starter) was developed by extrusion process in the feed mill of ICAR-CIFA, higher feed utilization by extrusion process may be responsible for improved growth and survival of carp fry (Das et al., 2018). Extrusion is the process in which the feed material is pushed through the barrel by means of screws of different configurations and pressed through the die at the end of barrel. The basic concept of extrusion process is high temperature, short time, whereby the high temperature is a direct result of friction or pre-conditioning and steam injection or a combination of both (Levic, 2010). Extrusion technology has been used in the feed industry for almost one century (Hardy and Barrows, 2000) and it has become popular for aqua feed production (FMT, 2005) as extruded pelleted feed reduces the feed wastage and improve the over all growth performane of fish.

The survival percentage of fish fed on carp starter was higher compared to fish fed on control feed. The survival percentage in control group of fish was 68 percentage (53 -80 percent) where as survivability in treatment group of fish was 90.6 percent (82 - 95

percent). Survivability of carp fry is an important parameters as generally, it is observed in the farmers practice that recovery of spawn to fry and fry to fingerling is around 25-30% and 40-50%, respectively. Nutrition plays a vital role on growth and survival in nursery rearing of the carp. Non-availability of quality feed is one of the reasons for the poor survival and growth making the nursery rearing of carp less remunerative. Though planktons are available in natural rearing process, it is not sufficient to meet nutritional requirement of all nursery carps. In our experiment, survivability of fish in different experimental stations varied from 82 to 95 percent with an average of 90.6 percent. Availability of nutrition rich carp starter along with natural planktons are prime factors for improving survival of carp fry.

Carp starter as used in this experiment contained many feed ingredients like maize, soyabean meal, groundnut oil cake (GNOC), fish meal and de-oiled rice bran (DORB). Supplementation of many feed ingredients in the production of fish feed improve the keeping quality of feed compared to two or three feed ingredients as has been observed by Das et al. (2016). There may be balancing of amino acids like methionine, lysine and arginine in carp starter produced from soyabean meal, ground nut cake and fish meal compared to feed of control group. This might be due to better utilization of protein in Carp starter compared to control feed. It has been reported that the feed prepared by using more than one protein sources always resulted in better growth of fish due to proper balancing of amino acids (Djissou et al., 2016; Gaylord et al., 2017). This showed that, carp starter had better growth performance as compared to control feed. The palatability of feed was higher in carp starter compared to control feed as observed during feeding time. Use of fish meal/fish oil in carp starter might have increased palatability of feed as observed by many workers. This indicated that, carp starter is a very high quality feed and may be provided to carp fry for higher growth performance and survivability.

#### **CONCLUSION**

Result of this experiment showed that, the fish fed with developed nursery feed had 1.5 to 2 times higher growth performance, 25% higher survivability and better feed efficiency compared to the feed which is used by most of the farmers of this region. Thus, the carp starter developed at Fish Nutrition and Physiology Division of ICAR- Central Institute of Freshwater Aquaculture (CIFA), Bhubaneswar could be used at farm level as an efficient nursery feed for carp which would ensure better survival and growth.

#### **ACKNOWLEDGEMENTS**

The work was conducted in ICAR-CIFA, Bhubaneswar, India and financed under institute funded project. Authors acknowledge Dr P. Jayasankar, Director of the institute for providing official and moral support for conducting the research work.

# **REFERENCES**

AOAC. 2012. Official Methods of Analysis. Association of Official Analytical Chemists International (19<sup>th</sup> ed.), Gaithersburg, MD, USA: Association of Official Analytical Chemists.

Barrows, F.T. and Hardy, R.W. 2000. Feed Manufacturing Technology. In: Stickney RR, editor. Encyclopedia of aquaculture. New York: Wiley, p. 354-9.

BIS. 2018. *The National Standard Body of India for Fish Feed.* Bureau of Indian Standards.

Cheng, Z. J. and Hardy, R. W. 2003. Effects of extrusion processing of feed ingredients on apparent digestibility coefficients of nutrients for rainbow trout (*Oncorhynchus mykiss*). *Aquacult. Nutr.* 9: 77-83.

Das, K.C., Toppo, S., Mohanty, T., Pradhan, C., Mohanta, K.N. and Giri, S.S. 2016. Cost effective floating feeds for Indian Major Carps (IMC) by replacement of soyabean meal with alternative feed ingredients. *Indian J. Anim. Res.* 50: 526-528.

Das, K.C., Mohanta, K.N., Nayak, S.K., Mohanty T., Toppo, S. and Swain, P. 2018. Effect of Extrusion Temperature on Quality of Carp Floating Feed Prepared from Local Feed Resources. *Anim.Nutr. Feed Technol.* 18: 117-123.

Djissou, A.S.M., Vodounnou, J.V., Tossavi, C.E., Toguyeni, A., Fiogbe, E.D. 2016. Complete replacement of fish meal by unconventional proteins sources in diet of *Oreochromis niloticus* (L.,1758) fingerlings: growth performance, feed utilization and body composition. *Int. J. Fish Aquacult. Std.* 4:242-247.

AFIA. 2005. *Feed Manufacturing Technology V.* American Feed Industry Association, Inc., 15001 Wilson Blvd., Suite 1100, Arlington, VA 22209.

Gaylord, T.G., Sealey, W.M., Barrows, F.T., Myrick, C.A. and Fornshell, G. 2017. Evaluation of ingredient combinations from differing origins (fishmeal, terrestrial animal and plants) and two different formulated nutrient targets on rainbow trout growth and production efficiency. *Aquacult. Nutr.* 23: 1319-1328.

Hardy, R.W. and Barrows, F.T. 2000. Diet formulation and manufacturing In: Halver JE, Hardy RW (Eds.), *Fish Nutrition*, 3<sup>rd</sup> edition. Academic Press Inc, New York, N.Y, pp. 506-600.

ICAR. 2009. Grow out carp culture. *In: Aquaculture Technologies for Farmers* (Jena, JK, Das PC, Das BK, Mohapatra BC, Sarangi N, Modayil MJ., Vass KK., Ravichandra P, Ponniah AG, Syda Rao S, Eknath AE & Ayyappan S *eds*), *pp.* 17-22. *Indian Council of Agricultural Research*, New Delhi, India.

Levic J. 2010. *Thematic proceedings of 2<sup>nd</sup> workshop on* "Extrusion Technology in feed and food processing", International Feed Industry Federation, Edited by Institut for Food Technology Bulevar cara Lazara 1, 21000 Novi Sad. Muyang Group 2009. R&D Center, No.1, Muyang Road, Hanjiang Industrial Park Yangzhou Jiangsu 225127 China.

Received on 14-08-2019 and accepted on 20-12-2019

Indian J. Anim. Nutr. 2019. 36 (4): 382-387 doi: 10.5958/2231-6744.2019.00062.8

# Effects of Feeding Graded Levels of Distillers Dried Grains with Soluble (DDGS) With or Without Supplementation of Multi-enzymes on Blood Bio-Chemical Constituents of Indigenous Chicken

Ashim Kumar Saikia<sup>1</sup>, Robin Bhuyan<sup>2</sup>, Bibeka Nanda Saikia<sup>3</sup>, Digendra Nath Sarma<sup>4</sup>, Ranajit Roychaudhury<sup>5</sup>, Arundhati Bora<sup>6</sup> and Joga Dev Mahanta<sup>7</sup>

Department of Animal Nutrition, College of Veterinary Science, AAU, Khanapara, Guwahati-781 022, India

#### **ABSTRACT**

An experiment was conducted using distillers dried grains with soluble (DDGS) at 0, 10 and 20% levels with or without enzymes supplementation in the diets of indigenous chicken to study the effect on different blood bio-chemicals parameters. A total of one hundred and eighty (21 days old) indigenous chicks were divided into six groups viz.  $T_1$ ,  $T_2$ ,  $T_3$ ,  $T_4$ ,  $T_5$  and  $T_6$  containing 30 chicks in each group. The birds of  $T_1$  (control) and T, groups were offered the standard chick, grower and layer feeds as per BIS, 2007 without and with multi-enzymes (Xzyme), respectively. The birds of  $T_2$ ,  $T_4$ ,  $T_5$  and  $T_6$  groups were fed the rations containing 10% DDGS without and with enzymes and 20% DDGS without and with enzymes, respectively. The feeding trial was conducted for a period of 13 fortnights. The average levels of different blood parameters, estimated at the end of the experiment, viz. haemoglobin, glucose, serum protein, lipid, cholesterol, aspartate aminotransferase (AST), alanine aminotransferase (ALT), calcium and phosphorus ranged from 9.96±0.20 to 10.91±0.24 g/dl,  $185.08 \pm 0.54$  to  $188.67 \pm 1.86$  mg/dl,  $3.05 \pm 0.17$  to  $3.32 \pm 0.09$  g/dl,  $72.45 \pm 0.29$  to  $72.96 \pm 0.49$  mg/dl,  $130.91 \pm 0.59$  to 133.71±2.07 mg/dl, 71.66±0.44 to 72.28±0.40 U/L, 11.42±0.32 to 11.93±0.12 U/L, 10.16±0.44 to 10.81±0.29 mg/dl and 4.93±0.11 to 5.94±0.14 mg/dl, respectively. Non-significant (P>0.05) differences were observed with respect to average values of different blood parameters among the treatment groups except for blood inorganic phosphorus. Serum concentration of inorganic phosphorus was significantly (P<0.05) lower in T<sub>1</sub> as compared to other groups. It can be concluded that the incorporation of DDGS up to 20% level in the diets of indigenous chicken did not have any adverse effect on the blood bio-chemical constituents of experimental birds.

Key words: Blood bio-chemical, Distillers dried grains with soluble (DDGS), Enzyme, Indigenous chicken

#### INTRODUCTION

The maize and soybean meal are the major conventional sources of energy and protein, respectively, in poultry feeds. These ingredients are becoming scarce day by day due to their increased demand in livestock and poultry sector. Further, due to its use for production of biofuel ethanol, availability of maize for use in poultry sector may be challenging. So, utilization of nonconventional feed resources in the poultry feeding is indispensable. The replacement of costlier traditional ingredients with cheaper non-conventional ingredients is probably the most viable proposition to address this situation. Distillers dried grains with soluble (DDGS), a co-product of ethanol production process, has been identified as one of the promising non-conventional feed resource that can be used in the poultry ration as energy and protein source. In the recent years there is an escalation of DDGS production and its quality has also seen a positive change when derived from new generation ethanol plants (Panda et al., 2016). It is a good source of energy, protein, exogenous amino acids, B-group vitamins and minerals, including phosphorus (Koreleski and Swiatkiewicz, 2006; Thaker and Widyaratne, 2007; Min et al., 2008). The DDGS is also better source of fibre, protein and fat than cereal grains (Koreleski and Swiatkiewicz, 2008). However, DDGS contains considerable amount of non-starch polysaccharides (NSP) (Ward et al., 2008), which restrict its extensive use in poultry ration. In this regard, exogenous enzymes are able to offer nutritional benefits by hydrolyzing NSP (Cost et al., 2008). The use of appropriate enzymes to hydrolyze these compounds can increase the nutritional value of DDGS and enables greater inclusion in poultry diets (Juanpere

Corresponding author Email: ashimkumar.saikia@gmail.com

at al., 2005). In view of the above facts, the present study was undertaken to investigate the effect of dietary incorporation of distillers dried grains with solubles (DDGS) at different levels with or without multienzyme supplementation on the blood bio-chemical constituents of indigenous chicken.

# MATERIALS AND METHODS

All procedures followed in this experiment was approved by Institutional Animal Ethics Committee, Assam Agricultural University, Khanapara, Guwahati (770/ac/CPCSEA/FVSc/U/IAEC/17-18/479 dated: 09.08.2017). Following a completely randomized design, a total of one hundred and eighty numbers of 21 days old indigenous chicks were randomly distributed into six groups viz. T<sub>1</sub>, T<sub>2</sub>, T<sub>3</sub>, T<sub>4</sub>, T<sub>5</sub> and T<sub>6</sub> containing 30 chicks with 3 replicates of 10 chicks in each group. The birds were reared under deep litter system of management throughout the experimental period following standard and uniform managemental practices and in similar environmental condition (temperature 17-28°C, humidity 80-92% and rainfall 0-14 mm). The birds of T<sub>1</sub> group (control) were offered the standard chick, grower and layer feeds as per BIS, 2007 (Table 1). The birds of T<sub>2</sub> group were fed with the same standard chick, grower and layer feeds as per BIS, 2007 with supplementation of multi-enzyme (Xzyme). Maize DDGS was incorporated at 10% level in the rations of T<sub>3</sub> and T<sub>4</sub> groups, whereas, the rations of T4 group was additionally supplemented with Xzyme. In the same way, the birds of T<sub>5</sub> and T<sub>6</sub> groups were fed with rations containing 20% DDGS without and with enzymes, respectively. The feeding trial was conducted for a period of 182 days (13 fortnights) using chick feeds for first 3 fortnights (0-42 days), grower feeds for next 7 fortnights (43-140 days) and layer feeds for last 3 fortnights (141-182 days). Weighed quantities of feeds were offered thrice daily at 6.30 AM, 12.30 PM and 5.00 PM to the birds of different treatment groups ensuring ad libitum feeding throughout the study period. During the last week of the experiment, blood was collected from four numbers of birds from each experimental group at 9.30 AM, after three hours of post feeding. Blood samples were collected aseptically from brachial vein of birds in two different tubes, one part in clean sterilized tube containing EDTA for haemoglobin analysis and another part in clean sterilized glass tubes and kept in slanted position at room temperature for serum collection. The samples were then centrifuged at 3000 rpm for 5 minutes, serum harvested and transferred to 2 ml Eppendorf tubes which were then stored at -20°C for estimation of other blood biochemical parameters.

The blood haemoglobin and glucose as well as other blood bio-chemical constituents (serum total protein, cholesterol, lipids, AST, ALT, calcium and inorganic phosphorous) were analyzed by using commercial kits following particular protocols as per the manufacturers' guidelines. The statistical analyses of the experimental data were carried out according to the method described by Snedecor and Cochran (1994).

# RESULTS AND DISCUSSION

Total feed consumption per bird for the last fortnight (13th) and entire period of experiment were 109.25 and 11653, 110 and 11665, 108.75 and 11664, 111.50 and 11669, 110.25 and 11739, and 111.25 and 11748 g for groups  $T_1$ ,  $T_2$ ,  $T_3$ ,  $T_4$ ,  $T_5$  and  $T_6$ , respectively. Feed intakes (g/bird) were found to be comparable among the birds. Other research workers (Thacker and Widyaratne, 2007; Youssef et al. 2013; Hassan and Aqil, 2015) also reported no significant difference in feed consumption among different treatment groups with DDGS inclusion levels up to 10-15%. In this experiment, maximum and minimum total consumption was recorded in T<sub>6</sub> and T<sub>1</sub> groups, respectively. Similar trend was also observed by Pineda et al. (2008) who reported that the higher levels of incorporation of DDGS in the layer ration increased feed intake.

No significant (P>0.05) difference was observed in Hb. levels of experimental birds among various treatment groups (Table 2). However, a non-significant (P>0.05) increase in blood Hb. levels in birds fed DDGS supplemented with multi-enzyme was observed, which might be due to the positive effects of enzymes as well

Table 1: Composition of chick, grower and layer rations

Ingradiants (%)									Rations	one								
			Chick	ick					Grower	wer					Layer	/er		
	$\mathbf{T}_1$	$\mathbf{T}_2$	$T_3$	$T_4$	$T_5$	$T_{\epsilon}$	$T_1$	$T_2$	$T_3$	$T_4$	$T_{\rm s}$	T <sub>6</sub>	$\prod_1$	$T_2$	$T_3$	$\mathbf{T}_4$	$T_{\rm s}$	$\Gamma_{\epsilon}$
Maize	40.40	40.40	34.00	34.00	29.32	29.32	40.40	40.40	32.26	32.26	29.32	29.32	43.18	43.01	34.21	34.21	32.27	32.27
Soybean meal	15.50		15.50 10.50	10.50	5.00	5.00	15.50	15.50	10.00	10.00	5.00	5.00	25.00	25.10	19.85	19.85	14.30	14.20
Rice polish	5.00	5.00	5.00	5.00	5.00	5.00	5.00	5.00	5.00	5.00	5.00	5.00	5.00	5.00	7.00	7.00	8.00	8.00
Distillers dried grains with soluble	8	8	10.00	10.00	20.00	20.00	8	00	10.00	10.00	20.00	20.00	8	8	10.00	10.00	20.00	20.00
Di-calcium phosphate	1.30	1.30	1.20	1.20	1.00	1.00	1.30	1.30	1.20	1.20	1.00	1.00	1.30	1.30	1.20	1.20	1.00	1.00
Limestone powder	1.70	1.70	2.00	2.00	2.00	2.00	1.70	1.70	2.00	2.00	2.00	2.00	7.00	7.00	7.20	7.20	7.30	7.30
Methionine	0.10	0.10	0.07	0.07	0.08	0.08	0.10	0.10	0.07	0.07	0.08	0.08	0.10	0.10	0.07	0.07	0.08	0.08
284 Lysine	0.00	0.00	0.05	0.05	0.18	0.18	0.00	0.00	0.05	0.05	0.18	0.18	0.00	0.00	0.05	0.05	0.18	0.18
Salt	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27	0.27
$Minpremix^1$	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10	0.10
Toxin binder	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05
BrokenRice	7.05	7.05	10.00	10.00	10.00	10.0	7.05	7.05	10.00	10.00	10.00	10.00	00.9	6.50	10.0	10.0	00.9	00.9
De-oiled rice bran	28.53	28.53	26.76	26.76	27.00	27.0	28.53	28.53	29.00	29.00	27.00	27.00	12.00	11.50	10.00	10.00	10.45	10.50
Enzymes	8	0.05	8	0.05	8	0.05	8	0.05	8	0.05	8	0.05	8	0.05	8	0.05	0.00	0.05
Crude Protein (%)	19.25	19.37	19.33	19.41	19.22	19.33	15.49	15.53	15.54	15.61	15.45	15.53	17.55	17.65	17.51	17.53	17.47	17.48
$\mathrm{ME}^2(\mathrm{kcal/kg})$	2798	2798	2793	2793	2800	2800	2527	2527	2503	2503	2539	2539	2580	2580	2600	2600	2604	2604
	,		1	į	,	i i		ľ		,	,	,		(				

<sup>1</sup>Mineral mixture contains (per 1.2 kg): Calcium- 255 g, Phosphorous- 127.5 g, Magnesium- 6 g, Manganese- 1.5 g, Iron- 1.5 g, Iron- 1.5 g, Iodine- 325 mg, Copper- 4.2 g, zinc-9.6 g, Cobalt- 150 mg, Sulphur- 7.2 g, Potassium- 100 mg, Sodium- 6mg, Selenium- 10 mg, Vitamin A- 700000 IU, Vitamin D<sub>1</sub>- 700000 IU, Vitamin B- 250 mg, Nicotinamide- 1000 mg & Chromium- 78 mg.; Composition of multi-enzyme (Xzyme: Each kg of Xzyme premix contains: Lactic Acid Bacillus-30,000 million spores, Saccharomyces Cervisiae- 100 billion CFU, Amylase- 29,000 IU, Betaglucanase-4,05,000 IU, Phytase- 44,500 IU, Lipase- 31,000 IU, Protease- 7,40,000 IU, Cellulase- 5,500 IU, Pectinase -1,01,000 IU and Hemicellulase- 25,000 IU; <sup>2</sup>(Metabolizable Energy) Calculated value

as higher iron contents of DDGS. Similarly, Gupta *et al.* (2017) reported that the inclusion of DDGS at 5, 7.5 and 10% levels enhanced the Hb level as compared to control. further improvement in Hb level was observed when DDGS was supplemented with multi-enzyme.

The mean concentration of blood glucose were  $188.67\pm1.86$ ,  $188.40\pm1.96$ ,  $187.20\pm1.61$ ,  $187.61\pm1.37$ ,  $185.08\pm0.54$  and  $185.79\pm0.30$  mg/dl in  $T_1$ ,  $T_2$ ,  $T_3$ ,  $T_4$ , T<sub>5</sub> and T<sub>6</sub> groups, respectively. The mean values for blood glucose levels in birds of different groups were comparable though some numerical differences existed among the values. Findings of Jiang et al. (2013) was in the same line with the present study who reported that dietary inclusion of rice DDGS up to 25% did not affect the plasma content of glucose in juvenile red seabream. However, Gupta et al. (2017) reported that glucose level decreased significantly in rice DDGS supplemented groups. In this study, the non-significant (P>0.05) decrease in blood glucose levels in DDGS added groups might be due to lower starch and higher fibre contents in the rations of various treatment groups.

The highest  $(3.32\pm0.09 \text{ g/dl})$  mean total serum protein was recorded in the birds of  $T_6$  group while the lowest  $(3.05\pm0.17)$  value for same was recorded in  $T_1$  group (Table 2). No significant (P>0.05) difference was observed in total serum protein values of experimental birds among the treatment groups. Bor-Ling *et al.* (2011) also reported that feeding of corn DDGS at different levels (0, 6, 12 and 18) did not influence the serum

protein in laying hen. Numerically increased total serum values were recorded in  $T_2$ ,  $T_4$  and  $T_6$  groups, where enzymes were supplemented, in comparison to  $T_1$ ,  $T_3$  and  $T_5$  groups. Osmana *et al.* (2011) conducted a feeding trial in Hubbard broiler chicks using DDGS and reported higher plasma protein values in birds fed 20% DDGS along with enzymes supplementation. In this study, the non-significant (P>0.05) increase of serum total protein levels in enzyme treated groups might be due to the effect of protease present in the multi-enzyme used in the experiment.

There was no significant (P>0.05) difference in blood lipid values of experimental birds among the treatment groups (Table 2). Similarly, Bor-Ling et al. (2011) reported that feeding of corn DDGS at different levels (0, 6, 12 and 18) did not influence the serum triglycerides of laying hens. Findings of Jiang et al. (2013) are also in the same line with the present study, who reported that dietary inclusion of rice DDGS up to 25% did not affect the plasma content of triglycerides in juvenile red seabream. No significant (P>0.05) difference was observed in blood cholesterol values of experimental birds among different treatment groups (Table 2). Jiang et al. (2013) also reported that dietary inclusion of rice DDGS up to 25% did not affect the plasma content of cholesterol in juvenile red seabream. Activity of AST and ALT were similar among the groups (Table 2). The present findings corroborate well with that of Ghazalah et al. (2011), who reported substitution

Table 2. Mean (±SE) blood parameters in experimental birds of different treatment groups

Parameters	Treatments groups						P value
	T <sub>1</sub>	T <sub>2</sub>	T <sub>3</sub>	$T_4$	T <sub>5</sub>	$T_6$	
Haemoglobin (g/dl)	9.96±0.20	10.40±0.16	10.65±0.27	10.74±0.36	10.85±0.22	10.91±0.24	0.118446
Glucose (mg/dl)	188.67±1.86	188.40±1.96	187.20±1.61	187.61±1.37	185.08±0.54	185.79±0.30	0.439116
Total serum protein (g/dl)	3.05±0.17	3.20±0.13	3.18±0.09	3.25±0.10	3.21±0.10	3.32±0.09	0.714567
Total lipids (mg/dl)	72.51±0.08	72.45±0.29	72.96±0.49	72.84±0.19	72.78±0.30	72.64±0.09	0.771293
Cholesterol (mg/dl)	133.71±2.07	131.85±2.28	132.61±1.84	131.71±1.04	131.90±0.97	130.91±0.59	0.869824
AST (U/L)	72.28±0.40	72.09±0.04	72.00±0.12	71.66±0.44	72.11±0.25	71.97±0.43	0.831994
ALT (U/L)	11.79±0.33	11.75±0.10	11.93±0.12	11.42±0.32	11.69±0.17	11.67±0.28	0.776553
Serum calcium (mg/dl)	10.59±0.36	10.81±0.29	10.37±0.31	10.53±0.41	10.16±0.44	10.63±0.34	0.851216
Serum phosphorous (mg/dl)	4.93°±0.11	5.62b±0.11	5.56 <sup>b</sup> ±0.22	5.75 <sup>b</sup> ±0.17	5.69b±0.18	5.94 <sup>b</sup> ±0.14	0.009138

Means bearing the same superscript within the row do not differ significantly (P<0.01)

of soybean meals with DDGS at four levels (0, 25, 50 or 75%) and supplementation of Avizyme 1500 at two levels (0 or 0.075%) did not cause any change in the activity of these enzymes.

The mean blood ALT levels of experimental birds of different treatment groups ranged from 11.42±0.32 to 11.93±0.12 U/L. There was no significant (P>0.05) difference in blood ALT values of experimental birds among various treatment groups. The results of the present study was in agreement with the findings of Ghazalah *et al.* (2011), who reported from a feeding trial in Bovans Brown layers by substituting soybean meals with DDGS at four levels of 0 or 25 or 50 or 75% and adding Avizyme 1500 at two levels (0 or 0.075%) that there was no significant (P>0.05) difference among the groups for ALT levels.

The mean serum concentration of calcium of experimental birds of different treatment groups ranged from 10.16±0.44 to 10.81±0.29 mg/dl. No significant (P>0.05) difference was observed in average serum calcium values of experimental birds among the treatment groups. The overall values of blood calcium in all the treatment groups were found to be on higher side which might be due to the fact that the birds of all the groups were in egg laying stage at the time of collection of blood. The results of the present study were in agreement with the findings of Ghazalah et al. (2011), who reported that there was no significant difference among the birds for calcium levels in Bovans Brown layers. Hack et al. (2015) also noticed that serum calcium was not altered when hens were fed with diets containing DDGS.

The serum inorganic phosphorus values were found to be maximum  $(5.94\pm0.14 \text{ mg/dl})$  in  $T_6$  and minimum  $(4.93\pm0.11 \text{ mg/dl})$  in  $T_1$  groups (Table 2). Statistically significant (P<0.05) difference was observed in serum phosphorus levels between  $T_1$  (control group) and all other treatment groups. There was increased levels of serum phosphorus in all the DDGS as well enzyme supplemented groups in comparison to control group, which corroborated well with the findings of Ali *et al.* (2006), who reported that the addition of enzyme preparation to laying hen diet increased plasma

phosphorous level. Dorra *et al.* (2013) also observed significant differences in plasma phosphorous level with different inclusion level of corn DDGS. In the present study the significant difference in blood inorganic phosphorus levels between T<sub>1</sub> and rest of the treatment groups might be due to presence of higher amount of available phosphorus in DDGS and supplementation of phytase in the diet which are in concurrence with the observations of earlier researchers.

# **CONCLUSION**

Based upon the results of this experiment, it is evident that the incorporation of DDGS up to 20% level in the diets of indigenous chicken did not have any adverse effect on the blood bio-chemical constituents of experimental birds. Therefore, it can be concluded that DDGS can be used as a cheap source of protein and energy in the rations of indigenous chicken for economical and profitable poultry production.

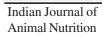
#### REFERENCES

- Ali, M.N., Hassan, M.S. and Abaza, I.M. 2006. Effect of improving the utilization of wheat bran on productive and physiological performance for local laying hens. *Egypt. Poult. Sci.* 26: 137-158.
- BIS. 2007. Bureau of Indian Standards: *Poultry Feed Specification*. 4<sup>th</sup> Review. Manak Bhawan, 9 Bahadur Shah Zafar Marg, New Delhi, India.
- Bor-Ling, S., Hsu, A.L. and Chen, Y.K. 2011. Effects of corn distillers dried grains with soluble on productive performance and egg quality of laying hens. Dept. of Ani. Sci., National Chia-Yi University and AGAPE Nutrition Consultant, Taiwan.
- Cost, F.G.P., Gourlart, C.C., Figueiredo, D.F., Oliveira, C.F.S. and Silva, I.H.V. 2008. Economic and environmental impact of using exogenous enzymes on poultry feeding. *Int. J. Poult. Sci.* 7: 311-314.
- Dorra, T.M.I., Kalaba, Z.M. and Younis M.E.E. 2013. Effect of using corn distillers dried with soluble in broiler diets without or with adding galzym on growth performance and serum physiological parameters. *J. Anim. Poult. Prod.* 4: 263-283.
- Ghazalah, A.A., Abd. Elsamee, M.O., Abd El-Hakim, A.S. and Ibrahim, M.M. 2011. Evaluation of using Distillers Dried Grains with Soluble (DDGS) in broiler diets. Egypt. *Poult. Sci.* 32: 381-397.
- Gupta, S.L., Tyagi, P.K., Tyagi, P.K., Mandal, A.B., Dinani, O.P. and Rokade, J.J. 2017. Feeding effect of rice based

- dry distiller's grains with solubles (DDGS) on hemato- biochemical and egg sensory attributes during 45<sup>th</sup> to 54<sup>th</sup> week of laying. *Int. J. Pure Appl. Biosci.* 5:1521-1527.
- Hack, M.E.A.E., Alaguwary, M., Rajab, M. and Kuldeep, F. 2015. Use of maize distillers dried grain with soluble (DDGS) in laying hen diets: trends and advances. *Asian J. Anim. Vet. Adv.* 10: 690-707.
- Hassan, S.M. and Al Aqil, A.A. 2015. Effect of adding different dietary levels of DDGS on productive performance of laying hens. *Int. J. Poult. Sci.*, 14: 279-284.
- Jiang, W., Zhang, L. and Shan, A. 2013. The effect of vitamin E on laying performance and egg quality in laying hens fed corn distillers dried grains with soluble. *Poult. Sci.* 92: 2956-2964.
- Juanpere, J., Perez, V.A.M., Angulo, E. and Brufau, J. 2005. Assessment of potential interactions between phytase and glycosidase enzyme supplementation on nutrient digestibility in broilers. *Poult. Sci.* 84:571-580.
- Koreleski, J. and Swiatkiewicz, S. 2006. Nutritive value and use of biofuel byproducts in poultry nutrition, *Wiadomosci ZOOT.* 44: 29-37.
- Min, Y.N, Liu, F.Z., Wang, Z., Coto, C., Cerrate, S., Costa, F.P., Yan, F. and Waldroup, P.W. 2008. Evaluation of DDGS in combination with glycerin in broiler diets. *Int. J. Poult. Sci.* 7: 646-654.
- Osmana M., Yakouta, H.M., El-Komyb, A.E.S. and Khalifahb, A.M. 2011. Effect of using distillers dried grains with solubles (DDGS) plus commercial enzymes on productive performance of broiler chickens. In: *Proc.* 18th European Symposium on Poultry Nutrition, Oct. 31–Nov. 04, 2011. Çesme Izmir, Turkey.

- Panda, A. K., Sahoo, B. and Kumar, A. 2016. Alternate feed resources in poultry. In: *Proc. XXXIII Annual Conference and National Symposium on "Rural Poultry for Livelihood, 2016*, College of Veterinary Science, Khanapara, Guwahati.1: 58-66.
- Pienda, L., Roberts, S., Kerr, B., Kwakkel, R., Verstegen, M. and Bregendahl, K. 2008. Maximum dietary content of corn dried distiller's grains with solubles in diets for laying hens: Effects on nitrogen balance, nanure excretion, egg production, egg quality. *Iowa State University Animal Industry Report* 2008, A.S. Leaflet, R 2334.
- Snedecor, G.W. and Cochran, W.G. 1994. Statistical Methods, 8<sup>th</sup> edn., The Iowa State University Press, Ames, Iowa.
- Swiatkiewicz, S. and Koreleski, J. 2008. The use of distillers dried grains with soluble (DDGS) in poultry nutrition. *World Poult. Sci. J.* 64: 257-266.
- Thacker, P.A. and Widyaratne, G.P. 2007. Nutritional value of diets containing graded levels of wheat distillers dried grains with solubles fed to broiler chicks. *J. Sci. Food Agric*. 87: 1386-1390.
- Ward, N.T., Zijlstra, R.T., Parsons, C. and Starkey, C. 2008. Non-starch polysaccharides (NSP) content of US commercial corn DDGS. *Poult. Sci.* 60, 1479-1484.
- Youssef, A.W., Abd El-Azeem, N.A., El-Daly, E.F. and El-Monairy, M.M. 2013. The impact of feeding graded levels of DDGS on broiler performance, hematological and histological parameters. *Asian J. Poult. Sci.*, 7: 41-54.

Received on 23-09-2019 and accepted on 28-09-2019







# Effect of Inclusion of Black Pepper Powder as Natural Feed Additive on the Performance of Japanese quail

V. Sri Divya, D. Srinivas Kumar\* and E. Raghava Rao

Department of Animal Nutrition, NTR College of Veterinary Science, Gannavaram – 521102, Andhra Pradesh, India

# **ABSTRACT**

This study was conducted to determine the effect of inclusion of black pepper (BP) powder in the diet on nutrient digestibility, production performance, and carcass characteristics of Japanese quail. One hundred and fifty, day-old quail chicks were randomly allotted into five experimental groups, each with 3 replicates of 10 chicks and were offered respective diets for 5 weeks. BP was ground and was included at 0% ( $T_1$ ; Control), 0.25 ( $T_2$ ), 0.50 ( $T_3$ ), 0.75 ( $T_4$ ) and 1.0 ( $T_5$ ) percent levels in iso-caloric and iso-nitrogenous broiler quail diets to meet the nutrient requirements. Results indicated increased body weight gain (P<0.05), decreased overall feed intake (P<0.01) and improved feed conversion ratio (FCR) (P<0.01) with inclusion of BP at 1.0% in the diet as compared to the control. There was a significant increase (P<0.01) in the digestibility of gross nutrients and fibre fractions with incorporation of BP at 1.0% in the diet as compared to the control group. Similarly, incorporation of BP at 1.0% level in the diet had increased the carcass yield, ready to cook yield (P<0.05), dressing percentage and meat to bone ratio (P<0.01) in quails as compared to the control. However, incorporation of BP upto 1.0% level in the diet had no effect (P>0.05) on feed cost/kg gain. Thus, it is concluded that BP can be included at 1.0% level in quail diets as natural feed additive without any adverse effects.

Keywords: Black pepper, Carcass characteristics, Nutrient digestibility, Production performance, Quail

#### INTRODUCTION

Dietary additives have an important role in animal production as growth and health promoters. Black pepper (BP) is one such alternative growth promoter. Black pepper (Piper nigrum) is a well-known medicinal plant and has many therapeutic effects like anti-ache effect (Moorthy et al., 2009), anti-oxidant and anti-bacterial effects (Gulcin, 2005). It is a good source of dietary protein and fatty acids for consumption by human and animals. The active principle, piperine present in BP has hepatoprotective and gastroprotective properties and can increase absorption of selenium, vitamin B-complex, β- carotene, curcumin and other nutrients (Khalaf et al., 2008). BP as an alternative to antibiotic growth promoters has been recommended for feeding broilers (Ghaedi et al., 2013; Valiollahi et al., 2013) while some other report observed no significant improvement in body weight gain and FCR (Puvaca et al., 2014).

Quail is considered a prolific meat and egg producer with dual production capacity and thus attained a valuable status in commercial enterprises. They are popular for their high protein (26%) and less fatty (3%) meat (Shinde *et al.*, 2014). However, studies on the feeding value of BP in Japanese quails are very limited. Hence, the present investigation was conducted to study the effect of BP powder inclusion at varying levels in the diet on growth performance, nutrient digestibility, carcass characteristics and cost economics in Japanese quails.

#### **MATERIALS AND METHODS**

One hundred fifty, day-old Japanese quail chicks were weighed individually (mean BW,  $7.42 \pm 0.38$  g), wing banded and randomly divided in to 5 equal groups of 3 replicates each with 10 chicks/replicate. The experiment was carried out for 5 weeks in a completely randomized design (CRD). During the experiment, BP seed was ground and included at 0% ( $T_1$ ; control), 0.25 ( $T_2$ ), 0.50 ( $T_3$ ), 0.75 ( $T_4$ ) and 1.0 ( $T_5$ ) percent levels in iso-caloric and iso-nitrogenous quail diets (Table 1) to meet the nutrient requirements (NRC, 1994). All the chicks were housed in battery brooders under uniform management conditions. Feed and water were provided *ad libitum*. The feed offered and feed left over was

 $<sup>*</sup> Corresponding \ author \ Email: kumardhulipalla@gmail.com$ 

Table 1. Ingredient (%) and chemical composition (% DM basis) of quail diets

Constituent		Cost/kg (₹)				
_	$T_1$	$T_2$	T <sub>3</sub>	$T_4$	T <sub>5</sub>	_
Maize	47.5	47.5	47.5	47.0	47.0	16.00
De-oiled rice bran	7.5	7.25	7.0	7.25	7.0	13.50
Soybean meal	36.5	36.5	36.5	36.5	36.5	38.00
Fish meal	5.00	5.00	5.00	5.00	5.00	30.00
Black pepper	0.00	0.25	0.50	0.75	1.00	550
Oil	0.50	0.50	0.50	0.50	0.50	20.00
Di-calcium phosphate	1.00	1.00	1.00	1.00	1.00	26.00
Shell grit	1.00	1.00	1.00	1.00	1.00	0.00
Salt	0.25	0.25	0.25	0.25	0.25	3.00
Trace min mix	0.15	0.15	0.15	0.15	0.15	240.0
Feed additives	0.60	0.60	0.60	0.60	0.60	1116.00
Total	100	100	100	100	100	
ME* Kcal/kg	2899.56	2902.39	2905.21	2902.29	2905.11	
Crude Protein (%)	23.98	23.99	23.99	24.00	24.00	
Feed cost/100 kg (₹)	2616	2750	2885	3017	3152	

\*Calculated Value

weighed daily, to quantify the feed utilized. The data for growth rate was recorded at weekly intervals.

During the last 3 days of the trial, faeces were collected of birds from all the 5 treatments which were previously fasted for 12 hours to empty the bird gut to mark the beginning and end of faeces collection. At the end of the collection period, the faecal samples collected from each treatment per day were pooled, ground and thoroughly mixed to obtain a homogenous mixture. Samples of feed and excreta samples were analyzed for proximate principles (AOAC, 2007). The retention of a nutrient was calculated by applying the following formula:

Nutrient retention (%) =

$$\frac{\text{Nutrient intake - nutrient excreted}}{\text{Nutrient intake}} \times 100$$

At the end of 5<sup>th</sup> week, two birds per replicate and thus six birds per treatment were randomly selected, weighed and slaughtered. The data on dressing percentage, carcass yield, meat bone ratio, ready-to-cook yield and per cent weight of heart, liver, gizzard and giblet were recorded. All the data were analyzed statistically (SPSS, version 17) as per Snedecor and Cochran (1993) and comparison of means was done

using Duncan's multiple range tests (DMRT) (Duncan, 1955).

#### RESULTS AND DISCUSSION

The ingredient composition of quail diets formulated by incorporating BP at varying levels was shown in Table 1. The diets were iso-nitrogenous and iso-caloric with a protein: energy ratio of 1:120. Animal fat was added to the diets to make up the energy deficit. The CP (12.87%) content of BP reported in the present study was higher than the value (9.3%) reported by Bolanle *et al.* (2014) and lower than the value (25.45%) reported by Mohammed *et al.* (2013).

Body weight gain, feed intake and FCR under different dietary treatments are presented in Table 2. The mean body weight gain was higher (P<0.05) in quails fed diets containing BP at 0.5,0.75 and 1.0 % level as compared to the control group, but inclusion at 0.25% level in feed did not show any significant effect (P>0.05) on body weight gain over control group. The increased body weight gains observed in quails may be attributed to the stimulating effect of BP on the digestive system thus leading to improved body weight gain. Further, the level of BP used reflects the high activity of piperine citrate included in the diet which may have affected the

Table 2. Effect of dietary inclusion of black pepper on performance and feed cost

Parameter		Level of				
		T <sub>2</sub>	T <sub>3</sub>	$T_4$	T <sub>5</sub>	significance
Body weight gain (g)	187.73°±3.57	195.04ab±3.52	198.53b±3.48	202.36b±3.41	205.36b±3.70	P<0.05
Overall feed Intake (g)	726.08°±7.07	701.11 <sup>d</sup> ±3.37	686.09°±3.45	659.60b±3.73	639.62a±1.15	P<0.01
Feed conversion Ratio	$3.90^{\circ} \pm 0.07$	$3.63^{b} \pm 0.07$	$3.46^{b}\pm0.07$	$3.26^{a}\pm0.06$	$3.11^{a}\pm0.07$	P<0.01
Feed cost / kg gain (₹)	102.02±1.80	99.825±1.90	99.82±1.80	98.35±2.08	98.03±0.86	NS

abcdeMeans with different superscripts in a row differ significantly

flow of digestive juices across the stomach (Al-Kassie et al., 2011). The results of the present study corroborated with earlier workers who also reported increased body weight gain in quails (El-Tazi et al., 2014; Valiollahi et al., 2014) with incorporation of BP powder in the diet. In contradiction to the present findings, Ndelekwute et al. (2015) reported that the final live weight gain was improved (P<0.05) in broilers fed BP at 0.25% in diet while there was a negative effect when fed at 0.75 and 1.0% level. However, Puvaca et al. (2014) reported that feeding BP upto 1.0% level in the diet had no effect (P>0.05) on body weight gain in broilers. Feed intake for overall growth period (0-5 weeks) was lower (P<0.01) in quails fed diets incorporated with BP at 0.25, 0.5 and 0.75 % and 1.0% as compared to the control group. Herbal feed additives inhibit the growth and colonization of various pathogenic microorganisms including E. coli in the intestinal tract of chicken due to their antimicrobial activity (Galib et al., 2010). When the number of harmful bacteria in the intestinal tract is low, more nutrients are absorbed by the birds. As a result, the birds gain higher weight with lower feed consumption. Another reason for decreased

feed intake with the addition of black pepper in the diet might be due to strong and spicy flavour that could reduce the palatability of the feed (Windisch *et al.*, 2008). Similarly, decreased feed intake upon feeding diets containing BP powder were reported by Ndelekwute *et al.* (2015), Ghaedi *et al.* (2013) and Tripathi *et al.* (2013) in broiler chicken. In contrast, increased feed intake upon feeding diets containing BP were also reported earlier (Al-Kassie *et al.*, 2011; Shahverdi *et al.*, 2013; El-Tazi *et al.*, 2014). However, Moorthy *et al.* (2009), Akbarian *et al.* (2012) and Valiollahi *et al.* (2014) reported that feeding BP in broiler diet had no effect (P>0.05) on feed intake.

The FCR improved significantly (P<0.01) with increased level of inclusion of black pepper up to 1.0% level in the diet of quails as compared to those fed control diet. The improved feed conversion ratio observed in quails fed diets containing BP may be attributed to the presence of piperine citrate which probably enhances the secretion of digestive juices in the stomach leading to greater efficiency in utilization of feed resulting in enhanced growth (Al-Kassie *et al.*, 2011). These results corroborated with the findings of

Table 3. Effect of dietary inclusion of black pepper on nutrient digestibility

Parameter		Level of				
	$T_1$	$T_2$	T <sub>3</sub>	$T_4$	T <sub>5</sub>	significance
Dry matter	65.23°±0.08	66.26b±0.40	67.27°±0.55	68.93 <sup>d</sup> ±0.38	69.16°±0.90	P<0.01
Organic matter	59.68°±0.33	61.10b±0.20	62.24°±0.10	$63.42^{d} \pm 0.09$	64.91°±0.24	P<0.01
Crude protein	55.58°±0.67	57.86b±0.18	58.04°±0.29	$60.36^{d} \pm 0.42$	$61.86^{e} \pm 0.33$	P<0.01
Crude fibre	47.06°a±0.22	49.98b±1.5	50.71 <sup>bc</sup> ±1.31	51.63bc±1.91	52.83d±0.39	P<0.01
Ether extract	49.87°±2.33	50.82b±2.19	55.58b±0.49	60.39°±2.12	$62.07^{d} \pm 1.15$	P<0.01
NFE	50.00°±0.45	51.66b±0.11	52.97°±0.16	54.10 <sup>d</sup> ±0.19	55.07 <sup>d</sup> ±0.16	P<0.01

abcde Means with different superscripts in a row differ significantly

Table 4. Effect of dietary inclusion of black pepper on carcass characteristics

Parameter		Level of				
		T <sub>2</sub>	T <sub>3</sub>	$T_4$	T <sub>5</sub>	significance
Carcass yield (g)	118.33°±2.7	123.83ab±2.3	126.17b±2.8	129.17b±2.4	129.67b±1.3	P<0.05
Dressing (%)	58.16 <sup>a</sup> ±1.15	59.59 <sup>b</sup> ±0.92	60.30 <sup>b</sup> ±1.6	$60.55^{b}\pm2.8$	$60.79^{b} \pm 1.0$	P<0.01
Ready to cook yield (g)	109.17 <sup>a</sup> ±1.6	113.33ab±2.1	115.50 <sup>bc</sup> ±2.1	119.33°±1.5	120.67°±1.8	P<0.05
Meat: bone ratio	$3.67^{a}\pm0.02$	$4.19^{b}\pm0.00$	4.61°±0.01	$5.20^{d} \pm 0.00$	$5.45^{e} \pm 0.02$	P<0.01
Heart (%)	$1.50\pm0.22$	$1.52 \pm 0.22$	$1.54 \pm 0.22$	1.67±0.21	1.90±0.16	NS
Liver (%)	$4.00\pm0.40$	$3.83 \pm 0.30$	$3.67 \pm 0.40$	$3.49\pm0.20$	2.83±0.40	NS
Gizzard (%)	$3.83 \pm 0.30$	$4.00\pm0.02$	4.17±0.30	4.33±0.20	4.76±0.20	NS
Giblet (%)	9.33±0.30	9.35±0.20	9.38±0.30	$9.39 \pm 0.30$	9.49±0.80	NS

abcdeMeans with different superscripts in a row differ significantly

other researchers (Al-Kassie *et al.*, 2011; Ghaedi *et al.*, 2013; Shahverdi *et al.*, 2013; Tripathi *et al.*, 2013; El-Tazi *et al.*, 2014; Valiollahi *et al.*, 2014). In contradiction, Ndelekwute *et al.* (2015) reported poor feed conversion ratio in broilers upon feeding BP in the diet.

Incorporation of BP up to 1.0% level in the diet of quails resulted in increased (P<0.01) digestibility of DM, OM, CP, CF, EE and NFE (%) as compared to the control group (Table 3). The improvement in the digestibility of proximate constituents observed with the addition of BP in the diet might be attributed to the ability of BP to induce saliva secretion, hydrochloric acid and mucus production (Srinivasan *et al.*, 2007). On the other hand, Jang *et al.* (2004) attributed better nutrient digestion to antimicrobial property of the essential oil in BP. In line with the present findings, Ndelekwute *et al.* (2015) also reported increased (P<0.05) digestibility of DM, CP, EE and CF with inclusion of BP upto 1.0% in the diet of broilers.

Data results pertaining to carcass characteristics are presented in Table 4. Inclusion of BP up to 1.0% level in the diet of quails resulted in increased carcass yield, ready to cook yield (P<0.05), dressing (%) and meat to bone ratio (P<0.01) as compared to the control. The increase in the carcass yield in quails observed in the present study may be attributed to the presence of essential oils in BP. Similarly, increased carcass yield (Ghaedi *et al.*, 2013; Shahverdi*et al.*, 2013) and dressing per cent(Al-Kassie *et al.*, 2011; El-Tazi *et al.*,

2014) were also reported earlier. However, Moorthy *et al.* (2009) reported that feeding BP powder at 0.2 % level in the diet had no effect (P>0.05) on dressed weight and ready to cook yield of broilers. Inclusion of BP up to 1.0% level in the diet had no effect (P>0.05) on per cent weight of heart, liver, gizzard and giblets in quails, which is in line with the findings of earlier workers (Moorthy *et al.*, 2009; Nath *et al.*, 2012; El-Tazi *et al.*, 2014).

The present study indicated that the feed cost/kg gain decreased by ₹ 2.19 in  $T_2$ , ₹ 2.20 in  $T_3$ , ₹ 2.67 in  $T_4$  and ₹ 3.99 in  $T_5$  group of quails fed diets containing BP at varying levels as compared to the control group (Table 2). Similarly, El-Tazi (2014) reported that inclusion of a mixture of red pepper and BP upto 1.0% level in broiler diets resulted in higher net profit/kg meat.

# **CONCLUSIONS**

The results of the present study indicated that incorporation of BP powder at 1.0 % level in the diet of quails had improved the performance of quails as evidenced from increased body weight gains, improved FCR and increased digestibility of nutrients in comparison to the control group. Thus, it is concluded that black pepper can be incorporated up to 1.0% level in the diet of quails without any adverse effect.

#### REFERENCES

AI-Kassie, G.A.M., Mamdooh, A.M., AI-Nasraw, S., and Ajeena, J. 2011. Use of black pepper (*piper nigrum*) as feed additive in broiler diet. *Res. Opin. Anim. Vet. Sci.* 1: 169-173.

- Akbarian, A., Golian, A., Kermanshahi, H., Gilani and Moradi, S. 2012. Influence of turmeric rhizome and black pepper on blood constituents and performance of broiler chickens. *Afr. J. Biotechnol.* 11: 8606-8608.
- AOAC (2005) Official Methods of Analysis. 18<sup>th</sup>edition, Association of Official Analytical Chemists, Washington, D.C. USA.
- Bolanle, A.O., Funmilola, A.S. and Adedayo, A. 2014. Proximate analysis, mineral contents, amino acid composition, anti- phytochemical screening of Brachystegia eurycoma harms and *Piper guineense* schum and thonn. *Am. J. Food Nutr.* 2: 11-17.
- Duncan, D.B. 1955. Multiple range and multiple F-tests. *Biometrics*. 11: 1-42.
- El-Tazi, S.M., Mukhtar, M.A., Mohamed, K. and Tabidi, M.H. 2014. Effect of using black pepper as natural feed additive on performance and carcass quality of broiler chicks. *Int. J. Pharm. Res. Anal.* 4: 108-113.
- Galib, A.M.A., Mamdooh, A.M.A. and Saba, J.A. 2011. Use of black pepper (*Piper nigrum*) as feed additive in broilers diet. *Roavs*. 1: 169-173.
- Ghedi, H., Nasr, J., Kheir, F., Miri, Y. and Rahimian, Y. 2013. Effect of use virginiamycin as probiotic, black pepper extract as phytogenic feed additive on performance of broiler chicks. *J. Agric. Sci.* 3: 521-525.
- Gulcin, I. 2005. The antioxidant and radical scavenging activities of black pepper (*Piper nigrum*) seeds. *Int. J. Food. Sci. Nutr.* 56: 491-499.
- Jang, I.S., Ko, Y.H., Yang, H.Y., Ha, J.S., Kim, J.Y., Kang, S.Y., Yoo, D.H., Nam, D.S., Kimand, D.H. and Lee, C.Y. 2004. Influence of essential oil components on growth performance and the functional activity of the pancreas and small intestine in broiler chickens. *Asian-Australas. J. Anim. Sci.* 17: 394-400.
- Khalaf, N.A., Shakya, A.K., Othman, A.A.L., EL-Agbar, Z. and Farah, H. 2008. Antioxidant activity of some common plants. *Turk. J. Biol.* 32: 51-55.
- Mansoub, N.H. 2011. Comparison of using different level of black pepper with probiotic on performance and serum composition of broiler chickens. *J. Basic Appl. Sci. Res.* 1: 2425-2428.
- Moorthy, M., Ravikumar, S., Viswanathan, K. and Edwin, S.C. 2009. Ginger, pepper and curry leaf powder as feed additive in broiler diet. *Int. J. Poult. Sci.* 8: 779-782.
- Mohamed, S.A. and El-Tazi, W. 2014. Response of broiler chicken to diets containing different mixture powder levels of red pepper and black pepper as natural feed additive. *Anim. Vet. Sci.* 2: 81-86.
- Nath, D.D., Rahman, M.M., Akter, F. and Mostofa, M. 2012.

- Effects of tulsi, blackpepper and cloves extract as a growth promoter in broiler. *Bangl. J. Vet. Med.* 10: 33-39.
- Ndelekwute, E.K., Afolabi, K.D., Uzegbu, H.O., Unah, U.L. and Amaefule, K.U. 2015. Effect of dietary Black pepper (*Piper nigrum*) on the Performance of Broiler. *Bang. J. Anim. Sci.* 44: 120-127.
- NRC 1994. *Nutrient Requirements of Poultry*. 9th Edition. National Academy Press, Washington, DC., U.S.A.
- Puvaca, N., Ljubojevic, D., Lukac, D., Kostadinovic, L., Stanacev, V., Popovic, S., MiliBalos, M.Z. and Nikolova, N. 2014. Digestibility of Fat in Broiler Chickens Influenced by Dietary Addition of Spice Herbs. *Macedonian J. Anim. Sci.* 4: 61-67.
- Shahverdi, A., Kheiri, F., Faghani, M., Rahimian, Y. and Rafiee, A. 2013. The effect of use red pepper (*capsicum annum* L) and black pepper (*Piper nigrum* L) on performance and hematological parameters of broiler chicks. *Eur. J. Zool. Res.* 2: 44-48.
- Shinde, A.S., Laxmi, C., Ingale, A.M., Upadhyay, D. and Munj, C.P. 2014. Quail farming small but worthy. *Poultry Line*. September 2014.
- Snedecor, G.W. and Cochran, W.G. 1993. *Statistical Methods*. 9<sup>th</sup> ed. IOWA, USA: Iowa State University Press.
- SPSS. 2008. *Statistical Packages for Social Sciences*, Version 17.0, SPSS Inc., Illinois, USA.
- Srinivasan, K. 2007. Black Pepper and its pungent principle-piperine. A Review of diverse physiological effects. *Food Sci. Nutr.* 47: 735-748.
- Tripathi, D., Kumar, A., Mondal, B.C., Rahal, A. and Palod, J. 2013. Effect of ajwain, hot red pepper and black pepper on the performance of Japanese quail. *Indian J. Anim. Nutr.* 30: 431-433.
- Valiollahi, M.R., Rahimain, Y., Miri, Y. and Rafiee, A. 2013. Effect of Use of Ginger (*Zingiberofficinale*), Black pepper (*Piper nigrum* L) powders on performance, some blood parameters and antibody titer against new castle vaccine on broiler chicks. *Scholarly J. Agric. Sci.* 3: 535-540.
- Windisch, W., Schedle, K., Plitzner, C. and Kroismayr, A. 2008. Use of phytogenic products as feed additives for swine and poultry. *J. Anim. Sci.* 86: 140-148.
  - Received on 18-08-2019 and accepted on 27-12-2019



# Indian J. Anim. Nutr. 2019. 36 (4): 393-398 doi: 10.5958/2231-6744.2019.00064.1

# Chemical Evaluation and Nutrient Digestibility of Shrimp Waste Meal in Broilers

N. Mounica, J.V. Ramana\*, D. Srinivasa Rao, J. Suresh and P. Kavitha

Department of Animal Nutrition,

College of Veterinary Science, SVVU, Tirupati-517502, Andhra Pradesh, India

#### **ABSTRACT**

This experiment was conducted to study the nutrient utilization in broilers fed shrimp waste meal (SWM) at various levels supplemented with or without amino acids as a substitute for fish meal in broiler diets. The percent dry matter, crude protein, ether extract, crude fibre, total ash, nitrogen free extract of shrimp waste meal (SWM) were 94.17, 50.5, 8.2, 15.2, 19.1, and 7.0, respectively. The calcium and phosphorous content of the SWM was 6.0, and 1.2 \%, respectively. The percent lysine and methionine were 1.66, and 0.88, respectively. Metabolizable energy content of SWM was calculated as 1515 kcal/kg diet. The experimental diets in pre-starter phase were prepared by replacing fish meal protein of the basal diet with the shrimp waste meal protein at 20% level  $(T_2)$ , 30% level  $(T_3)$ ; and  $T_4$   $T_5$  diets were prepared by adding synthetic lysine and methionine to  $T_2$  and  $T_3$ diets. In starter and finisher phases, five experimental diets were prepared by replacing fish meal protein of the basal diet with the shrimp waste meal protein at 50% level (T<sub>2</sub>), 100% level (T<sub>3</sub>); and T<sub>4</sub> T<sub>5</sub> diets were prepared by adding synthetic lysine and methionine to T<sub>2</sub> and T<sub>3</sub> diets. The basal diet T<sub>1</sub> was used as control containing maize, SBM, DORB and 10 % fish meal. All diets were iso-nitrogenous and iso-caloric. Metabolism trials were conducted to study the digestibility of nutrients during starter (0-28 days) and finisher (29-42 days) phases. The nutrient digestibility of DM and CF was found to be non-significant among treatments during starter and finisher phase. However, during starter and finisher phases CP and EE digestibilities were significantly (P<0.01) higher in birds fed  $T_4$  diet when compared to birds fed other diets  $(T_1, T_2, T_3 \text{ and } T_5)$ . The results of the present study conclude that the protein from fishmeal can be safely substituted up to 30% with the SWM protein in pre-starter and up to 50% in starter and finisher broiler diets along with the supplementation of synthetic lysine and methionine.

Key words: Amino acids, Broilers, Chemical composition, Fish meal, Nutrient digestibility, Shrimp waste meal

# INTRODUCTION

Escalating cost of poultry feed has promoted the use of alternative feed sources, especially agroindustrial by-products. To avoid the problem of disposal of agro-industrial wastes, which pose threat to the environment, can be better utilized as potential feed resources. Shrimp waste meal is one of such unconventional protein source that has the potential of being an alternative source of protein in broiler rations, partially or totally replacing conventional protein sources like fish meal. Shrimp waste meal is the dried and milled waste of the shrimp industry which consists of heads, shells and appendages of shrimp (Ingweye *et al.*, 2008). Shrimp waste contains high CP content and reasonably good balance of essential amino acids (Ngoan *et al.*, 2000a). It is particularly rich in lysine which makes

it an ideal supplement for cereals (Fanimo *et al.*, 1996). It is palatable and has a pleasant aroma. This study was therefore designed to provide an insight into the chemical composition and the nutrient utilization potential of diets containing various levels of SWM with or without amino acids supplementation.

# **MATERIALS AND METHODS**

Shrimp waste meal used in the preparation of experimental diets was procured from Om Sai Aqua Industry, Venkateswarapuram, Nellore district. Feed ingredients like maize, soybean meal, fish meal and de-oiled rice bran (DORB) for preparation of experimental diets were procured from the local market. Representative samples of feed ingredients were analyzed for their proximate composition (AOAC, 2005). The ME values for all feed ingredients except fish meal

was estimated using equation suggested by NRC (1994), while for fish meal the ME value was derived as per equation suggested by Leeson and Summers (2008). Representative samples of feed ingredients and broiler diets were analyzed for calcium and phosphorous as per the method of Talapatra *et al.* (1940).

Three hundred and seventy-five, day old commercial broiler chicks were distributed randomly to five treatments with three replicates of twenty five birds each. Experimental diets were formulated for pre-starter (0-14 days), starter (15-28 days) and finisher (29-42 days) phases. The experimental diets in pre-starter phase were prepared by replacing fish meal protein of the basal diet with the SWM protein at 20% level  $(T_2)$ , 30% level (T<sub>3</sub>); and T<sub>4</sub> T<sub>5</sub> diets were prepared by adding synthetic lysine and methionine to T<sub>2</sub> and T<sub>3</sub> diets. In starter and finisher phases five experimental diets were prepared by replacing fish meal protein of the basal diet with the SWM protein at 50% level (T<sub>2</sub>), 100% level (T<sub>2</sub>) and T<sub>4</sub> T<sub>5</sub> diets were prepared by adding synthetic lysine and methionine to T<sub>2</sub> and T<sub>3</sub> diets. The basal diet T<sub>1</sub> was used as control containing maize, SBM, DORB and 10 % fish meal.

All the five diets during each phase were made iso-nitrogenous and iso-caloric and other ingredients like palm oil, L-Lysine and DL-Methionine were also used to meet the dietary concentration for energy, protein, lysine and methionine as per ICAR (2013) specifications for broiler diets. The CF and AIA content of the basal diets was within the permissible limits of BIS (1992). The synthetic lysine and methionine were added to the T<sub>4</sub> and T<sub>5</sub> diets as the SWM was having these amino acids in lesser amounts as compared to FM.

Metabolism trials were conducted during the starter and finisher phases of the biological trial. Two birds from each replicate, thus a total of six birds per treatment were kept separately in six metabolic cages. Birds in the cages were fed with the respective experimental diets consecutively for 3 days and the total feed offered was weighed and recorded for each cage. Similarly feces voided and feed left over in each cage was carefully collected, weighed and recorded.

The representative samples of experimental diets offered and fecal samples from each cage were collected separately and analyzed for dry matter, crude protein, ether extract and crude fiber as per AOAC (2005). All the data obtained in this experiment were subjected to analysis of variance (Snedecor and Cochran, 1994).

# RESULTS AND DISCUSSION

The proximate composition of shrimp waste meal revealed that it contained 94.17% DM, on a dry matter basis it contained 50.5 % CP, 8.2 % EE, 15.2 % CF, 19.1 % TA, 7.0 % NFE, 6.0% Ca, 1.2 % P, 1.6 % lysine, 0.88% methionine and 1515 kcal ME /kg diet. The nutrient content (%DM) of feed ingredients, ingredient composition and nutrient content (%DM) of experimental diets are presented in the Tables 1,2,3 and 4.

The CP content of SWM in the present study was 50.5 % which is similar to the values reported by Rosenfeld *et al.* (1997). On the contrary lower, CP values were reported by Okonkwo *et al.* (2012) and Mahata *et al.* (2008). The EE, CF, TA values of SWM used in the present study were higher than those reported by Rosenfeld *et al.* (1997). This variation among different studies may be due to the difference in shrimp species (Ngoan *et al.*, 2000; Heu *et al.*, 2003), source of shrimp meal (head/shell) (Meyers, 1986) and/or processing method, as these can affect the nutritional values of SWM. Shrimp waste meal made from heads contains more CP and EE, less CF and TA than shrimp shells (Meyers *et al.*, 1973).

During starter phase and finisher phases no significant differences were noticed regarding the digestibility of DM and CF among treatments (Table 5 and 6). These findings were on line with Khempaka *et al.* (2011) who reported that DM, OM and ash digestibility values and nitrogen retention did not change significantly when SWM was added in the ration at or below 15% level. Ojewola and Annah (2006) also reported non-significant differences regarding fat, ash, crude fibre digestibility and nitrogen retention among treatment groups fed with SWM.

During starter and finisher phases the digestibility

Table 1. Chemical composition of feed ingredients used in the experiment

Constituents			Ingredients		
_	Maize	Soybean meal	Fish meal	De-oiled rice Bran	Shrimp waste meal
Dry matter (%)	90.2	90.8	91.17	93.6	94.17
		%D	M		
Crude protein	10.5	45.00	55.00	13.0	50.5
Ether extract	1.50	1.02	2.5	0.80	8.20
Crude fibre	6.7	9.46	2.4	16.8	15.2
Nitrogen free extract	75	36.28	19.6	53.9	7.0
Total ash	6.30	8.24	20.50	15.5	19.1
Calcium	0.01	0.20	6.50	0.06	6.0
Phosphorus	0.13	0.3	3.50	0.80	1.2
Lysine*	0.2	1.28	3.5	0.51	1.66
Methionine*	0.2	0.33	1.19	0.29	0.80
ME (k.cal /kg)**	3300	2300	2200	1400	1515

<sup>\*</sup>Calculated as per Panda et al. (1984); \*\*Calculated as per Leeson and Summer (2008).

of CP and EE was significantly (P<0.01) higher in birds fed  $T_4$  ration (50% SWM protein + amino acids) and lower values were observed with the rations in which

fish meal protein was completely substituted with SWM protein. Similar findings were reported by Mahata *et al.* (2008) that there was a significant decrease in the

Table 2. Ingredient composition (%) and chemical composition (%) of broiler pre-starter diets

Ingredients	$\mathbf{T_{_{1}}}$	$\mathbf{T_2}$	$T_3$	$\mathbf{T_4}$	$\mathbf{T_{5}}$
Maize	58	57.9	58	57.9	58
Soybean meal	20.55	20.4	20.3	20.34	20.21
Fish meal	10	8	7	8	7.0
Shrimp waste meal	0	2.18	3.27	2.18	3.27
De-oiled rice bran	5	5	4.83	5	4.83
Palm oil	3.7	3.77	3.85	3.77	3.85
Mineral mixture*	2	2.0	2.0	2.0	2.0
DL-methionine	0.2	0.2	0.2	0.21	0.21
L-lysine	0.55	0.55	0.55	0.6	0.63
Feed additives**	+	+	+	+	+
Total	100	100	100	100	10
Chemical compositi	ion				
CP (%)	22.1	22.05	22.0	22.08	22.04
ME (kcal/kg)	3002	2998	2999	2997	3000
Lysine (%)	1.18	1.15	1.13	1.20	1.19
Methionine (%)	0.51	0.51	0.50	0.52	0.51
Ca (%)	1.01	1.02	1.02	1.02	1.02
Available P (%)	0.7	0.66	0.63	0.66	0.63
Cost (₹/kg)	28.7	28.96	28.89	29.04	29.04

 $^{\circ}$ Contained Ca, 25; P, 15; NaCl, 2.5; Fe, 0.35% and Cu, 100; Mn, 200; Co, 50; I, 100 ppm;  $^{**}$ All diets contained Meriplex® – FDS @ 10g/100 kg: (Each gram contains: Vit-B<sub>1.</sub> 8mg; Vit-B<sub>6.</sub> 16 mg Vit-B<sub>12.</sub> 80µg; Vit-E<sub>50.</sub> 80 mg; Niacin, 120 mg; Folic acid, 8 mg; Calcium D Pantothenate, 80 mg, Merivite®-AB<sub>2</sub>D<sub>3</sub>K@10g/100kg: (Each gram contains: Vit-A 82,500 IU, Vit-B<sub>2.</sub> 52mg, Vit-D<sub>3.</sub> 1200 IU, Vit-K 10mg, Calcium 166 mg, Phosphate 395 mg) and Cosmodot @ 50g/100 kg: (3-5, Dinitro-O-Toluamide:25 percent W/W)

Table 3. Composition of broiler starter and finisher diets

Ingredient	Т	1	7	2	T	3		$\Gamma_4$	7	$\overline{\Gamma_5}$
	Starter	Finisher	Starter	Finisher	Starter	Finisher	Starter	Finisher	Starter	Finisher
Ingredient compo	sition (	<b>%</b> )								
Maize	59	65	60.25	65	59.51	64.81	60.25	65	59.77	65.24
Soybean meal	19.3	14	19.5	14.2	19.8	14.7	19.39	14.06	19.3	14
Fish meal	10	10	5	5	0	0	5	5	0	0
Shrimp	0	0	5.45	5.45	10.89	10.89	5.45	5.45	10.89	10.89
De-oiled rice bran	5	4.8	3	3.65	2.5	2.5	3.0	3.65	2.5	2.5
Palm oil	4.1	3.8	4.2	4.3	4.7	4.7	4.2	4.3	4.7	4.7
Mineral mixture*	2	2	2	2	2	2	2	2	2	2
DL-methionine	0.17	0.1	0.17	0.1	0.17	0.1	0.18	0.11	0.2	0.13
L-lysine	0.43	0.3	0.43	0.3	0.43	0.3	0.53	0.43	0.64	0.54
Feed additives**	+	+	+	+	+	+	+	+	+	+
Total	100	100	100	100	100	100	100	100	100	100
Chemical compos	sition (%	DM Ba	asis							
CP	21.5	19.55	21.49	19.5	21.5	19.5	21.5	19.6	21.5	19.5
ME (kcal/kg)	3049	3096	3049.3	3102	3048	3099	3047	3099	3049.2	3098
Lysine	1.09	0.92	0.99	0.83	0.99	0.74	1.01	0.93	1.01	0.92
Methionine	0.48	0.4	0.46	0.39	0.4	0.37	0.47	0.4	0.47	0.4
Ca	1.01	1.07	1.01	1.09	1.02	1.01	1.01	1.08	1.01	1.09
Available P	0.7	0.69	0.58	0.57	0.46	0.45	0.58	0.57	0.46	0.45
Cost (₹/kg)	28.39	26.39	28.30	26.53	28.49	26.71	28.53	26.82	29.06	27.56

\*Contained Ca, 25; P, 15; NaCl, 2.5; Fe, 0.35% and Cu, 100; Mn, 200; Co, 50; I, 100 ppm; \*\* Il diets contained Meriplex® – FDS @ 10g/100 kg : (Each gram contains: Vit-B $_1$ , 8mg; Vit-B $_6$ , 16 mg Vit-B $_{12}$ , 80µg; Vit-E $_{50}$ , 80 mg; Niacin, 120 mg; Folic acid, 8 mg; Calcium D Pantothenate, 80 mg, Merivite®-AB $_2$ D $_3$ K@10g/100kg : (Each gram contains: Vit-A 82,500 IU, Vit-B $_2$  52mg, Vit-D $_3$  1200 IU, Vit-K 10mg, Calcium 166 mg, Phosphate 395 mg) and Cosmodot @ 50g/100 kg: (3-5, Dinitro-O-Toluamide:25 percent W/W)

Table 4. Chemical composition (%) of broiler basal experimental diets on dry matter basis

Nutrient		<b>Basal diets</b>	
-	Pre-Starter	Starter	Finisher
Dry matter	90.1	90.36	91.06
Crude protein	22.04	21.50	19.57
Crude fat	5.37	5.12	5.18
Crude fibre	6.16	6.98	6.96
Total ash	10.40	10.43	10.56
Acid insoluble ash	2.08	2.15	2.16
Nitrogen free extract	56.03	55.97	57.73
Calcium	0.97	1.0	0.94
Phosphorus	0.57	0.55	0.56
Lysine*	1.20	1.09	0.97
Methionine*	0.52	0.48	0.47
ME (k.cal/kg)*	3000	3049	3099

<sup>\*</sup>calculated values

Table 5. Effect of supplementation of shrimp waste meal on digestibility of nutrients during starter phase

Treatment		Digesti	bility %	
_	DM <sup>NS</sup>	<b>CP</b> **	EE**	CF <sup>NS</sup>
$T_1$	66.70±0.32	68.23°±0.10	79.02ab±0.13	30.45±0.09
$T_2$	66.90±0.17	69.06b±0.36	79.23°±0.23	30.24±0.15
$T_3^2$	66.70±0.1	66.78 <sup>d</sup> ±0.09	78.49 <sup>b</sup> ±0.32	29.94±0.23
$T_4$	67.60±0.25	$70.25^{a}\pm0.1$	79.72°±0.35	30.70±0.06
$T_5$	66.78±0.26	67.05 <sup>d</sup> ±0.07	79.11ab±0.12	30.15±0.28

abcdValues in a column not sharing common superscripts differ significantly \*\*(P<0.01), NS- Non-significant

Table 6. Effect of supplementation of shrimp waste meal on digestibility of nutrients during Finisher phase

Treatment	eatment Digestibility %					
_	DM <sup>NS</sup>	<b>CP</b> **	EE**	CF <sup>NS</sup>		
$T_1$	70.77±0.2	68.58 <sup>b</sup> ±0.22	77.9 <sup>b</sup> ±0.21	31.49±0.02		
$T_2$	70.64±0.04	$68.99^{ab} \pm 0.29$	77.79b±0.28	31.61±0.06		
$T_3$	70.47±0.27	67.22°±0.25	77.08°±0.27	31.39±0.09		
$T_4$	71.22±0.23	69.51°±0.19	$78.6^{a}\pm0.10$	31.01±0.33		
$T_5$	70.41±0.14	67.84°±0.17	77.33 <sup>bc</sup> ±0.21	31.19±0.25		

abe Values in a column not sharing common superscripts differ significantly \*\*(P<0.01), NS- Non-significant

nitrogen retention when SWM was used beyond 8% level of inclusion in the diet. The enhanced nutrient digestibility in birds fed T<sub>4</sub> ration may be due to reduced number of pathogenic bacteria *Escherichia coli, Salmonella typhimurium* (Choi *et al.*, 1994; Le M ieux *et al.*, 2003; Wang *et al.*, 2003), increased immune function (Gibson and Roberfroid, 1995; Patterson and Burkholder, 2003), better secretion of digestive enzymes (Hou and Gao, 2001) and better absorption of nutrients (Wu, 1998; Huang *et al.*, 2005; Li *et al.*, 2007; Khambualai *et al.*, 2008, 2009).

The exoskeleton of the shrimp is composed mainly of chitin, an N-acetylated glucosamine polysaccharide that forms part of the protein complex, and is considered to have low digestibility when fed to animals (Austin *et al.*, 1981). Inclusion of SWM at higher levels in broiler diets caused impairment of digestion due to increase in chitin levels as it physically blocks the access of digestive enzymes to lipids and proteins, thus affecting the utilization of these nutrients (Castro *et al.*, 1989; Karasov, 1990).

### **CONCLUSION**

Results indicates that shrimp waste meal can be utilized as a potential alternative protein source to fish meal in broiler diets. Along with the supplementation of synthetic lysine and methionine, the protein from FM can be safely substituted up to 30% with the SWM protein in pre-starter and up to 50% in starter and finisher broiler diets.

#### **REFERENCES**

AOAC 2005. Official Methods of Analysis, 15<sup>th</sup> edn. Association of Official Analytical Chemists. Washington, D.C.

Austin, P.R., Brine, C.J., Castle, J.E. and Zikakis, J.P. 1981. Chitin: New facets of *research*. *Science*. 212: 749-753.

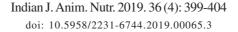
BIS. 1992. Bureau of Indian Standards. *Requirements for Chicken Feeds*. IS 1374-1992. Manak Bhavan, 9, Bhadursha Zafar Marg, New Delhi.

Castro, G., Stoyan, N. and Nyers, J.P. 1989. Assimilation efficiency in birds, a function of taxon and food type. *Comp. Biochem. Physiol.* 92: 271-278.

Choi, K. H., Namkung, H. and Paik, I. K. 1994. Effects of dietary fructo oligosaccharides on the suppression of intestinal colo-nization of *Salmonella typhimurium* in

- broiler chickens. Korean J. Anim. Sci. 36: 271-284.
- Fanimo, A. O., Mudama, E., Umukoro, T. O. and Oduguwa, O.O. 1996. Substitution of shrimp waste meal for fish meal in broiler chicken rations. *Trop. Agric*. (Trinidad) 73: 201-205.
- Gibson, G.R. and Roberfroid, M.B. 1995. Dietary modulation of the human colonic microbiota: Introducing the concept of prebiotics. *J. Nutr.* 125: 1401-1412.
- Heu, M.S., Kim, J.S. and Shahidi, F. 2003. Components and nutritional quality of shrimp processing by-products. *Food Chem.* 82: 235-242.
- Hou, Q.L. and Gao, Q.S. 2001. *Chitosan and Medicine*. Shanghai Science Technology Press, Shanghai, China.
- Huang, R.L., Yin, Y.L., Wu, G.Y., Zhang, Y.G., Li, T.J., Li, L.L., Li, MX., Tang, Zhang, J., Wang, B., He, J.H. and Nie, X.Z. 2005. Effect of dietary oligo chitosan supplementation on ileal digestibility of nutrients and performance in broilers. *Poult. Sci.* 84: 1383-1388.
- ICAR. 2013. *Nutrient requirements of Poultry*. ICAR publications, New Delhi.
- Ingweye, J.N., Okon, B.I., Ubua, J.A. and Essien, A.I. 2008. Performance of broiler chicken fed fish and shrimp waste. *Asian J. Anim. Sci.* 2: 58-63.
- Karasov, W.H. 1990. Digestion in birds: Chemicaland physiological determinants and ecological implications. *Stud. Avian Biol.* 13: 391-415.
- Khambualai, O., Yamauchi, K.E., Tangtaweewipat, S. and Cheva-Isarakul, B. 2008: Effects of dietary chitosan diets on growth performance in broiler chickens. *J. Poult. Sci.* 45:206-209.
- Khambualai, O., Yamauchi, K.E, Tangtaweewipat, S. and Cheva-Isarakul, B. 2009: Growth performance and intestinal histology in broiler chickens fed with dietary chitosan. *Br. Poult. Sci.* 50: 592-597.
- Khempaka, S., Chitsatchapong, C. and Molee, W. 2011. Effect of chitin and protein constituents in shrimp head meal on growth performance, nutrient digestibility, intestinal microbial populations, volatile fatty acids and ammonia production in broilers. *J. Appl. Poult. Res.* 20: 1-11.
- Leeson, S. and Summers, J.D. 2008. *Scott's Nutrition of the Chicken* 12<sup>th</sup> edn. Guelph, Ontario, Canada.
- Le Mieux, F.M., Southern, L.L. and Bidner, T.D. 2003. Effects of mannan oligosaccharides on growth performance of weanling pigs. *J. Anim. Sci.* 81: 2462-2487.
- Li, X.J., Piao, X.S., Kim, S.W., Liu, P., Wang, L., Shen, Y.B., Jung, S.C. and Lee, H.S. 2007. Effects of Chitooligosaccharide supplementation on performance, nutrient digestibility and serum composition in broiler

- chickens. Poult. Sci. 86: 1107-1114.
- Mahata, M.E., Dharma, A., Ryanto, I. and Rizal, Y. 2008. Effect of substituting shrimp waste hydrolysate of Peneus merguensis for fish meal in Broiler. *Pak. J. Nut.* 7: 806-810.
- Meyers, S.P., Rutledge, J.E. and Sonu, S.C. 1973. Variability in proximate analysis of different processed shrimp meals. *Feedstuffs*. 45: 34.
- Meyers, S.P. 1986. *Utilization of Shrimp Processing Wastes*. Infofish Marketing Digest, National Agriculture and Food. 4: 18-19.
- National Research Council. 1994. *Nutrient Requirement of Poultry*. 9<sup>th</sup> rev. edn. National Academy Press. Washington, D.C.
- Ngoan, L.D., Lindberg, J.E., Ogle, B. and Thomke, S. 2000a. Anatomical proportions and chemical and amino acid composition of com-mon shrimp species in central Vietnam. Asian-Australas. J. Anim. Sci. 13: 1422-1428.
- Ojewola, G.S. and Annah, S.I. 2006. Nutritive and economic value of Danish fish meal, Cray fish dust meal and Shrimp waste meal inclusion in broiler diets. *Int. J. Poult. Sci.* 5: 390-394.
- Okonkwo, A.C., Apkan, I.P. and Issac, L.J. 2012. Performance and carcass characteristics of finisher broilers fed shrimp waste meal. *Agric. J.* 7: 270-272.
- Patterson, J. A. and Burkholder, K.M. 2003. Prebiotic feed additives: Rational and use in pigs. Pages 319-332 in *Proc. 9<sup>th</sup> Int. Symp*. Digest. Physiol. Pigs. Banff, Alberta, Canada.
- Rosenfeld, D.J., Gernat, A.G., Marcano, J.D., Murillo, J.G., Lopez, G.H. and Flores, J.A. 1997. The effect of using different levels of shrimp meal in broiler diets. *Poult. Sci.* 76: 581-587.
- Snedecor, G.W. and Cochran, W.S. 1994. *Statistical Methods*. 8<sup>th</sup> edn. Oxford and IBH Publishing Co., Kolkata.
- Talapatra, S.K., Roy, S.C. and Sen, K.C. 1940. Estimation of phosphorous, chlorine, calcium, magnesium and potassium in food stuffs. *Indian J. Vet. Sci Anim. Husb.* 10: 243-258.
- Wang, X.W., Du, Y.G., Bai, X.F. and Li, S.G. 2003. The effect of oligo chitosan on broiler gut flora, microvilli density, immune function and growth performance. *Acta Zoonutr. Sin.* 15: 32-45.
- Wu, G. 1998. Intestinal mucosal amino acid catabolism. *J. Nutr.* 128: 1249-1252.
  - Received on 13-08-2019 and accepted on 25-12-2019



## Growth Performance, Carcass Traits and Economics of Kadaknath birds Rearing under Intensive Condition in Hot and Humid Climate

P.K. Jena<sup>1</sup>, B. Panigrahi<sup>1</sup>, N. Panda<sup>2\*</sup>, L.M. Mohapatra, B.K. Mallik<sup>3</sup>, J. Bagh<sup>1</sup>, P.K. Pati<sup>4</sup> and A. Baliarsingh<sup>5</sup>

Department of Livestock Production and Management, College of Veterinary Science and Animal Husbandry, Odisha University of Agriculture and Technology, Bhubaneswar-751003, Odisha, India

#### **ABSTRACT**

This experiment was conducted to study the growth performance, carcass traits and economics of Kadaknath chicken reared under intensive managemental condition in hot and humid climate of Odisha. A total of 104 day-old straight run chicks of Kadaknath breed of chicken were taken for this experiment, divided into 4 replicates, each comprising of 26 chicks and reared in deep litter system. The weekly body weight, feed conversion ratio (FCR), and mortality of birds were recorded upto 20<sup>th</sup> weeks of age and carcass traits were studied at the end of the experiment. No significant difference of body weight gain (P>0.05) between male and female chicks was noticed upto 4<sup>th</sup> week, while males showed higher (P≤0.05) body weights gains from 5<sup>th</sup> to 20<sup>th</sup> week of age. The incidence of mortality recorded were 3.8% in the first week of age, whereas, no further mortality of birds was noticed during the rest of the experimental period. Males had higher (P<0.01) slaughter weight, eviscerated weight and drumstick percentage, while females had higher (P<0.01) breast and giblet percentage. Breast muscle showed higher crude protein, total ash compared to thigh muscle. From the above study it is concluded that Kadaknath breed can profitably be reared under intensive system of management in hot and humid climatic condition.

Key words: Carcass trait, Economics, Kadaknath chicken, Intensive rearing, Growth

#### INTRODUCTION

Kadaknath is an important indigenous breed of poultry inhabiting vast areas of Western Madhya Pradesh, mainly the Jhabua and Dhar Districts and adjoining areas of Gujarat and Rajasthan. It tolerates extreme climatic conditions and thrives very well under minimal management inputs. It shows appreciable degree of resistance to diseases compared to other exotic breeds of fowl (Thakur et al., 2006). Back yard poultry farming is a part and parcel of typical rural/tribal household. The meat and eggs are also reckoned to be a rich source of protein (Rao and Thomas 1984). The meat of the Kadaknath breed contains high percentage (25.47%) of protein and is believed to have aphrodisiac properties (Mohan et al., 2008). Continual efforts are to be made for the improvement of this breed through effective healthcare, nutrition, management and breeding. However, most of the experiments that have been conducted on Kadaknath are under free range system of management. The present investigation was planned to study the growth performance and carcass traits of Kadaknath poultry birds reared intensive system in hot and humid climate of Odisha.

#### MATERIALS AND METHODS

The research work was carried out at the Central Poultry Development Organization (CPDO), Eastern region, Bhubaneswar from February to June. The maximum and minimum temperature and relative humidity were recorded daily and mean was calculated weekly. The weekly average maximum temperature ranged from 31.30 to 39.50°C, minimum temperature ranged from 17.80 to 28.10°C. Similarly, the weekly average maximum relative humidity ranged from 79.00 to 95.00 % and minimum ranged from 32.00 to 75.00 %. The experiment was conducted on 104 day-old straight run chicks of Kadaknath breed. Birds were divided into 4 replicates, each comprising of 26 chicks. All chicks were vaccinated with Marek's vaccine and

\*Corresponding author Email: npandaouat@gmail.com; <sup>1</sup> Department of Livestock Production and Management, <sup>2</sup> Department of Animal Nutrition, <sup>3</sup> Central Poultry Development Organisation (ER), <sup>4</sup> Department of Livestock Products Technology, <sup>5</sup>Nodoal officer, GKMS College of Veterinary Science and Animal Husbandry, Odisha University of Agriculture and Technology, Bhubaneswar-751003

wing banding was done at hatchery before transfer of chicks to the experimental shed. The experimental birds were reared under deep litter system and placed in the naturally ventilated house providing with 1 sq ft floor space per bird. Male and female birds were differentiated at 14th week by observing their comb pattern, then at 21 weeks of age birds were shifted to layer house. During the experiment, birds were offered weighed quantities of feed starting from day one till the end of the experiment as per the feeding practices carried out at CPDO, Bhubaneswar. All the birds were fed chick mash (0-8 weeks of age), grower mash (9-16 weeks of age) and breeder mash (17 week onwards). The weighed quantity of feed was offered daily in the morning and in the afternoon while fresh drinking water was supplied ad libitum. The composition of rations fed to the birds during different phases is presented in Table.1.

The body weight of birds was recorded individually at weekly intervals upto 20th weeks of age using a digital electronic balance. The birds were

weighed in the morning before supplying feed. The weekly body weight gain at a given age was found out by deducting the body weight of the previous week from that of the current week. Daily feed given was weighed and recorded. Feed consumption was calculated by subtracting the residual feed at the end of each week from the total feed provided during that week to the birds in each replicate, from which cumulative feed consumption was calculated. Feed conversion ratio (FCR) was derived by dividing the feed consumed with weekly body weight gain. Mortality of chicks was recorded daily. The mortality of all the chicks of the replicate groups were added and expressed on weekly basis.

At the end of twenty weeks, 6 healthy birds (3 male and 3 female) were sacrificed for the carcass parameters. Before slaughter the selected birds were separated from the flock and fasted overnight but drinking water was provided *ad libitum* to facilitate proper bleeding and also to know their actual live weights. The birds were slaughtered by severing the jugular vein and

Table 1. Composition of rations provided to the birds

Feed ingredients (kg)	Chick mash	Grower mash	Breeder mash
	(0-8 weeks)	(9-16wks)	(17th week onwards)
Crushed yellow maize	45	45	45
Rice polish	22	25	15
Soya bean meal	30	19	27
De-oiled rice bran	-	8	6
Mineral Mixture	3	3	3
Shell grit	-	-	4
Total	100	100	100
Nutritive value (% DM basi	(s)		
CP	20.15	16.20	17.95
ME (kcal/kg)*	2608	2515	2588
Ether extract	4.15	3.98	4.25
Crude fibre	4.65	8.50	5.25
Total ash	10.52	11.25	9.85
AIA	3.52	4.56	4.58
NFE*	53.25	51.25	52.82
Calcium	1.20	1.12	3.45
Lysine*	1.23	0.85	1.10
Methionine*	0.65	0.54	0.60

<sup>\*</sup>Calculated value

carotid artery below the left ear lobe by a single incision and allowed to bleed for five minutes by holding the birds' heads down. After complete bleeding and cessation of movement the carcass weight was recorded. The carcass was then scalded at 55-58°C for 1 minute 30 seconds. This helps in easy removal of feathers and gives uniform skin colour along with good flesh quality.

The eviscerated carcass along with the edible offal was weighed and recorded as edible carcass yield. The total meat yield was calculated by subtracting the giblet weight (the weight of the heart without pericardium, liver without gall bladder and gizzard without the serous lining) from the weight of the edible carcass.

1. Dressing 
$$\% = \frac{\text{Dressed weight}}{\text{Live weight}} \times 100$$

2. Giblet weight 
$$\% = \frac{\text{Giblet weight}}{\text{Live weight}} \times 100$$

3. Neck 
$$\% = \frac{\text{Neck weigh t}}{\text{Eviscerated weight}} \times 100$$

4. Wing 
$$\% = \frac{\text{Wing weight}}{\text{Eviscerated weight}} \times 100$$

5. Back % = 
$$\frac{\text{Back weigh t}}{\text{Eviscerated weight}} \times 100$$

6. Breast % = 
$$\frac{\text{Breast weight}}{\text{Eviscerated weight}} \times 100$$

7. Thigh 
$$\% = \frac{\text{Thigh weight}}{\text{Eviscerated weight}} \times 100$$

8. Drumstick 
$$\% = \frac{\text{Drumstick weight}}{\text{Eviscerated weight}} \times 100$$

The proximate composition such as moisture, crude

Table. 2. Body weight, cumulative feed intake and cumulative FCR

Age	I	Body weights (g	)	Cumulative	Cumulative	Cumulative
(weeks)	Male	Female	Combined	feed intake/ bird (g)	body weight gain (g)	FCR
1	57.00±1.47	53.08±1.13	53.89±0.71	100±0.32	31.95±0.17	3.13±0.01
2	105.40±1.61	94.63±2.08	96.87±1.28	220.00±0.70	74.93±0.53	$2.93 \pm 0.02$
3	155.90±2.62	138.16±3.34	141.93±2.04	$350.00\pm0.72$	119.99±0.78	2.91±0.01
4	212.80±4.57	182.026±4.94	188.43±3.16	494.0±0.50	166.49±1.20	$2.96 \pm 0.04$
5	278.00±6.84**	232.47±6.80	241.95±4.45	652.0±0.75	220.01±0.64	$2.96 \pm 0.02$
6	348.80±9.55**	293.55±8.12	305.06±5.42	826.0±1.04	283.66±0.39	2.91±0.03
7	416.40±11.56**	349.00±9.73	363.04±6.53	1022.0±0.91	341.10±2.14	$2.99 \pm 0.04$
8	506.40±15.96**	413.31±11.30	432.35±8.08	1232.0±0.75	410.41±1.78	3.00,0.05
9	591.50±18.02**	476.44±12.30	500.41±9.12	1460.0±1.29	478.47±1.98	$3.05\pm0.06$
10	698.30±18.58**	559.52±13.06	588.43±10.02	1702.0±1.43	566.49±2.06	$3.00\pm0.04$
11	787.00±16.82**	628.71±9.04	661.68±10.31	1962.0±1.82	639.74±2.43	$3.06\pm0.12$
12	905.70±19.02**	706.86±9.65	748.29±11.91	2236.0±1.38	726.35±1.54	$3.07 \pm 0.03$
13	922.80±18.32**	718.47±10.03	761.04±12.20	2490.0±2.21	739.10±1.25	$3.36 \pm 0.02$
14	1028.40±20.45**	790.84±10.73	840.33±13.68	2776.0±1.32	818.39±2.87	$3.40\pm0.04$
15	1054.80±17.81**	821.50±11.48	870.10±13.78	3086.0±0.48	848.16±3.14	$3.63\pm0.02$
16	1143.80±21.42**	885.23±12.72	939.10±15.37	3368.0±1.88	917.16±2.16	$3.67 \pm 0.03$
17	1214.30±22.94**	941.42±13.76	998.27±16.42	3662.0±3.50	976.33±3.18	$3.75 \pm 0.05$
18	1274.20±23.95**	990.86±14.37	1049.89±17.09	3978.0±3.10	1027.95±3.36	3.86±0.06
19	1323.80±25.35**	1043.44±14.81	1101.85±17.33	4308.0±2.72	1079.91±2.74	$3.98 \pm 0.04$
20	1380.80±24.99**	1098.36±15.08	1157.20±17.51	4652.0±4.21	1135.26±3.48	4.10±0.07

<sup>\*\*</sup>Mean differ significantly within the rows (P<0.01)

protein, ether extract and total ash content of the chicken meat from the breast and thigh muscles were measured according to the procedure of AOAC (1995). The data obtained from the study were statistically analysed according to Snedecor and Cochran (1994).

#### RESULTS AND DISCUSSION

The body weight of males birds were 1028.40 g and female were 790.84 g at 20th week (Table-2). It was observed that males have significantly (P<0.01) higher body weight than females from 5th week to 20th week. But no difference in body weight was found upto 4th week. This may be due to higher rate of cell multiplication in males as well as influence of androgenic hormones on the growth rate of males (Chandra et al., 2004). In contrast to the present findings, Pathak et al. (2015) recorded higher body weights of Kadaknath at different ages i.e. from day old to 20 weeks under intensive system of management, but Jha et al. (2013) reported lower body weights for the same ages of Kadaknath. However, Thakur et al. (2006) found significantly lower body weights for the same genetic group under free range system. The higher body weight in intensive system might be due to the supplementation of balanced diet and other proper managemental care.

The body weight gains increased progressively from 1st week upto 20th week. The male chicks recorded significantly (P<0.01) higher cumulative body weight gain compared to female chicks from 5th week to 20th week. In this study, at the age of 20th week, the body weight was 1157.20 g which was comparable with the value of 1145.73 g for indigenous chickens reared under intensive system of management as reported by Doley et al. (2009). Thakur et al. (2006) reported that Kadaknath attain 886.12 g body weight at 20 weeks of age with an overall gain of 858.68 g during 0-20 weeks under field condition which was much lower than the present findings of 1135.26 g gain in body weight during the same age which could be attributed to better feeding and managemental practices adopted in the present study. Jha and Prasad (2013) reported that the body weight of Vanaraja, Gramapriya and Aseel at 20th week was 2013.39, 1574.31 and 1038.75 g respectively. It was observed that the bodyweight of Kadaknath was comparable with Aseel birds.

Data pertaining to the weekly feed intake and cumulative feed intake of the birds are presented in Table 2. The cumulative feed intake per bird increased progressively. At the end of 20<sup>th</sup> week the cumulative feed intake of Kadaknath bird was 4652. 00 g and the

Table. 3. Carcass traits and percentage yield of different cut up parts

Parameter	Male (g)	Female (g)
Live body weight (g)	1150.33±25.78*	964.66±2.96
Dressing (%)	74.17±1.33	75.46±0.52
Eviscerated weight	812.66±24.53*	686.33±6.64
Heart (g)	3.66±0.33	3.66±0.33
Liver (g)	17.00±2.00	18.00±0.57
Gizzard (g)	19.66±1.33	20.00±0.57
Giblet (%)	3.50±0.22	4.31±0.08*
Neck (%)	8.88±0.26	7.91±0.1
Wing (%)	13.52±1.13	15.10±0.07
Breast (%)	23.16±0.11	24.86±0.06**
Back (%)	21.06±1.08	20.30±0.23
Thigh (%)	16.96±0.44	16.70±0.04
Drumstick (%)	16.32±0.08**	15.10±0.03

<sup>\*/\*\*</sup> Mean differ significantly within rows (P≤0.05, P≤0.01)

Table. 4. Meat quality traits

Parameters (%)	Thigh muscle	Breast muscle
Moisture	74.83±0.60	73.77±0.40
Crude protein	80.08±0.90	83.31±0.44
Ether extract	9.07±0.40	6.26±0.13
Crude fibre	0.57±0.01	0.58±0.02
Total ash	3.66±0.15	7.75±0.18
Acid insoluble ash	$0.06 \pm 0.01$	$0.08 \pm 0.01$

cumulative body weight gain was 1135.26 g. The Feed conversion ratio of the birds upto 4th, 8th, 12th, 16th and 20th week were recorded to be 2.96, 3.00, 3.07, 3.67 and 4.10, respectively. An FCR value of 5.24 at 40 weeks of age was reported by Jha *et al.* (2013) in Kadaknath birds maintained under deep litter system of management. The mean cumulative FCR was 3.07 at 12th weeks of age which was in agreement with the observation of Gupta and Mehta (2016). They also found the FCR of 3.18 at 12 weeks of age in Kadaknath birds kept in deep litter system of housing. Jha and Prasad (2013) also reported that the FCR (upto 40th week) of the three coloured breeds like Vanaraja, Gramapyiya and Aseel were 4.28, 3.85 and 5.47, respectively.

During the whole experimental period mortality rate of 3.8% was recorded during starter period (0-8 weeks) which was lower than 12.86% reported by Malik *et al.* (2009). On the other hand, the mortality of birds was higher under extensive system (17.13%) followed by those under semi-intensive system (15.28%) and intensive system (14.35%), but the differences were statistically non-significant in indigenous chicken as reported by Doley *et al.* (2009).

Data pertaining to mean carcass traits for the male and female birds are presented in Table 3. The live weight of males and females that was selected for carcass study was 1150.33 g and 964.66 g, respectively. Similarly the eviscerated weight was 812. 66 and 686.33 g in males and females, respectively. As the live weights of the male birds was higher than females at 20 week, the eviscerated weight was also significantly higher in male birds. The breast yield (%) of the female birds was higher whereas the drumstick % was higher in male birds. No difference was observed with respect to other

cuts between the sex. Haunsi *et al.* (2013) reported lower dressing percent of 65-66% in Kadaknath and Aseel bird. But in the present experiment higher dressing % was observed. Other cut up parts percentage were similar as reported by Haunsi *et al.* (2013). Sharma and Naryankhedkar (2005) reported comparable carcass traits in the crosses of Kadaknath breeds.

From the proximate composition of thigh and breast muscle of male birds, it was found that the protein content of breast muscle (83.31%) was higher than thigh muscle (80.08%), whereas the ether extract of the thigh muscle was 9.07% as against 6.25% in breast muscle (Table 4). No difference was observed in moisture, crude fibre and acid insoluble ash content of both the muscle, but the total ash content of breast muscle was higher than the thigh muscle. Similar findings were also reported by Haunsi et al. (2013) who found higher protein value of 83.75 and 87.65 % for thigh and breast muscle, respectively from the meat of 20 week male Kadaknath bird. The economics of Kadaknath rearing in the intensive system was evaluated on 20 week taking the feed consumption and body weight gain (Table 5). From the dressing (%) the

Table. 5. Economics of production

Particulars	Values
Cumulative feed intake/bird (g)	4652
Cost of feed (₹/kg)	27.01
Feed cost/bird (₹)	125.60
Cumulative body weight gain/bird (g)	1135.25
Feed cost/ kg live weight gain (₹)	110.63
Dressing (%)	74.81
Feed cost/kg of meat (₹)	129.41

feed cost per kg of meat was also evaluated. The cumulative FCR up to  $20^{th}$  week was 4.1. As the  $20^{th}$  week cumulative weight gain was 1135.25 g and feed intake was 4652 g, the feed cost per bird was ₹ 125.60 feed cost and per kg live weight gain was ₹ 110.63. Taking the dressing % of 74.81 (male birds) into account, the feed cost per kg of meat produced was ₹ 129.41.

#### **CONCLUSION**

From the above study it could be concluded that Kadaknath breed of chicken can be reared under intensive system of management in hot and humid climatic condition. From the economics of production it was found that the cost of production of per kg live weight gain was ₹ 110.63.

#### **ACKNOWLEDGMENTS**

The authors are thankful to the Director, Central Poultry Development Organization (E.R), Bhubaneswar for providing necessary facilities to conduct the research work at their farm.

#### **REFERENCES**

- A.O.A.C. 1995. *Official Methods of Analysis*, Association of Official Analytical chemists, 13<sup>th</sup> edn. Washington DC
- Chandra, C.V.S., Rao, S., Rao, G.N., Sharma, R.P., Reddy, B.L.N., Gupta, B.R. and Satyanarayana, S. 2004. Genetic study on juvenile traits of Vanaraja chickens. *Indian. J. Anim. Sci.* 74: 1229-1231.
- Doley, S., Barua, N., Kalita, N and Gupta J.J. 2009. Performance of indigenous chickens of North-Eastern region of India under different rearing systems. *Indian J. Poult. Sci.* 44:249-252.
- Gupta, S and Mehta, M. 2016. Effect of cysteine protease in diet to reduce soybean meal without affecting performance of Kadaknath birds in India. *J. Anim. Res.* 6:951.
- Haunshi, S., Panda, A.K., Padhi, M.K and Bhanja, S.K. 2013.
  Effects of Varying Nutrient Densities on Production
  Performance of Aseel Layers. *Anim. Nutr. Feed Technol.* 15: 51-58.
- Jha, D.K and Prasad, S. 2013. Production performance of improved varieties and indigenous breed of chicken in Jharkhand. *Indian J. Poult. Sci.* 48: 109-112.
- Jha, D.K., Prasad, S., Soren, S.K and Bharti, A.2013. Production performance of indigenous chicken in

- intensive farming system. *Indian J. Poult Sci.* 48: 105-108.
- Malik S, Singh N.P. and Ngachan S.V. 2009. Study of the different economic traits of CARI Shyama in field and farm conditions of Tripura, *Indian J. Poult. Sci.* 44: 296-300.
- Mohan, J., Sastry, K.V., Moudgal, R.P and Tyagi, J.S. 2008. Performance profile of Kadakanath desi hens under normal rearing system. *Indian J. Poult. Sci.* 43: 379-381.
- Pathak, P., Dubey, P.P., Dash, S.K and Chaudhary, M.L. 2015. Studies on growth and carcass traits of Aseel and Kadaknath chicken. *Indian J. Poult Sci.* 50: 327-328.
- Rao, G.V and Thomas, P.C. 1984. The breed characteristics of Kadaknath breed of indigenous (Desi) chicken. *Avian Res.* 68: 55-57.
- Sharma, D.C and Narayankhedkar, S.G. 2005. Carcass evaluation traits of crosses between Kadaknath breed of Indian native chicken, Rhode Island Red and White Leghorn. *Indian Vet. J.* 82: 221-222.
- Singh, D.P. 2009. Overview of existing breeds and issues of conserving indigenous breeds. Pages 54-55 in *Proc. Natl. Workshop on Synthesizing Experiences in the Promotion of Backyard Poultry*. Commissioner, Rural Development, Govt. of Andhra Pradesh, Hyderabad, India.
- Snedecor, G.W. and Cochran, W.G. 1998. *Statistical Methods*, 5<sup>th</sup> ed. Oxford and IBH Publishing Company, Kolkata, India.
- Thakur, M.S., Parmar, S.N.S and Pillai, P.V.A. 2006. Studies on growth performance in Kadaknath breed of poultry. *Livest. Res. Rural Dev.* 18: 1-9.

Received on 22-11-2019 and accepted on 11-12-2019

# Effect of Dietary Supplementation of Carrot Meal on Survival, Growth and Pigmentation of Freshwater Ornamental Fish, Koi Carp, *Cyprinus Carpio* (L.)

Abhinka Jain<sup>1</sup>, Vaneet Inder Kaur<sup>2\*</sup> and Shathanagouda Admane Hollyappa<sup>3</sup> Department of Acquaculture, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana-141004, Punjab, India

#### **ABSTRACT**

The study was conducted to assess the efficacy of experimental diets supplemented with carrot meal (CM) @ 1, 3, 5 and 7 % with respect to pigmentation, survival and growth in freshwater ornamental fish koi carp, *Cyprnius carpio* (Linnaeus) for 120 days. Five diets including one control (CM0) without any supplemented carotenoid source and four supplemented experimental pelleted diets with 1% (CM1), 3% (CM3), 5% (CM5) and 7% (CM7) of powdered carrot meal were prepared. At the termination of the experiment, colouration studies indicated significantly higher (P<0.05) skin and muscle carotenoid (µg g<sup>-1</sup> wet weight basis) content in fish fed with CM5 (4.87) followed by diets CM3 (3.28), CM7 (2.78), CM1 (1.95) and CM0 (1.24). RGB colour intensity based on digital photographs analysis was also found to be highest in CM5 followed by CM3, CM7, CM1 and CM0. Fish growth in terms of % NWG, PER and SGR was highest in CM5 (94.09%, 1.48, 0.55), with lowest values of NWG and SGR in CM7 (65.71% and 0.41), while PER in CM1 (0.75). Further, supplementation of CM @ 5% in Koi carp diet also improved the FCR (2.16) as compared to control (4.17). 100% survival of fish was recorded with diets supplemented with CM @ 3, 5 and 7 %, while it was 80 % in CM0 and CM1. During the experimental period, the water quality parameters did not vary significantly among control and treatments and remained in optimum range for fish growth. Hence, carrot meal can be supplemented in koi carp diet up to 5 % level both as colour enhancer as well as growth promoter under indoor rearing conditions.

Key words: Carotenoids, Carrot meal, Colour enhancement, Koi carp, Pigmentation

#### INTRODUCTION

Ornamental fish are nature's wonderful creation and ornamental fish keeping is the second most preferred hobby in the world and this sector plays a vital role in the international fish trade, having high export value (Jain et al., 2016). The value of ornamental fish is determined by their unique body characteristics, good health and coloration. The beautiful and flamboyant coloration exhibited by ornamental fishes is due to the presence of carotenoid pigments along with overall nutrition. Therefore, there is a direct relationship between dietary carotenoids and pigmentation in fish (Halten et al., 1997). Carotenoids are the main pigments, representing a class of over 600 natural lipid soluble pigments found in plants, algae, photosynthetic, some non-photosynthetic bacteria and animals, which are required for healthy growth, metabolism, photosynthesis and reproduction, besides colour development. Chemical composition of carotenoids include approximately 20%  $\beta$ -carotene, 30%  $\gamma$ -carotene and 30% lycopene (oxygen free hydrocarbons) and 15% of oxygen containing carotenoids *i.e.* xanthophylls (Zeb, 2004). Fish contain various kinds of carotenoids, the proportion of which depends upon their physiological status and dietary conditions. Synthetic or natural carotenoids are required to be incorporated @ 50-400 mg kg<sup>-1</sup> in diet of indoor reared fish to develop colour similar to fish feeding on live food like algae, tubifex, zooplankton, *etc.* (Boonyaratapalin and Lovell, 1977) under outdoor conditions.

For supply of carotenoids, various synthetic as well as natural sources (animal and plant) are available and are being developed for use in aquaculture for food as well as ornamental fishes. Synthetic processes results in production of only specific carotenoids like  $\beta$ -carotene or astaxanthin, whereas, natural sources are always a combination of several carotenoids

\*Corresponding author Email: vinnygulati@gmail.com; ¹ICAR-Central Institute of Fisheries Education, Mumbai-400061, India; ²Department of Aquaculture; ³Department of Aquatic Environment; College of Fisheries, GADVASU, Ludhiana-Punjab, 141004, India

like α-carotene, β-carotene, zeaxanthin, lutein, cryptoxanthin etc. (Gupta et al., 2007). Moreover, synthetic carotenoids are expensive and their use in aqua feed is limited due to species specificity. Natural carotenoids of animal origin are mostly extracted from crustacean such as crayfish meal, shrimp meal, crab meal, etc. (Torrissen and Naevdal, 1984) and are rich source of astaxanthin. The major concerns with the production of animal-based carotenoid are limited supply due to declining catches of crustaceans along with high expenses, which leads to increase in cost of carotenoid incorporated aquaculture diets. Plant based natural carotenoids are mainly derived from flowers, fruits, vegetables, algae, yeast, etc. (Tsushima and Matsuno, 1998; Chapman, 2000), which are not only inexpensive, but also a potential source of mixed carotenoids.

Few studies have demonstrated the effect of natural carotenoids on the growth, survival, colour development and antioxidation responses in ornamental fishes. Among natural vegetable pigment source, carrot (family Apiaceae) is one of the important root vegetables rich in bioactive compounds *i.e.* carotenoids along with several other functional components having significant health promoting properties (Sharma *et al.*, 2012). The total carotenoid content in the edible portion of the carrot root range from 6-55 mg/100g (Simon and Wolff, 1987) with  $\beta$ -carotene (44-79 % of total carotenoids, 5.3 – 10 mg/100 g) as major component (Holland *et al.*,

Table 1. Composition (%) of experimental diets

1991) and is the single major source of β-carotene providing 17% of total vitamin A. Due to these properties, carrot alone or in combination with other natural antioxidants, is found to induce better growth, enhance colour intensity and improve health of ornamental fish in a natural way. In view of unique properties of carrot, the present study was designed to assess the efficacy of carrot (*Daucus carota*) meal on survival, growth and pigmentation of ornamental Koi, *C. carpio* (Linn.).

#### MATERIALS AND METHODS

The experimental fish, Koi carp, *C. carpio* L. were procured from ornamental fish culture unit of College of Fisheries, Guru Angad Dev Veterinary and Animal Sciences University, Ludhiana. The fishes were acclimatized under indoor conditions for one month with control diet. Ten uniformly sized fishes (av. length 2-4 cm, av. weight 1-2 g) were stocked in glass aquaria, having 50 liters of water with continuous oxygen supply. The experiment (in triplicate) was carried out for 120 days.

Four experimental supplemented pelleted diets were prepared by adding carotenoid source *i.e.*, carrot meal (CM) @ 1 % (CM1), 3 % (CM3), 5 % (CM5) and 7 % (CM7) in the basal diet (CM0) by adjusting the level of other feed ingredients (Table 1). Carrot was procured from market, washed, grated and dried under shade and grounded to prepare the powdered meal. All the feed ingredients were powdered, mixed thoroughly

Ingredients		E	xperimental die	ets	
	CM0(Control)	CM1	CM3	CM5	CM7
Soybean meal*	25	25	24.5	24	24
Groundnut meal*	25	25	24.5	24	23
Mustard meal*	25	25	24	24	23
Rice bran*	20	19	19	18	18
Vegetable oil	02	02	02	02	02
Vitamin-mineral mixture	02	02	02	02	02
Salt	01	01	01	01	01
Carrot meal	-	1	3	5	7

\*Solvent extracted

and palletized. The proximate composition of feed ingredients and experimental diets (Table 2) was analyzed by following the methods of AOAC (2000). Carotenoid analysis of pigment source (carrot meal) was done by following method of Cyanotech (2002). Pelleted feed was provided to the fish @ 5 % fish body weight twice a day. Amount of feed was adjusted at every sampling (fortnightly interval) according to increase in fish weight.

Water quality parameters in terms of water temperature, pH, dissolved oxygen, total alkalinity, total hardness and ammonical nitrogen were measured at fortnightly intervals by following the methods of APHA (1991). Survival of the fish was calculated by subtracting the number of fishes harvested at the end of the experiment from the fish stocked at the initiation of the experiment.

Fish growth in terms of total body length and body weight was recorded at fortnightly intervals. At the end of the experiment, total length gain (TLG), per cent total length gain (%TLG), net weight gain (NWG), percent net weight gain (% NWG), specific growth rate (SGR), feed conversion ratio (FCR), protein efficiency ratio (PER) and condition factor were calculated as follows

TLG = Final total body length (cm)-initial total body length (cm)

%TLG = Final total body length (cm)-initial total body length (cm) /Initial total body length (cm) ×100

NWG = Average final body wt. (g) - Average initial body weight (g)

%NWG = Final body weight (g) - initial body weight (g) /initial body weight (g)  $\times$  100

SGR (% increase in weight /day) =

 $\frac{\ln \text{ final body weight - ln initial body weight}}{\text{Culture days}} \times 100$ 

Where, ln = Natural logrithum

FCR = Feed given (g) / Weight gain (g)

PER = Weight gain (g) / Protein intake (g)

Condition factor (K) =

Body weight / (Body length) $^3 \times 100$ 

Carotenoid analysis of fish muscle and skin was done by following method of Olson (1979) at initiation, middle and termination of the experiment. The digital photographs of experimental fish were taken at monthly intervals with a Sony full HD camera (DSC-HX 300). Colour of the fish skin was measured from opercular region, base of dorsal fin and caudal peduncle area of at least three fishes from each replicate. While taking the photographs all camera conditions were kept constant.

Table 2. Proximate composition (% DM basis), gross energy (kcal/g) and total carotenoid content (mg/ 100g) of carrot meal and carrot meal incorporated diets

Ingredients/ feed	Crude protein	Ether extract	Crude fibre	Ash	NFE	Gross energy (kcal/g)	Carotenoid content (mg/ 100g)
Rice bran*	21.94	1.46	10.86	7.00	58.72	3.77	-
Ground nut meal*	35.62	0.76	20.26	8.40	34.94	3.50	-
Mustard meal*	34.67	0.76	14.40	8.00	41.56	3.78	-
Soybean meal*	39.37	1.46	9.20	6.13	43.82	4.14	-
Carrot meal	17.33	1.33	10.33	8.50	59.03	3.50	16.45
CM0	31.93	1.04	13.20	7.60	46.46	3.79	-
CM1	31.20	1.06	13.06	6.95	47.73	3.81	0.152
CM3	31.38	1.08	13.04	7.12	47.38	3.81	0.431
CM5	31.20	1.06	12.93	7.16	47.65	3.81	0.670
CM7	30.85	1.00	12.81	7.11	48.23	3.80	0.987

\*Solvent extracted

Photographs were analyzed in adobe photoshop (Hancz et al., 2003; Tlusty, 2005) to compare these on the basis of RGB (Red, Green and Blue) values. To quantify the colour of the fish skin from digital photographs, CIE (Commission International de l'Eclairage) Lab is applied. Adobe Photoshop has a colour space called "Lab colour mode" which is based on CIE Lab. It consists of three parameters: L\*, a\* and b\*. Where L\* stands for luminosity (lightness ranging from 0 for black to 100 for white), channel a\* is the balance between red / green and channel b\* describes the balance between yellow/ blue.

#### RESULTS AND DISCUSSION

The study revealed that during the experimental period, water quality parameters (Table 3) such as temperature, pH, dissolved oxygen, total alkalinity, total hardness and ammonical-nitrogen did not vary significantly among the treatments and were within the recommended range for ornamental fish culture (Wurts and Durborow, 1992; OATA, 2008; Rinna *et al.*, 2013).

At the termination of the experiment, 100% survival of fish was recorded with diets supplemented with CM @ 3, 5 and 7 %, while it was 80 % in control (CM0) and CM1 (Table 4), indicating positive effect of carrot on survival of fish. According to Krinsky (1993),

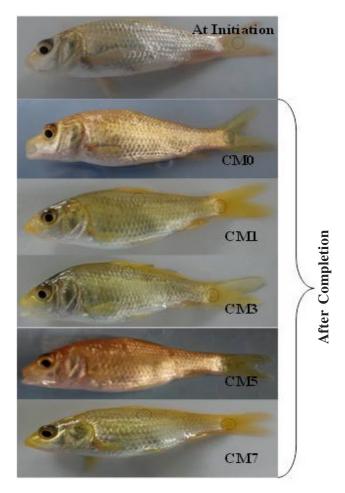


Fig. 1. Changes in skin colour of Koi carp, *C. carpio* L. before and after feeding

Table 3. Physico-chemical parameters of water in different treatments

Parameters	Diets							
	CM0(Control)	CM1	CM3	CM5	CM7			
Water temperature (°C)	29.28°±0.50	29.15°±0.62	29.36°±0.57	29.00°±0.65	29.24°±0.64			
	(27.50-31.76)	(26.66-31.70)	(26.73-31.93)	(26.20-31.86)	(26.33-32)			
pH	8.59°±0.08	8.50°±0.07	(8.24-8.75)	8.57°±0.09	8.61°±0.09			
	(8.33-8.90)	(8.24-8.76)	8.51°±0.07	(8.20-8.97)	(8.18-9.02)			
Dissolved oxygen (mgl <sup>-1</sup> )	6.78°±0.29	$6.86^{a}\pm0.28$	6.91°±0.28	$7.04^{a}\pm0.28$	7.07±0.22			
	(5.86-8.40)	(5.86-8.40)	(5.80-8.20)	(6.20-8.20)	(6.33-8.20)			
Total alkalinity(mg CaCO <sub>3</sub> l <sup>-1</sup> )	211.14 <sup>a</sup> ±12.2	203.33°±9.98	214.67°±11.79	214.22°±10.92	219.44°±14.1			
	(140-254)	(154-254)	(160-256)	(150-257)	(148-277)			
Total hardness(mg CaCO <sub>3</sub> l <sup>-1</sup> )	266.56°±5.5	269.33°±6.1	275.89°±5.7	274.11a±4.0	266.22°±6.68			
-	(235-282)	(246-298)	(255-296)	(252-292)	(229-296)			
$NH_3$ - $N (mgl^{-1})$	0.185°±0.020	0.179°±0.022	0.159°±0.027	$0.188^{a}\pm0.024$	0.171°±0.024			
J	(0.116-0.308)	(0.063-0.251)	(0.045-0.260)	(0.049-0.268)	(0.0440.249)			

Values are Mean ± S.E.; Values with different superscript (abcd) in a row differ significantly (P<0.05); values in parentheses indicate range

absorption of carotenoids enhances fish survival. In agreement to present study, Mirzaee et al. (2013) recorded colour enhancement in jewel cichlid (Hemichromis bimaculatus) with carrot as compared to red pepper. Likewise, Kop et al. (2010) reported carrot as an efficient carotenoid source in comparison to red pepper (Capsicum annum) for colour enhancement in cichlid (Cichlasoma severum). Comparison of carrot (D. carota) with three flower pigment sources i.e. marigold petal (Tagestes erecta), China rose petal (Hibiscus rosasinensis), rose petal (Rosa cinensis) also revealed that carrot resulted in significantly higher carotenoid content in fish skin/muscle (Ramamoorthy et al., 2010). Yanar and Tekelioglu (1999) also suggested utilization of carrot as potential carotenoid source for colour development in gold fish, Carassius auratus.

At the end of culture period, maximum average final total body length (cm) was observed in CM1 followed by CM7, CM5 and CM0 and CM3, respectively and the differences among treatments were significant (P<0.05). TLG and % TLG were also

maximum in CM1 and minimum in CM3 and the differences among different treatments were significant (P<0.05). The final body weight was highest in CM5 followed by CM1, CM0, CM3 and CM7 with significant differences. Likewise, NWG, %NWG and SGR was highest in CM5 and minimum in CM7. PER and CF were highest in CM5, while PER was lowest in CM0, and CF was lowest in CM1. Feed efficiency in terms of FCR also improved maximally in CM5 as compared to all other treatments and control.

Improved fish growth has been found in pigment supplemented diets as carotenoids are known to play a positive role in the metabolism of fish and might have improved nutrient utilization leading to improved growth (Amar *et al.*, 2001). In the present study also, it is clear that carotenoid do play a role in fish growth, as increasing level of CM (1-5%) resulted in positive effects on growth, in terms of increased weight gain and reduced FCR. Mirzaee *et al.* (2013) conducted comparative study to see the effect of carrot and red pepper (both providing 60 mg/kg of total pigment) on growth of jewel cichlid. Feeding of carrot resulted in

Table 4. Growth performance and of koi carp, C. carpio L. in different treatments

Parameters			Diets		
-	CM0(Control)	CM1	CM3	CM5	CM7
% Survival	80	80	100	100	100
Final total body length (cm	6.24 <sup>bc</sup> ±0.20	$6.48^{a}\pm0.19$	6.13°±0.21	6.24 <sup>bc</sup> ±0.11	$6.26^{bc} \pm 0.06$
TLG	0.70	0.96	0.62	0.82	0.67
		(+26.32%)	(-11.43%)	(+17.14%)	(-4.28%)
%TLG	12.63	17.39	11.25	14.49	11.98
Final body weight (g)	$3.83^{cd} \pm 0.44$	4.18b±0.26	$3.62^{cd} \pm 0.21$	4.27°±0.24	$3.48^{cde} \pm 0.16$
NWG	1.60	1.96	1.44	2.07	1.38
		(+22.50 %)	(-10.00 %)	(+29.37 %)	(-13.75 %)
% NWG	71.74	88.28	66.05	94.09	65.71
FCR	4.17	3.40	3.32	2.16	3.46
PER	0.75	0.94	0.95	1.48	0.93
SGR	0.45	0.53	0.43	0.51	0.42
CF	1.57	1.53	1.57	1.75	1.41

Values are Mean  $\pm$  S.E.; Values with different superscript (ab...d) in a row differ significantly (P<0.05); Values in parentheses indicate % change over control; TLG = Total length gain; NWG = Net weight gain, FCR = Feed conversion ratio, PER = Protein efficiency ratio, SGR= Specific growth rate, CF = Condition factor

significantly higher growth as compared to red pepper. Ramamoorthy et al. (2010) too showed enhanced growth of marine ornamental fish, Amphiprion ocellaris fed on four pigment sources (carrot, D. carota; marigold meal, T. erecta; China rose petal, H. rosasinensis and rose petal, R. chinensis) as compared to non-pigmented diet, where D. carota resulted in maximum growth rate as compared to other diets as well as control. Positive effects in terms of growth through carotenoid were also observed by Ahilan et al. (2008) in Gold fish, C. auratus, when amaranth and coriander were given @ 1 and 3 %, respectively. Spirulina, a rich source of carotrenoids, has also proved to be protein rich nutrient and resulted in significantly higher growth (@ 30 mg kg<sup>-1</sup> of diet) in blue gourami, Trichogaster trichopterus (Rinna et al., 2013) Similar to natural carotenoid sources, synthetic carotenoid source such as astaxanthin was also reported to influence fish and shellfish growth in a positive manner (Thongrod *et al.*, 1995). Liang *et al.* (2012) showed significant higher weight gain and specific growth rate along with significantly reduced FCR in red white koi carp with 150, 200 and 250 mg of astacin kg<sup>-1</sup> of diet. Similarly, Nguyen *et al.* (2014) found that supplementation of astaxanthin @ 80 mg kg<sup>-1</sup> in koi carp resulted in higher growth and feed utilization. However, few of the earlier reports suggested the role of carotenoids only on colour enhancement without having much effect on the fish growth (Kop *et al.*, 2010; Mirzaee *et al.*, 2013; Seyedi *et al.* 2013).

The result of the present study revealed maximum colour enhancement in terms of carotenoid content at 5% incorporation level of carrot meal as compared to both low (1% and 3%) and high incorporation level (7%). The digital photographs also exhibited improved values for RGB along with increased a\*and b\* values in CM5. In the present study, carrot meal resulted in significantly increased carotenoid

Table 5. Colouration parameters in skin of Koi carp (C. carpio L.) in different treatments

Days	Particulars			Diets*		
		CM0	CM1	CM3	CM5	CM7
			Caro	tenoid content (	μg/g)	
120		1.24°±0.011	$1.95^{d} \pm 0.008$	3.28b±0.035	4.87°±0.384	2.78°±0.018
			(57.25)	(164.52)	(292.74)	(124.19)
				<b>RGB Values</b>		
0	R	59.00°±1.17	59.00°±1.10	60.00°±1.11	59.00°±1.12	58.00°±1.15
	G	51.00°±1.21	50.00°±1.12	52.00°±1.15	52.00°±1.14	51.00°±1.16
	В	28.00°±1.20	29.00°±1.13	29.00°±1.14	30.00°±1.16	29.00°±1.19
120	R	$75.66^{d} \pm 2.60$	179.00 <sup>ab</sup> ±5.77	187.66°±3.17	189.00°±9.81	150.66 <sup>bc</sup> ±5.48
	G	$66.66^{de} \pm 0.33$	166.00 <sup>bc</sup> ±2.30	155.66 <sup>cd</sup> ±3.17	185.00°±0.57	150.00c <sup>cd</sup> ±4.61
	В	51.66 <sup>d</sup> ±3.17	$143.66^{bc} \pm 1.45$	174.66°±7.79	179.00°±1.73	167.00 <sup>ab</sup> ±5.19
				Lab values		
0	L*	22.00°±0.25	21.00°±0.30	22.00° ±0.25	21.00° ±0.24	21.00°±0.26
	a*	1.00°±0.26	$1.00^{a}\pm0.21$	$1.00^{a}\pm0.24$	$1.00^{a}\pm0.23$	$1.00^{a}\pm0.28$
	b*	16.00°±0.15	15.00°±0.19	17.00°±0.21	18.00°±0.22	18.00°±0.20
120	L*	$31.00^{cd} \pm 0.57$	$69.00^{ab} \pm 1.15$	64.66 <sup>b</sup> ±0.88	62.00 <sup>bc</sup> ±1.15	75.66°±0.33
	a*	$1.89^{de} \pm 0.57$	$4.00^{bc} \pm 0.00$	$3.66^{cd} \pm 0.88$	$7.66^{a}\pm1.45$	$3.00^{cd} \pm 0.57$
	b*	15.00°±2.00	$34.00^{cd} \pm 0.57$	$35.66^{bc} \pm 0.33$	39.00° ±2.00	$34.66^{bc} \pm 2.02$

Values are Mean  $\pm$  S.E.; Values with different superscript (ab...d) in row differ significantly (Pd" 0.05); Values in parentheses indicate % change over control

concentration at 5 % incorporation level, showing direct relationship with concentration of carotenoids on deposition of carotenoids. A positive relationship between feed dose and deposition of carotenoids in muscle of Atlantic salmon was also observed by Baker (2002). In agreement to present studies, Mirzaee et al. (2013) also recorded more colour enhancement in jewel cichlid (H. bimaculatus) with carrot as compared to red pepper. Likewise, Kop et al. (2010) also reported carrot (D. carota) as a more efficient carotenoid source in comparison to red pepper (Capsicum annum) for colour enhancement in cichlid (C. severum). Comparison of carrot with three flower pigment sources i.e. marigold petal (T. erecta), China rose petal (H. rosasinensis), rose petal (R. cinensis) also revealed that carrot resulted in significantly higher carotenoid content in fish skin/muscle (Ramamoorthy et al., 2010). Yanar and Tekelioglu (1999) also suggested utilization of carrot as potential carotenoid source for colour development in gold fish, C. auratus.

Tlusty (2005) while feeding of American Lobster (Homarus americanus) also found the colouration values on RGB scale were proportional to amount of astaxanthin carotenoid in the diet. Liang et al. (2012) analyzed the red white ornamental koi carp by visual evaluation, photography as well as total carotenoid concentration after feeding with different levels of astacin. Red areas of skin, scale, head and fins showed marked increase in carotenoid content, when fed with 200 and 250 mg kg<sup>-1</sup> of astacin. In addition, highest deposition of carotenoid was observed in red skin followed by scales, red head and fins. Sun et al. (2012) compared the effectiveness of synthetic and natural pigment sources in colouration of Japanese ornamental koi carp (*C. carpio*). Synthetic carotenoid *i.e.* carophyll red @ 1.5 g kg<sup>-1</sup> and natural pigment source *i.e.* Spirulina platensis @ 75 g kg-1 of diet resulted in similar colour improvement. These studies clearly indicate the effectiveness of natural carotenoids similar to synthetic sources in colour enhancement of koi.

The results in terms of CIE colouration were in agreement with the findings of Yeider *et al.* (2014) in

Zebra cichlid (Maylandia estnerae) with spirulina feeding, in which decreased L\* values (lightness) along with significantly increased a\* and b\* values (red and yellow) were observed. Similarly, in the present study also, increased a\* and b\* values resulted in enhanced orange appearance of fish skin. These results are in accordance with the study of Nguyen et al. (2014) on koi carp, in which significantly higher values of a\* and b\* were reported, when fish were provided with synthetic astaxanthin @ 80 mg kg-1 of the diet. Skin pigmentation in terms of total carotenoids (0.9 µg/g) and L\*a\*b\* values were significantly affected by paprika @ 16 % in the diet of pale chub (Zacco platypus). The skin lightness (L\* values) significantly decreased with increased values of a\* and b\* in fish fed diets containing paprika (Lee et al., 2010).

#### **CONCLUSION**

It can be concluded that the carrot meal has a positive effect on growth as well as colour enhancement of koi carp, *Cyprinus carpio* L. in indoor rearing conditions. Hence, carrot meal can be supplemented in koi carp diet at 5 % level both as colour enhancer and growth promoter.

#### **ACKNOWLEDGMENTS**

The authors are thankful to the Dean, College of Fisheries, Guru Angad Dev Veterinary and Animal Sciences University, for providing necessary research facilities and funding for conducting the study.

#### REFERENCES

Ahilan, B., Jegan, K., Felix, N. and Ravaneswaran, K. 2008. Influence of botanical additives on the growth and colouration of adult gold fish (*Carrassicus auratus*) (Linn.). *J. Vet. Anim. Sci.* 4: 129-134.

Amar, E.C., Kiran, V., Satoh, S. and Watanbe T. 2001. Influence of various dietary synthetic carotenoids on bio defense mechanism in rainbow trout (*Onchorhynchs mykiss* Walbaum). *Aquacult. Res.* 32: 162-163.

Arscott, S.A. and Tanumihardjo, S.A. 2010. Carrots of many colour provide basic nutrition and bio available phyto-chemicals acting as a functional food. *Comp. Rev. Food Sci. Food Saf.* 9: 223-239.

AOAC. 2000. *Official Methods of Analysis*. 17<sup>th</sup> edn. Association of Official Analytical Chemists,

- Washungton, D.C.
- APHA. 1991. Standard Methods for the Examination of Water and Wastewater. 18th ed. American Public Health Association, Washungton, DC.
- Baker, R.T.M. 2002. Canthaxanthin in aquafeed application: is there any risk. *Trends in Food Sci. Technol.* 12: 240-243.
- Boonyaratpalin, M. and Lovell, R.T. 1977. Diet preparation of aquarium fish. *Aquacult*. 12: 53-62.
- Chapman, F.A. 2000. Ornamental fish culture freshwater. In: *Encyclopedia of Aquaculture* (R. R. Stickney; Ed.) John Wiley and Sons, New York. pp. 602-610.
- Cyanotech. 2002. Analysis of Beta-carotene and Total carotenoids from spirulina (Spectrophotometric method), *Spirulina pacifica*. Technical Bulletin No. 003b, Cyanotech Corporation. www.cyanotech.com.
- Gupta, S.K., Jha, A.K., Pal, A.K. and Venkateshwarlu, G. 2007. Use of natural carotenoids for pigmentation in fishes. *Nat. Prod. Radiance*. 6: 46-49.
- Halten, B., Arnmesan, A., Jobling M. and Bjerkeng, B. 1997. Carotenoid pigmentation in relation to feed intake, growth and social integration in Arctic char, Salvelinus aipinus (L.), from two anadromous strains. *Aquacult. Nutr.* 3: 189-199.
- Hancz, C., Magyary, I., Molnar, T., Sato. S., Horn, P. and Taniguchi, N. 2003. Evaluation of color intensity enhanced by paprika as feed additive in goldfish and koi carp using computer-assisted image analysis. *Fish. Sci.* 69: 1158-1161.
- Holland, B., Unwin, J.D. and Buss, D.H. 1991. Vegetables, herbs and spices: Fifth supplement to Mc Cance and Widdowson's. The composition of foods London. Royal Society of Chemistry, Cambridge, UK.
- Jain, A.K. 2016. An overview of freshwater ornamental fish production industry in India. pp. 7-20. In: *Best Management Practices for Freshwater Ornamental Fish Production.*, (A. K. Jain, V. P. Saini, and V. I. Kaur; Eds.) National Fisheries Development Board, Department of Animal Husbandry, Dairying and Fisheries, Ministry of Agriculture and Farmers Welfare, Government of India, Hyderabad.
- Kop, A., Durmaz, Y. and Hekimoglu, M. 2010. Effect of Natural Pigment Sources on Colouration. *Advances* 9: 566-569.
- Krinsky, N. 1993. Action of carotenoid in biological system. *Ann. Rev. Nutr.* 13: 561-587.
- Lee, C. R., Pham, M.A. and Lee, S. 2010. Effects of Dietary Paprika and Lipid Levels on Growth and Skin Pigmentation of Pale Chub (*Zacco platypus*). *Asian-Australas. J. Anim. Sci.* 23: 724-732.

- Liang, Y.J., Bai, D.Q., Yang, G., Wei, D., Mei, G., Yan, S.S., Wu, X. and Ning, B. 2012. Effect of astacin on growth and colour formation of juvenile Red-White Ornamental carp (*Cyrinus carpio* var. Koi L). *The Israeli J. Aquacult.-BAMIDGEH* 64: 1-6.
- Mirzaee, S., Beygi, M. M. and Shabani, H. N. A. 2013. Effect of placement carrot (*Daucus carota*) and red pepper (*Capsicum annuum*) in diets on coloration of Jewel Cichlid (*Hemichromis bimaculatus*). World J. Fish. Mar. Sci. 5: 445-448.
- Nguyen, N.V., Khanh, T.V. and Hai, P.D. 2014. Study on Development of Formulated Feed for improving Growth and Pigmentation of Koi carp (*Cyprinus carpio* L. 1758) Juveniles. *J. Life Sci.* 8: 433-441.
- OATA. 2008. Water quality criteria In: Ornamental Aquatic Trade Association. Version 2.0. A Company Limited by Guarantee and Registered in England No 2738119 Registered Office Wessex House, 40 Station Road, Westbury, Wiltshire, BA13 3JN,UK.(http://www.ornamentalfish.org/association/code/Water%20Quality%20Final.pdf).
- Olson, J.A. 1979. A simple dual assay for vitamin A and carotenoids in human liver. *Nutr. Rep. Int.* 19: 807-813.
- Ramamoorthy, K., Bhuvaneswari, S., Sankar, G. and Sakkaravarthi, K. 2010. Proximate composition and Carotenoid Content of Natural Carotenoid Sources and its Colour Enhancement on Marine Ornamental Fish *Amphiprion ocellaris* (Cuvier 1880). *World J. Fish. Mar. Sci.* 2: 545-550.
- Rinna, H. S., Jansi, M. and Vasudhevan, I. 2013. Effect of dietary *Spirulina plantensis* on feeding parameter of blue gourami *Trichogaster trichopterus*. *Int. J. Res. Fish. Aquacult*. 3: 103-106.
- Seyedi, S.M., Sharifpour, I., Ramin, M. and Jamil, S.H. 2013. Effect of Dietary Astaxanthin on survival, growth, pigmentation Clownfish *Amphiprion ocellaris*, Cuvier. *Indian J. Fundam. Appl. Life Sci.* 3: 391-395.
- Sharma, K.D., Karki, S., Thakur, N.S. and Attri, S. 2012. Chemical composition, functional properties and processing of carrot a review. *J. Food Sci. Technol.* 49: 22-32.
- Simon, P. and Wolff, X.Y. 1987. Carotene in tropical and dark orange carrot. *J. Agri. Food Chem.* 135: 1017-1022.
- Sun, X., Chang, Y, Ye., Y, Ma., Z, Liang, Y. L. Li. T., Jiang. N., Xing, W. and Lou, L. 2012. The effect of dietary pigments on the colouration of Japnese ornamental carp (Koi, *Cyprinus carpio* L.). *Aquaculture*. 342: 62-68.
- Thongrod, S., Tansutapanich, A. and Torrissen. 1995. Effects

- of dietary astaxanthin supplementation on accumulation, survival and growth in post larvae of *Penaeus monodon* fabricus, *Bulletin* 24. Larvi 95-Fish and Shellfish larviculture symposium. *Europian Aquacuture Society Specia.*, Belgium.
- Tlusty, M. 2005. Astaxanthin deposition in the cuticle of the lobster. *Marine Biol.* 147: 113-119.
- Torrissen, O.J. and Naevdal, G. 1984. Pigmentation of salmonids-genetical variation in carotenoid deposition in rainbow trout. *Aquaculture*. 38: 59-66.
- Tsushima, M. and Matsuno, T. 1998. Carotenoid composition and two new 4-ketolutein isomer in the integuments of the red filefish, *Branchiostegus japonicas*. *Fish. Sci.* 64: 464-468.

- Wurts, W.A. and Duborow, R.M. 1992. Interactions of pH, carbon dioxide, alkalinity and hardness in fish ponds. *Southern Reg. Aquacult. Centre.* 464: 1-4.
- Yanar, M. and Tekelioglu. 1999. Dogalve Sentetik Karotenoyitlerin Japon Baliklarin in (*Carrasius auratus*) pigmentasyonu Uzerine Etkisi. *Turkish J. Vet. Anim. Sci.* 23: 501-505.
- Yeider, S., Gumus, E., Livengood, E.J. and Chapman, F.A. 2014. The relationship between carotenoid type and skin colour in the ornamental red zebra chichlid Maylandia estherae. *Int. J. Bioflux Soc.* 7: 207-216.
- Zeb, A. 2004. Chemical and nutritional constituents of sea buckthorn juice. *Pak. J. Nutr.* 3: 99-106.

Received on 01-11-2019 and accepted on 27-12-2019



## Effect of Dietary Incorporation of Short Chain and Medium Chain Fatty Acid on Feed Intake and Serum metabolites in Broiler Chickens

Banani Gantayat\*, S.M. Durge, S.A. Amrutkar and V.B. Dongre

Department of Animal Nutrition,

College of Veterinary and Animal Sciences, Udgir-413517, Maharashtra, India

#### **ABSTRACT**

An experiment was conducted to study the effect of incorporation of short chain and medium chain fatty acid (SCMCFA) on feed intake and serum parameters in broiler chickens. Three hundred one-day-old AP-95 strain broiler chicks were reared under uniform managemental conditions. The broiler chicks were individually weighed and distributed randomly into five treatments having four replicates with 15 birds per replicate. Birds were fed either a control diets (C), control plus probiotics (500 mg/kg; P), control plus SCMCFA (1000 mg/kg; F), control plus butyric acid supplement (500 mg/kg; B) or control plus SCMCFA + butyric acid (1000 mg/kg + 500 mg/kg; FB) during pre-starter, starter and finisher phases. The intake followed similar trend in all the groups during the six-week study period, except a significant decline in feed intake in the probiotic fed group during the fifth week. Blood glucose, cholesterol and serum triglycerides were similar among the groups. These results indicated that dietary supplementation of SCMCFA showed no adverse effect on intake, performance and serum metabolites of broiler chicken.

Keywords: Broilers, Feed intake, Serum chemistry, Short chain and medium chain fatty acid

#### **INTRODUCTION**

Emerging concerns on animal ethics, evolving customer demands and government regulations in India has resulted in changing the way of animal rearing, especially the use of antibiotics (Manyi-Loh et al., 2018). An endless number of alternatives are being considered in order to achieve the same feed efficiency and growth rate that are accomplished with antibiotics (Phillips et al., 2004; Den Hartog et al., 2005). Among potential alternatives for antibiotic growth promoters (AGPs), short chain fatty acids (SCFAs) and medium chain fatty acids (MCFAs) act similarly like AGPs (Khatibjoo et al., 2018). Short-chain fatty acids are saturated aliphatic organic acids that consist of one to six carbons, of which acetate  $(C_2)$ , propionate  $(C_3)$ , and butyrate  $(C_4)$  are found to be absorbed from both the small intestine and the ceca by passive transport (den Besten et al., 2013). MCFAs are effective alternatives for antibiotics, due to their high antibacterial activity, and they enter the cell un-dissociated (Khatibjoo et al., 2018). SCFA and MCFA have shown to have synergistic effect and may act by disrupting the cell wall membrane of the microorganisms helping the SCFCA to enter the

cytoplasm where they act (Del Alamo et al., 2007). Therefore, this experiment was carried out to determine the effect of SCFA and MCFA, in comparison with probiotic, butyric acid alone or SC&MCFA with butyric acid, on feed intake and serum parameters in broilers.

#### MATERIALS AND METHODS

The experiment was carried out at the experimental shed of College of Veterinary and Animal Sciences, Udgir, Maharashtra. Three hundred day-old straight run broiler chicks (AP-95) were purchased from a commercial broiler hatchery from Udgir, Maharashtra. The broilers were housed in identical sized pens, environmental temperature was set at 31°C for the first week and 28°C for the second week, which was further decreased to 22°C until the end of the experiment. The relative humidity was above 55% throughout the study. The basal diet was formulated to meet the nutrient requirements of broilers (BIS, 2007). Following a completely randomized design, broilers were allotted to five dietary treatments with four replicates of 15 birds each. The experimental diets consisted of control (C), control plus probiotics (500 mg/kg; P),

Corresponding author Email: bananigantayat@gmail.com

control plus SCMCFA (1000 mg/kg; F), control plus butyric acid t (500 mg/kg; B) and control plus SCMCFA + butyric acid (1000 mg/kg + 500 mg/kg; FB). The supplements were fed during pre-starter, starter and finisher phase as mentioned in Table 1 and 2.

The birds were reared under uniform managemental condition following standard protocols for vaccination, and maintenance of temperature, ventilation and lighting. They were fed experimental diets from 1 to 42 day of age. Feed intake was recorded for each period and during the whole experiment (1–42 d of age). On 42<sup>nd</sup> day, eight birds from each dietary treatment were sacrificed and blood samples were collected, alalyzed for total glucose, serum cholesterol and serum triglycerides content using diagnostic kits. The data obtained were statistically analysed as per one-way ANOVA following method of Snedecor and Cochran (1994) and significant differences between them were tested by Duncan's New Multiple comparison Test according to methods described by SAS (1985). Significance was considered at P<0.05 levels.

#### RESULTS AND DISCUSSION

Data pertaining to the feed intake of the birds are presented in Table 3. Overall, feed intake was not significantly influenced by dietary treatments, except the probiotic supplemented group whose intake was significantly reduced in the 5<sup>th</sup> week. However, Duncan Multiple Range Test (multiple comparison test) revealed that the effect of supplementation did not differ significantly from each other except for the said week. The weekly feed intake of birds fed probiotic diet was comparable with that of 0.05% butyrate supplemented group. In all the treatments, the feed intake followed similar trend and supplementation of SCMCFA had similar intake as that of control. In the fatty acid supplemented groups, the feed intake was marginally improved as compared to other groups. Feed intake was unaltered in broilers fed diets supplemented with medium-chain fatty acids at 0, 1, 2 and 4 g/kg (Saeidi et al., 2016). Khatibjoo et al. (2018) have also reported similar effect of treatments on feed intake during growing period of experiment. The inclusion of dietary

Table 1. Ingredient composition of experimental diets (%)

0 1		` '	
Ingredients (on DM basis)	Pre-starter (0-7 days)	Starter (8-21 days)	Finisher (21-42 days)
Maize	51.12	53.0	57.85
De-oiled soybean cake	42.00	39.5	33.44
Soybean oil, crude	3.00	4.0	5.00
Di-calcium phosphate	1.70	1.8	1.90
Limestone powder	1.00	0.8	0.80
Vitamin premix*	0.025	0.025	0.025
Salt	0.30	0.30	0.30
Methionine	0.17	0.20	0.21
Lysine	0.16	0.14	0.15
Trace mineral mixture** (TMM)	0.10	0.10	0.10
Coccidiostat	0.05	0.05	0.05
Choline	0.06	0.06	0.06
Toxin binder	0.10	0.10	0.10
Total	99.79	100.07	99.99

\*Each kg of vitamin premix contains Vitamin A-22.5 million IU, Vitamin D<sub>3</sub>-4.5 m IU, Vitamin E-60 g, Vitamin K<sub>3</sub>-4 g, Vitamin B<sub>1</sub>-4 g, Vitamin B<sub>2</sub>-20 g, Niacin-60 g, calcium D-panthonate-30 g, Vitamin B<sub>12</sub>-35 g, Folic acid-4 g, Biotin-0.2 g; \*\*Each kg of TMM contains Mn-50 g, Zn-50 g, Fe-55 g, Copper-10 g, Iodine-1.25 g, Selenium-250 mg

Table 2. Nutrient composition of pre-starter, starter and finisher diets

Nutrients			Treatments*		
_	С	P	В	F	FB
Pre-starter diet					
CP (%)	23.0	23.2	23.0	23.0	22.8
EE (%)	5.18	6.58	7.0	5.2	4.41
CF (%)	3.88	4.15	4.2	3.88	4.04
TA (%)	7.45	6.18	6.53	6.78	6.53
AIA (%)	1.6	1.63	1.67	1.62	1.87
ME (kcal)	3002	3006	3007	3005	3003
Starter diet					
CP (%)	22.0	21.8	21.9	21.9	21.9
EE (%)	7.04	6.98	7.5	7.4	7.67
CF (%)	3.97	3.92	3.9	3.94	3.95
TA (%)	7.08	6.8	6.9	6.5	6.4
AIA (%)	1.61	1.61	1.61	1.61	1.61
ME (kcal)	3107	3105	3108	3098	3097
Finisher diet					
CP (%)	20.0	20.1	19.0	19.9	20.0
EE (%)	8.41	8.6	8.7	8.6	8.7
CF (%)	3.2	3.4	3.5	3.4	3.3
TA (%)	6.1	6.0	6.2	6.1	6.3
AIA (%)	1.46	1.5	1.43	1.45	1.46
ME (kcal)	3230	3219	3220	3215	3219

<sup>\*</sup>Treatments: T, control diet; P, probiotic-500 mg/kg; B, butyric acid-500 mg/kg; F, SCMFCA-1000 mg/kg; FB, SCMFCA 1000 mg + butyric acid-500 mg

coconut oil as a source of medium-chain fatty acid by replacing 25, 50, 75 or 100 % of the soybean oil had no adverse effect on feed intake in broilers (Wang *et al.*, 2015). Supplementation of basal diet with 0.1, 0.15 or

0.2 % medium chain fatty acids for period 1-21 day of age, 0.15% for period 21-36 day of age and 0.1% for period 36-42 day of age did not affect feed intake (Mohammadzade *et al.*, 2013). Pinchasov and Jensen

Table 3. Effect of dietary supplementation of probiotics and SCMCFA on weekly feed intake (g)

Age			$Treatments^*$			P-value
in weeks	C	P	В	F	FB	
1	129.7±7.45	129.1±2.83	134.6±6.60	119.9±2.33	111.6±7.43	0.080
2	409.0±38.89	359.4±19.70	410.0±18.78	432.1±35.70	337.4±6.95	0.120
3	569.3±54.91	563.0±46.25	585.8±7.11	574.8±14.2	579.6±26.53	0.980
4	900.6±25.73	892.4±9.69	948.5±18.44	957.8±18.77	872.0±43.97	0.140
5	1102±23.63ab	1025±12.42a	1076±13.01ab	1145±20.52 <sup>b</sup>	$1124 \pm 17.8^{ab}$	0.002
6	874.4±35.11	876.5±19.75	909.6±54.19	900.3±13.3	877.1±4.56	0.88

<sup>&</sup>lt;sup>a,b</sup> Means bearing different superscripts within the same row are significantly different (P<0.05); \*Treatments: T, control diet; P, probiotic-500 mg/kg; B, butyric acid-500 mg/kg; F, SCMFCA-1000 mg/kg; FB, SCMFCA 1000 mg + butyric acid-500 mg

Table 4. Effect of dietary supplementation of probiotics and SCMCFA on serum profile

Parameters	Treatments*							
	С	P	В	F	FB			
Blood glucose (mg/dl)	128±23.5	125±29.5	156±7.0	222±14.0	139±24.0	0.64		
Serum cholesterol (mg/d)	163±12.0	139±15.0	149±11.0	178±8.0	176±34.0	0.55		
Triglycerides (mg/dl)	97.5±1.5	80±18.0	70.5±1.5	130±13.5	133±27.5	0.12		

\*Treatments: T, control diet; P, probiotic-500 mg/kg; B, butyric acid-500 mg/kg; F, SCMFCA-1000 mg/kg; FB, SCMFCA 1000 mg + butyric acid-500 mg/kg; B, butyric acid-500 mg/kg; F, SCMFCA-1000 mg/kg; FB, SCMFCA 1000 mg + butyric acid-500 mg

(1989) summarized that in case of well-nourished healthy chicks housed at a moderate stocking density and hygienic condition, dietary inclusion of MCFAs or SCFAs had no significant influence on the birds' performance. In an another study on MCFA, Miller *et al.* (2009) reported no differences with regard to feed intake, feed efficiency and weight gain in grower and finisher pigs when fed diets with 1, 3, or 6% of MCFA oil compared with diets containing the same amount of tallow, pig fat or corn oil. However, in our study birds were fed 1000 mg/kg of SCMCFA in feed and we did not observe any effect of supplementation on feed intake.

Serum concentration of glucose, cholesterol and triglycerides were similar among the groups (Table 4). In another study it was observed that the serum triglyceride concentrations was significantly increased in broilers with MCFA diet as compared to control (Baltic *et al.*, 2018). The present findings corroborate well with that Khatibjoo *et al.* (2018) who bserved no effects of supplemental dietary short-and medium-chain fatty acids on serum cholesterol and glucose of broiler chickens.

#### CONCLUSION

Our results suggest that combination of short chain and medium chain fatty acid supplementation in broiler diet to be beneficial and exert no adverse effect on feed intake and blood biochemistry.

#### REFERENCES

Baltic, B., Ciric, J., Šefer, D., Radovanovic, A., Đorðevic,
J., Glišic, M., Boškovic, M., Baltic, M.Z, Đorðevic,
V. and Markovic, R. 2018. Effect of dietary supplementation with medium chain fatty acids on growth performance, intestinal histomorphology, lipid profile and intestinal microflora of broiler chickens.

South Afr. J. Anim. Sci. 48: 885-896.

BIS. 2007. *Indian Standards of Poultry Feed Specification*, 5<sup>th</sup> rev. edn. IS:1374, Bureau of Indian Standards, Manak Bhavan, New Delhi, India.

Del Alamo, A.G., De Los Mozos, J., Van Dam, J.T.P. and De Ayala, P.P. 2007. The use of short and medium chain fatty acids as an alternative to antibiotic growth promoters in broilers infected with malabsorption syndrome. In: *Proc. the 16<sup>th</sup> European Symposium on Poultry Nutrition*, pp. 317-320.

Den Besten, G., Van Eunen, K., Groen, A.K., Venema, K., Reijngoud, D.J. and Bakker, B.M. 2013. The role of short-chain fatty acids in the interplay between diet, gut microbiota, and host energy metabolism. *J. Lipid Res.* 54: 2325-2340.

Den Hartog, L.A., Gutierrez Del Alamo, A., DoorenBos, J. and Flores, A. 2005. The effect of natural alternatives for anti-microbial growth promoters in broiler diets. In: *Proceedings of the 15<sup>th</sup> European Symposium on Poultry Nutrition*, Balatonfüred, Hungary.

Khatibjoo, A., Mahmoodi, M., Fattahnia, F., Akbari-Gharaei, M., Shokri, A.N. and Soltani, S. 2018. Effects of dietary short-and medium-chain fatty acids on performance, carcass traits, jejunum morphology, and serum parameters of broiler chickens. *J. Appl. Anim. Res.* 46:492-498.

Manyi-Loh, C., Mamphweli, S., Meyer, E. and Okoh, A. 2018. Antibiotic use in agriculture and its consequential resistance in environmental sources: potential public health implications. *Molecules*. 23: 795.

Miller, D., Johnston, Z.C., Mullan, B., Pluske, J.R. and Hansen,
C. 2010. Nutritional manipulation of the somatotropic axis in grower and finisher pigs 2
F-106. Report prepared for the Co-operative Research
Centre for an Internationally Competitive Pork Industry.

Mohammadzade, S., Shahriar, H.A., Ebrahimnejad, Y., Ahmadzadeh, A. and Tahmasebpour, B. 2013. Effect of different levels of medium chain fatty acids on

- performance, and some of microbial population of gastro in broiler chicks. *Res. J. Chem. Environ. Sci.* 1:5-7.
- Phillips, I., Casewell, M., Cox, T., De Groot, B., Friis, C., Jones, R., Nightingale, C., Preston, R. and Waddell, J. 2004. Does the use of antibiotics in food animals pose a risk to human health? A critical review of published data. *J. Antimicrobial. Chemother.* 53: 28-52.
- Pinchasov, Y. and Jensen, L.S. 1989. Effect of short chain fatty acids on voluntary feed intake of broiler chicks. *Poult. Sci.* 68: 1612-1618.
- Saeidi, E., Shokrollahi, B., Karimi, K. and Amiri-Andi, M. 2016. Effects of medium-chain fatty acids on performance, carcass characteristics, blood biochemical parameters and immune response in Japanese quail. *Br. Poult. Sci.* 57: 358-363.
- Snedecor, G. W. and Cochran, W. G. 1994. *Statistical Methods* (eighth edition). Oxford & IBH Publishing Co., Calcutta, India.
- Wang, J., Wang, X., Li, J., Chen, Y., Yang, W. and Zhang, L. 2015. Effects of dietary coconut oil as a medium-chain fatty acid source on performance, carcass composition and serum lipids in male broilers. *Asian-Australas. J. Anim. Sci.* 28: 223-230.

Received on 04-11-2019 and accepted on 15-12-2019



## Indian J. Anim. Nutr. 2019. 36 (4): 419-422 doi: 10.5958/2231-6744.2019.00068.9

#### **SHORT COMMUNICATION**

### Successful Management of Nitrite Poisoning in Crossbred Dairy Calves

E. Niyas¹\*, Shibu Simon², Surej Joseph Banglavan³, Jith John Mathew⁴, Ani S. Das⁵, Reni John⁶ and S. Reshma⁻

University Livestock Farm and Fodder Research Scheme, College of Veterinary and Animal Sciences, Veterinary and Animal Sciences University, Mannuthy, Thrissur, Kerala, India

#### **ABSTRACT**

Nitrate/ nitrite poisoning in dairy cattle occurs when there is an abrupt change in diet from the introduction of a bulk feed source with high nitrate levels. Over fertilization of fodder with untreated manure and occurrence of drought season followed by a rainy season could result in higher level of accumulation of nitrate in plants. A sudden onset of illness was reported in eight crossbred dairy calves (between 8-11 months of age) at University Livestock Farm and one among the affected calf died after showing symptoms of discomfort, excessive salivation, tympany, dyspnoea, sternal recumbency progressing to lateral recumbency, pedalling movements and, stiffness of the limbs and neck. On clinical examination, the affected animals exhibited sialorrhoea, tympany, increased pulse and respiratory rates, laboured breathing, brownish discoloration of mucous membrane and loss of co-ordination. Microscopical examination of rumen fluid revealed normal protozoan motility. Blood colour was dark chocolate brown in all the affected animals and coagulation time was delayed. Animals had a history of ad libitum feeding with chopped lush green fodder grasses namely hybrid Napier and fodder maize that were harvested from cattle slurry irrigated fodder plots. Based on the history and clinical signs noticed, the case was presumptively diagnosed as nitrate/nitrite poisoning, later confirmed with diphenylamine test. All the animals were treated with intravenous administration of 1 per cent methylene blue @ 1-2 mg/kg BW. Subsequently the dairy calves showed an uneventful recovery from the condition.

Key words: Methylene blue, Nitrate, Nitrite, Sodium thiosulphate

Nitrate/ Nitrite poisoning is the one of the most widespread toxic condition in ruminants. Fodder crops namely maize, sorghum, brassicas, Italian rye grass, tama and green oats are considered as the principal accumulators of nitrate that could reach toxic levels for bovines. Accumulation of nitrate is more in rapid growing plants with reduced photosynthesis, over fertilization of the soil with excessive use of animal manure and poultry litter are the principle cause of nitrate accumulation in forages (Follet and Hatfield, 2001) and the accumulation is more in young and rapidly growing plants as compared to the mature forages (Haliburton and Edward, 1978).

The occurrence of a rainy season followed by a draught period, less adaptation of the ruminants to a

nitrite rich diet and *ad libitum* feeding of bovines with nitrite rich plants have increased the risk of nitrate toxicity in ruminants (Gontigo *et al.*, 2017). Ruminants like cattle, goat and sheep are more prone to nitrate toxicity as the rumen bacteria reduce nitrate to a more toxic nitrite and when this conversion ratio is faster than the conversion of nitrite to ammonia, the excess nitrite that accumulates in the rumen is progressively absorbed into the blood stream. Nitrite in the blood reduces the oxygen carrying haemoglobin to a brownish black substance called methemoglobin that cannot carry oxygen and imparts the above colour to the blood (Osweiler 1996). Nitrite also causes vasodilatation that leads to a reduction in blood pressure and circulatory shock and foetal death in pregnant animals due to

<sup>&</sup>lt;sup>1</sup>\*Corresponding author Email: drniyasemad@gmail.com; Research Assistant, University Livestock Farm and Fodder Research Development Scheme, College of Veterinary and Animal Sciences, Mannuthy, Thrissur (Kerala), <sup>2,3 & 4</sup>Assistant Professor, College of Veterinary and Animal Sciences, Mannuthy, Thrissur (Kerala), <sup>5</sup>PhD Scholar, College of Veterinary and Animal Sciences, Mannuthy, Thrissur (Kerala), <sup>6</sup>Research Assistant, University Livestock Farm and Fodder Research Development Scheme, College of Veterinary and Animal Sciences, Thrissur (Kerala), <sup>7</sup>MVSc Scholar, College of Veterinary and Animal Sciences, Mannuthy, Thrissur (Kerala)

hypoxia. Affected animals show clinical signs when 20 per cent of haemoglobin is converted to methemoglobin and death results when it reaches to 60-80 per cent (Jones 1988). Clinical signs include chocolate brown discoloration of the mucous membranes, severe abdominal pain, ataxia, incordination, muscle tremor, weakness, severe dyspnoea, cyanosis, collapse and recumbency. In this paper we report a case of spontaneous occurrence of nitrate or nitrite intoxication in cross bred dairy heifers and its' successful management.

A sudden onset of illness was reported in eight crossbred dairy calves (8-11 months of age) in University Livestock Farm, Mannuthy and one among the affected calves died after showing symptoms like sudden onset of discomfort, excessive salivation, tympany, dyspnoea, sternal to lateral recumbency, pedalling movements and stiffness of the limbs and neck. It was reported that all calves were healthy and they were fed *ad libitum* chopped lush green fodder (hybrid Napier and fodder maize) harvested from cattle slurry irrigated fodder plots.

Detailed clinical examination was carried out on all the affected animals and observations were recorded. The affected animals exhibited sialorrhoea, ataxia, tachycardia, tachypnoea, dyspnoea, cyanotic mucous membrane and loss of balance. Microscopical examination of rumen fluid revealed normal protozoan motility. Blood colour was found to be dark chocolate brown in all affected animals.

Based on the feeding history and clinical signs noticed, the case was presumptively diagnosed as nitrite poisoning and treatment was carried out with 1 % methylene blue. The diphenylamine test was performed in grass samples by adding one drop of the reagent solution (0.5 g diphenylamine, 20 ml distilled water, and 80 ml concentrated sulfuric acid) to three drops of the plant extract on glass slides. The extract was obtained by manual pressure (Tokarnia et al., 2012). The results of the diphenylamine test associated with clinical signs and epidemiological data helped to conclude the diagnosis of nitrate/nitrite intoxication. All the affected animals were treated with intravenous administration of 1% methylene blue @ 1-2 mg/kg BW. Subsequently the dairy calves showed an uneventful recovery from the condition.

Even though the chopped lush green grass was fed to animals in all the age group the intoxication occurred only in the heifers group, whose diet was almost exclusively composed of fodder that corresponded to approximately 60-80% of the total dry matter intake. Dairy cows, on the other hand received larger amounts of concentrate, and the calves received milk and ate a small amount of concentrates and grass. Hybrid Napier and maize fodders are good accumulator nitrates (Medeiros *et al.*, 2003). The condition was further aggravated the fact that the fodder were harvested from plots that were fertilized with large amount of untreated manure and pig slurry. Additionally, the site experienced a long period of drought



Fig. 1 Brownish discoloration of the blood (A), mucus membrane of vulva (B) and rectum (C)

without rain followed by high rainfall during the incident. All these factors might have contributed to the accumulation of nitrate in the fodders offered to the animals and bulk consumption of these fodder caused the intoxication.

In ruminants, nitrate/nitrite toxicity occurs when high nitrate levels in the feed overwhelm the animal's digestive system to the extent that the rate of conversion of nitrate to nitrite is faster than the conversion of nitrite to ammonia (Al Qudah, 2009). Microbes in the rumen reduce nitrate contents in the ingested plants in to nitrite, which is five to six times more toxic than nitrates (Rogers and Hope-Cawdery, 1980). Following its absorption into blood through rumen capillaries, nitrite oxidizes the haemoglobin to methaemoglobin which is unable to transport oxygen and also results in vasodilatation (Leng, 2008). These two predominant effects of nitrite lead to tissue hypoxia and circulatory collapse (Haymond et al., 2005). The amount of nitrate ingested by the animal and methemoglobin formed influences the onset of clinical signs, disease progression and death (Jonk et al. 2013). The amount of methemoglobin formed in each animal depends on its nitrate recycling rates in the rumen and on the drop in nitrite levels, reflecting different levels of nitrate tolerance presented by the animals (Ozmen et al., 2005).

The poisoned heifers exhibited a cyanotic or brownish discoloration of mucosa of conjunctiva, vaginal vestibule and rectum; dyspnoea, sialorrhoea, tympany, and progression to sternal recumbence and death. Similar findings were also reported by Medeiros *et al.* (2003) and Jonck *et al.* (2013). Among the intoxicated animals one heifer died after showing symptoms like stiffness in limbs, neck and jaw, tympany and pedalling movements. Necropsy was performed soon after the animal's death. A strong nitrous odour was noticed while opening the carcass. The blood was dark chocolate brown in colour with difficult coagulation, all the compartments of stomach were distended due to gas accumulation and a brownish discoloration was observed in lungs, kidneys and liver.

These necropsy findings were in accordance with the report of Tokarnia *et al.* (2012) who noticed brownish discolouration of visceral organs in intoxicated animals. Chocolate-coloured and poorly clotted blood and intensely red coloured skeletal and cardiac musculature as observed in the present case were compatible with those described by Jonck *et al.* (2013).

The drug of choice for nitrate/nitrite poisoning is intravenous administration of 1% methylene blue at the dosage of 1 to 2 mg/kg body weight. Methylene blue acts by reducing Fe³+ to Fe²+, and receiving an H⁺ from NADPH (becoming leukotolene blue) and donating it to methemoglobin, which is converted to haemoglobin. Since the time taken to metabolize the methylene blue is short, the poisoned animal shows recovery within minutes after its administration. The rapid response to the treatment is another form of diagnosis of the intoxication (Radostits *et al.*, 2007; Jonck *et al.*, 2013).

The clinical signs, necropsy findings, epidemiological data and the positive diphenylamine test confirmed the nitrate/nitrite intoxication. The sudden change in diet, with introduction of a bulky food source containing high nitrate levels, the over fertilization of grass with untreated manure, and the occurrence of a drought period followed by a rainy season were some of the causes of the intoxication. Intra venous administration of one percent methylene blue @ 1-2 mg/kg body weight resulted in uneventful recovery of the animals.

#### REFERENCES

Al-Qudah, K.M., Rousan, L.M. and Ereifej, K.I. 2009. Nitrate/nitrite poisoning in dairy cattle associated with consumption of forages irrigated with municipally treated wastewater. *Toxicol. Environ. Chem.* 91: 163-170.

Haliburton, J.C. and Edwards W.C. 1978. Nitrate poisoning in oklahoma cattle during winter of 1977-1978. *Vet. Hum. Toxicol.* 20: 401-403.

Follett, R.F. and Hatfield, J.L. 2001. Nitrogen in the environment: sources, problems, and management. *Sci. World J.* 1: 920-926.

Gontijo, D.A., Borges, A.A and Wouters, F. 2017. Nitrate/ nitrite poisoning in dairy cattle from the Midwestern Minas Gerais, Brazil. *Ciência Rural*. 12: 1-7.

- Haymond, S., Cariappa, R., Eby, C.S. and Scott, M.G. 2005. Laboratory assessment of oxygenation in methemoglobinemia. *Clin. Chem.* 51: 434-444.
- Jonck, F., Gava, A., Traverso, S.D., Lucioli, J., Furlan, F.H. and Gueller, E. 2013. Spontaneous and experimental poisoning by nitrate/nitrite in cattle fed Avena sativa (oat) and/or Lolium spp. (ryegrass). *Pesquisa Veterinária Brasileira*. 33: 1062-1070.
- Jones, T.O.1988. Nitrate/nitrite poisoning in cattle. *In Practice*. 10: 199-203.
- Leng, R.A. 2008. The potential of feeding nitrate to reduce enteric methane production in ruminants. A Report to the Department of Climate Change, Commonwealth Government of Australia, Canberra. 82p.
- Medeiros, R.S., Lopez, B.A., Sampaio, L.A., Romano, L.A. and Rodrigues, R.V. 2016. Ammonia and nitrite toxicity to false clownfish Amphiprion ocellaris. *Aquacult. Int.* 24: 985-993.

- Osweiler, G. 1996. Methemoglobinemia. In: *Toxicology*, ed. G.D. Osweiler, 398-400. Philadelphia, PA: Williams and Wilkins.
- Ozmen, O., Mor, F., Sahinduran, S. and Unsal, A. 2005. Pathological and toxicological investigations of chronic nitrate poisoning in cattle. *Toxicol. Environ. Chem.* 87: 99-106.
- Radostits, O.M., Gay, C.C., Hinchcliff, K.W. and Constable, P.D. 2007. *Veterinary Medicine, A Text Book of the Disease of Cattle, Horses, Sheep, Pigs and Goats.* (10<sup>th</sup> edn), Saunders, Elsevier, London pp 1851-1895.
- Rogers, P.A.M. and Hope-Cawdery, M.J. 1980. Monensin, ketosis and nitrate toxicity in cows. *Vet. Rec.* 106: 311-312.
- Tokarnia, C.H., Brito, M.F, Barbosa, J.D, Peixoto, P.V. and Dobereiner, J. 2012. *Plantas Tóxicas do Brasil para Animais de Produção*. 2. ed. Rio de Janeiro: Helianthus, 2012. 586p.

Received on 21-11-2019 and accepted on 28-12-2019



Indian J. Anim. Nutr. 2019. 36 (4): 423-427 doi: 10.5958/2231-6744.2019.00069.0

#### **SHORT COMMUNICATION**

# In Sacco Degradability Kinetics of Cumbu Napier (COBN-5) Fodder at Two Different Stages of Harvest

M. Madesh, K. Raja Kishore<sup>1\*</sup>, D. Srinivas Kumar and A. Anitha

Department of Animal Nutrition, Sri Venkateswara Veterinary University NTR College of Veterinary Science, Gannavarm-521102, AP, India

#### **ABSTRACT**

In the present study, Cumbu Napier (COBN-5) fodder was evaluated for nutrient degradability *in sacco* using three rumen fistulated Murrah buffalo bulls (380 ± 9.36 kg BW) fed *ad libitum* COBN-5 fodder at two different stages (40 and 45 days) of harvest. The average *in sacco* disappearance values (%) of DM, CP, NDF and ADF were 80.49, 91.97, 74.11 and 73.79 after 72 h incubation in the rumen, respectively. The readily soluble fraction (a) for DM, CP, NDF and ADF was higher in the fodder harvested at 40 days, whereas, higher insoluble but degradable fraction (b) for DM, CP, NDF and ADF was reported in the fodder harvested at 45 days. Further, potentially degradable fraction, PD (a+b) for DM and CP was lower, and for NDF and ADF was higher in the fodder harvested at 40 days. The effective degradability (%) of DM, CP, NDF and ADF was higher in the fodder harvested at 40 days than in 45 days. Results indicated that COBN-5 fodder can be effectively included in the rations of livestock harvested at 40 days for effective utilization.

Keywords: COBN-5 fodder, Degradation kinetics, In sacco degradability, Stage of harvest

The ruminant sector provides a significant proportion of self-employment opportunities and forms a basis for supplementary income to most sections of India's agrarian society. Forages usually constitute the major portion of the ruminant feeds and high quality nutritious green fodder is required to exploit the productive potential of dairy animals. With the increase in industrialization, ever-increasing population pressure of human beings, there is a little chance of having good arable land available for cultivation of green fodder. To meet the current level of livestock production, cultivation of perennial fodders which can yield higher biomass per unit area with early harvesting period is the immediate solution.

A number of breeding programmes came into force for evolving best quality green fodder. Hybrid Napier varieties proved to be promising fodders for their high yielding capability and usage as forage for livestock (Woodard and Prine, 1991). Cumbu Napier (COBN-5) fodder is a newly developed hybrid Napier variety by Tamil Nadu Agricultural University (TNAU) at Coimbatore and released for commercial cultivation

in 2013 (Babu et al., 2014). The yield data from research station trials showed a higher fodder yield in COBN-5 (3260 g/ha/yr) compared to CO-3 (2960 q/ha/yr). A key measure of the nutritive value of feedstuffs is digestibility, either in vitro or in vivo (Minson, 1990). Data on chemical composition, ruminal degradation and their relationships determine the economic value of forage (Weiss, 1994). The time of harvest also effects the chemical composition and nutrient degradability of forage inside the rumen. It is very important to know the proportion of dry matter or protein or fibre that is degraded in the rumen in certain times (Topps, 1996). The in sacco degradability is the best available method for estimating digestibility similar to in vitro disappearance and it is easy to perform (Weiss 1994). Therefore, the present study was undertaken to ascertain the in sacco degradability of DM, CP, NDF and ADF in COBN-5 fodder at two different stages of harvest.

Cumbu Napier (COBN-5) fodder was procured from the cultivated fields of Livestock Farm Complex (LFC), NTR College of Veterinary Science,

\*Corresponding author Email: dr\_rajakishore@yahoo.co.in; Present address: Department of Animal Nutrition, NTR College of Veterinary Science, Gannavaram-521102, Andhra Pradesh, India.

Gannavaram. First cut was done after 75 days of plantation. Subsequently, the fodder was harvested on 30, 35, 40, 45 and 50 days in the second cut and is used for nutritional evaluation. Chemical composition (AOAC, 2007) and forage fibre constituents (Van Soest *et al.*, 1991) of the fodder were evaluated. Further, *in sacco* degradability of DM, CP, NDF and ADF was estimated in the rumen (Orskov and Mc Donald, 1979) using COBN-5 fodder harvested at 40 and 45 days.

Three adult rumen fistulated Murrah buffalo bulls  $(380 \pm 9.36 \text{ kg BW})$  maintained on COBN-5 fodder were used to determine the *in sacco* degradability of DM, CP, NDF and ADF in COBN-5 fodder harvested at 40 and 45 days. The COBN-5 fodder was chaffed, dried and ground in a Wiley mill (2 mm size mesh screen) for incubation in the rumen. The pre-weighed nylon bags (13.5 cm x 7.5 cm), containing 5 g of feed were incubated for 0, 3, 6, 12, 24, 48 and 72 h in the ventral sac of rumen. The bags containing samples for 0 h were washed without incubation in rumen. The bags with feed samples incubated in the rumen were drawn in a sequential removal method (Osuji *et al.*, 1993). After

removal, the bags were washed under slow running tap water by rubbing between fingers and the thumb for ten minutes, oven dried at 70°C for 48 h to constant weight and DM loss during the incubation was calculated. The CP, NDF and ADF content in the residue was analyzed to determine their respective degradabilities. From the degradability data obtained at different intervals, the constant a, b and c from the expression P=a+b (1-e<sup>-ct</sup>) were obtained. Where, P is the degradability at time t, the constant, a is the intercept or instantly degradable fraction, b is the potentially degradable and c is the degradation rate or rate constant. The effective nutrient degradability (%) of crop residues, conventional and complete rations were calculated by time measurements and fitted values in NEWAY programme (Mc Donald, 1981) using a computer, assuming an outflow rate (K) of 0.05%/hr. The data was analyzed statistically (Snedecor and Cochran, 1994) using SPSS 17.0 version.

The chemical composition and cell wall constituents of COBN-5 fodder harvested at different days is presented in Table 1. Data revealed that the

Table 1. Chemical composition (% DM basis) of COBN-5 fodder at different days of harvest

Nutrient			Days of	f harvest		
_	30	35	40	45	50	Mean ± SE
Dry matter	16.68a	17.02a	17.42ab	18.03 <sup>b</sup>	20.54°	17.94±1.09
Organic matter	$82.49^{a}$	84.27 <sup>b</sup>	86.05°	$87.20^{cd}$	88.17 <sup>d</sup>	85.64±1.61
Total ash	17.51 a	15.73 b	13.95°	12.80 <sup>d</sup>	11.83 e	14.36±1.61
Crude protein	$14.00^{a}$	13.41 <sup>b</sup>	$13.02^{\circ}$	$12.70^{d}$	12.10 <sup>e</sup>	13.05±0.51
Ether extract	$3.27^{a}$	$2.90^{b}$	$2.82^{b}$	2.72°	$2.41^{d}$	$2.82 \pm 0.22$
Crude fibre	19.24 <sup>a</sup>	$23.07^{b}$	$27.96^{\circ}$	$30.50^{d}$	34.69e	27.09±4.30
Nitrogen free extract	45.98 a	44.89 <sup>b</sup>	42.25 °	$41.28^{d}$	38.97 e	42.67±1.99
Neutral detergent fibre	$62.00^{a}$	$65.00^{b}$	67.06°	$69.10^{d}$	73.38e	67.31±3.03
Acid detergent fibre	$38.29^{a}$	39.26 <sup>b</sup>	$40.20^{\circ}$	41.64 <sup>d</sup>	$44.80^{e}$	$40.84 \pm 2.41$
Hemi-cellulose	23.71 <sup>a</sup>	25.74 <sup>b</sup>	$26.86^{\circ}$	$27.46^{d}$	28.58e	26.47±0.44
Cellulose	30.94 a	32.61 b	34.25 °	$36.70^{d}$	40.87 e	35.07±2.97
Acid detergent lignin	4.97°	5.31 b	5.68°	6.13 <sup>d</sup>	6.54 e	5.73±2.74
Silica	2.95 a	3.17 b	3.42°	$3.69^{d}$	3.82 e	3.41±0.25
Calcium (%)	$0.74^{\mathrm{a}}$	0.81 a	1.15 b	$0.48^{\mathrm{b}}$	0.47°	$0.73\pm0.20$
Phosphorus (%)	0.68 a	0.51 a	0.75 a	0.45 b	$0.44^{\rm b}$	$0.57 \pm 0.10$

Each value is a mean of 3 observations; a,b,c,d,e Values in the rows bearing different superscripts differ significantly (P<0.05)

protein content was higher, while NDF, ADF and lignin were lower in Cumbu Napier fodder at 40 days of harvest as compared to 45 days. Forage age is the most influential factor on animal response parameters when forages form the main source of diet for ruminants. The fodder harvested at 40 and 45 days is studied *in sacco* by incubating in the rumen upto 72 h. The average *in sacco* disappearance values (%) and degradation kinetics of DM, CP, NDF and ADF in COBN-5 fodder is presented in Tables 2 and 3.

Data revealed that as the period of incubation extended from 0 to 72 h in the rumen, the *in sacco* disappearance (%) values of DM, CP, NDF and ADF increased progressively and higher values were observed in the fodder harvested at 40 days compared to that harvested at 45 days with non-significant differences. The rapidly soluble fraction (a) was higher, while the insoluble but degradable fraction (b), potentially degradable fraction (a + b) and rate constant/h (c) were lower in DM in the fodder harvested at 40 days (Table 2). Further, ED (0.05%-h) of DM was higher in the fodder harvested at 40 days. Similar findings were

reported by other research workers (Abdul Razak et al., 1996; Tessema and Baars. 2004) in Napier grass. The rapidly soluble fraction 'a' and ED (0.05%-h) of CP was higher in fodder harvested at 40 days, while the insoluble but degradable fraction 'b' was higher in the fodder harvested at 45 days (Table 2). Further, decreases in protein content was caused by an increase in stem proportion (Gutteridge and Shelton, 1994), which had a lower protein percentage than leaves, and there will be a reduction in leaf and stem fraction with age. Muia et al. (2001) reported similar values for fractions 'b' and 'c' but lower values for 'a' and ED of CP in Napier fodder compared to the values in present study. The higher fraction 'a' and lower 'b' fraction in the present study indicates presence of more soluble nitrogen content as compared to other Napier varieties. The effective degradability of CP was higher than the other Napier grass variety suggesting high degradable nitrogen content (Kabi et al., 2005) in COBN-5 fodder. Higher fraction 'a', (a+b) and ED (0.05%-h) of NDF were reported in the fodder harvested at 40 days compared to fodder harvested at 45 days. The lower

Table 2. Average *in sacco* disappearance (%) and degradation kinetics of DM and CP in COBN-5 fodder at 40 and 45 days of harvest

Parameter		DM			CP	
Days of harvest	40	45	Mean ± SE	40	45	Mean ± SE
<b>Incubation period</b> (h	n)					
0	38.37	37.04	37.70±0.67	55.18 <sup>a</sup>	53.47 <sup>b</sup>	54.33±0.86
3	40.47	39.20	39.83±0.64	$62.99^{a}$	60.49 <sup>b</sup>	61.74±1.25
6	48.00	46.97	47.49±0.52	$69.74^{a}$	67.84 <sup>b</sup>	68.79±0.95
12	57.23	51.87	54.55±2.68	$73.85^{a}$	71.73 <sup>b</sup>	72.79±1.06
24	70.05	67.49	68.77±1.28	$79.05^{a}$	75.41 <sup>b</sup>	77.23±1.82
48	77.46	72.55	75.05±2.45	88.73 a	86.12 <sup>b</sup>	87.43±1.31
72	82.54	78.44	80.49±2.05	92.10a	91.84 b	91.97±0.13
<b>Degradation Kinetic</b>	cs					
a	38.37	37.04	37.70±0.67	55.18 <sup>a</sup>	53.47 <sup>b</sup>	54.33±0.86
b	43.98	50.27	47.12±3.15	39.33a	44.56 <sup>b</sup>	41.95±2.62
c	0.056	0.024	$0.04 \pm 0.02$	$0.036^{\mathrm{a}}$	$0.024^{b}$	$0.030\pm0.01$
PD (a+b)	82.35	87.31	84.83±2.48	94.51a	98.03 <sup>b</sup>	96.27±1.76
ED (0.05%/h)	58.50	54.20	56.35±2.15	$76.10^{a}$	$74.80^{b}$	75.45±0.65

Each value is a mean of 3 observations; a.bValues in the rows bearing different superscripts differ significantly P<0.05

Table 3. Average *in sacco* disappearance (%) and degradation kinetics of NDF and ADF in COBN-5 fodder at 40 and 45 days of harvest

Parameter		NDF			ADF	
Days of harvest	40	45	Mean ± SE	40	45	Mean ± SE
<b>Incubation period</b> (	h)					
0	25.34 a	$18.98^{b}$	22.16±3.18	19.11 a	14.46 <sup>b</sup>	$16.79 \pm 2.32$
3	30.16 <sup>a</sup>	24.36 <sup>b</sup>	27.26±2.90	25.39a	19.14 <sup>b</sup>	22.27±3.13
6	36.20a	$32.05^{b}$	34.13±2.08	$33.77^{a}$	26.74 <sup>b</sup>	$30.26 \pm 3.52$
12	44.98a	42.64 <sup>b</sup>	43.81±1.17	44.69a	38.46 <sup>b</sup>	41.58±3.11
24	63.13a	53.78 <sup>b</sup>	58.46±4.68	61.01a	52.96 <sup>b</sup>	$56.99 \pm 4.02$
48	71.54 <sup>a</sup>	62.08 <sup>b</sup>	66.81±4.73	71.42a	63.24 <sup>b</sup>	67.33±4.09
72	76.54 <sup>a</sup>	71.68 <sup>b</sup>	74.11±2.43	76.52a	71.06 <sup>b</sup>	73.79±2.73
<b>Degradation Kineti</b>	cs					
a	25.34a	18.98 <sup>b</sup>	22.16±3.18	19.11 <sup>a</sup>	14.46 <sup>b</sup>	16.79±2.32
b	52.41a	52.48a	52.45±0.04	58.11 <sup>a</sup>	57.05 <sup>b</sup>	57.58±0.53
c	$0.0501^{a}$	$0.0449^{b}$	$0.048 \pm 0.001$	$0.0530^{a}$	$0.0478^{b}$	$0.05\pm0.002$
PD (a+b)	77.75 <sup>a</sup>	71.46 <sup>b</sup>	74.61±3.15	77.22a	71.51 b	74.37±2.86
ED (0.05%/h)	$48.90^{\mathrm{a}}$	43.10 <sup>b</sup>	46.00±2.90a	46.74 <sup>a</sup>	39.99 <sup>b</sup>	43.37±3.38

Each value is a mean of 3 observations; a.bValues in the rows bearing different superscripts differ significantly P<0.05

ED (%) of NDF in the fodder harvested at 45 days might be due to higher cellulose and acid detergent lignin content (Table 3). The potentially degradable fraction of elephant grass was reported to be lower than the value in the present study (Gallardo et al., 2010) in Napier grass. The rapidly soluble fraction (a), insoluble but degradable fraction (b) and PD (a+b) of ADF were higher in the fodder harvested at 40 days. Further, the effective degradability of ADF was higher in the fodder harvested at 40 days (Table 3). Lower potential degradability values were reported for elephant grass than the values observed in the present study (Gallardo et al., 2010; Singh et al., 1992). Decreased ADF degradability in late cut forages, may be attributed to presence of higher lignified fibre. Further, as forage matures its content of less digestible nutrients decreases (Bosch et al, 1992).

It was concluded that the COBN-5 fodder was better utilized in terms of *in sacco* nutrient degradability harvested at 40 days as compared to 45 days. This, will help to use the fodder for animal feeding at early stages of harvest, minimizing the cost of cultivation practices.

Hence, the fodder can be better utilized by including it in the rations of livestock starting at 40 days of harvest for economical feeding.

#### REFERENCES

Abdul Razak, S.A., Muinga, R.W., Thorpe, W. and Orskov, E.R. 1996. The effects of supplementation with *Gliricidia sepium* or *Leucaena leucocephala* forage on intake, digestion and live weight gains of *Bos taurus* × *Bos indicus* steers offered Napier grass. *Anim. Sci.* 63: 381-388.

AOAC. 2007. Official Methods of Analysis 18<sup>th</sup> edn. Association of Official. Analytical Chemists, Washington, DC, USA.

Babu, C., Iyanar, K. and Kalamani, A. 2014. High green fodder yielding new grass varieties. *Elect. J. Plant Breed.* 5: 220-229.

Bosch, M.W., Tamminga, S.P.G., Leffering, C.P., and Muylaert, J.M. 1992. Influence of stage of maturity of grass silages on digestion processes in dairy cows. 1. Composition, nylon bag characteristics, digestibility and intake. *Livest. Prod. Sci.* 32: 245-264.

Gallardo, I., Ricardo Barcena., Juan Manuel Pinos-Rodríguez., Mario Cobos., Lorenzo Carreon and Maria Esther Ortega. 2010. Influence of exogenous fibrolytic enzymes on *in vitro* and *in sacco* degradation of

- forages for ruminants. Italian J. Anim. Sci. 9: 8.
- Gutteridge, R.C. and Shelton H.M. 1994. *Forage Tree Legumes in Tropical Agriculture*. Wallingford, G.B. CAB International Oxford. United Kingdom.
- Kabi, F., Bareeba F.B., Havrevoll O. and Mpofu, I.D.T. 2005. Evaluation of protein degradation characteristics and metabolizable protein of elephant grass (*Pennisetum purpureum*) and locally available protein supplements. *Livest. Prod. Sci.* 95: 143-153.
- McDonald. 1981. A revised model for the estimation of protein degradability in the rumen. *J. Agric. Sci.* Camb. 96: 251-252.
- Minson, D.J. 1990. *Forage in Ruminant Nutrition*. London: Academic Press.
- Muia, J.M.K., Tamminga, S., Mbugua, P.N. and Kariuki, J.N. 2001. Effect of supplementing Napier grass (*Pennisetum purpureum*) with poultry litter and sunflower meal based concentrates on feed intake and rumen fermentation in Friesian steers. *Anim. Feed Sci. Technol.* 92: 113-126.
- Orskov, E.R. and McDonald. 1979. The estimation of protein degradability in rumen from the incubation measurements weighted according to rate of passage. *J. Agric. Sci.* Camb. 92:499.
- Osuji, P.O., Nsahlai, I.V. and Khalili, H. 1993. Feed Evaluation. ILCA Manual 5, ILCA, Addis Ababa 10-11.
- Singh, B., Narang M.P. and Makkar, H.P.S. 1992. Rate and extent of *in sacco* degradation of dry matter and cell wall constituents of various grass species. *Indian J. Anim Nutr.* 9: 1-12.

- Snedecor, C.W. and Cochran, W.G. 1994. *Statistical Methods* (6<sup>th</sup> edition). Iowa State University Press, Anes, USA.
- SPSS. 17.0.2007. *Statistical Package for the Social Sciences* (SPSS).
- Tessema, Z. and Baars, R.M.T. 2004. Chemical composition, *in vitro* dry matter digestibility and ruminal degradation of Napier grass (*Pennisetum purpureum* (*L.*) *Schumach.*) mixed with different levels of Sesbania sesban (L.) Merr. *Anim. Feed Sci. Technol.* 117: 29-41.
- Topps, J.H. 1996. Assessment of forage legumes as protein-rich supplement in ruminant production systems in Zimbabwe. In: *Ndikumana J and de Leeuw* (eds). Proceedings, 2<sup>nd</sup> African Feed Resources Network, p. 201. Harare.
- Van Soest, P.J., Robertson, J.B. and Lewis, B.A. 1991. Methods for dietary fibre, neutral detergent fibre and non-starch polysaccharides in relation to animal nutrition. *J. Dairy Sci.* 74:3583-3597.
- Weiss, P.W. 1994. Estimation of digestibility of forages by laboratory methods. In: Fahey GC, Collins M, Mertens DR and Moser E (eds). *Forage Quality, Evaluation and Utilization*, pp. 644-682. Madison: American Societies of Agronomy, Crop Science and Soil Science.
- Woodard, K.R. and Prine, G.M. 1991. Forage yield and quality of elephant grass as affected by harvesting frequency and genotype. *Agron. J.* 83:541-546.

Received on 13-09-2019 and accepted on 30-12-2019

## INDIAN JOURNAL OF ANIMAL NUTRITION

/			1	1 / - 1 1	/:11 /TT A NT)
(www.indianjournals.	com: www.ansi.oi	rg.in: http://e	pubs.icar.org	.in/eiournal	/index.php/IJAN

Marc	h, 2019 Vol. 36	#1
	CONTENTS	
	Review	
1.	Metagenomics and CAZymes in Rumen: A review Anju Kala, Devki Nandan Kamra, Lal Chandra Chaudhary and Neeta Agarwal	1
	Ruminants	
2.	Increasing Dairy Cows Productivity through New Balanced Concentrate Feed:  A Study in Bihar, India  Dhiraj K. Singh, Shree Prasad Sahu and Nils Teufel	11
3.	Effect of UMMB Supplementation on Metabolic and Oxidative Parameters in Rambouillet Cross Sheep during Peripartum Period A.P. Singh, R. Singh, A.M. Bhat, A. Tikoo and A.K. Pathak	17
4.	Evaluation of <i>kharif</i> Forage Crops for Biomass Production and Nutritional Parameters in Indo-Gangetic Plains of India  Phool Singh Hindoriya, Rajesh Kumar Meena, Magan Singh, Rakesh Kumar, Hardev Ram, Vijendra Kumar Meena and Manish Kushwaha	25
5.	Bioavailability of Major Minerals from Commonly Used Ruminants Feeds Using Dialyzability Technique  Abhilasha Singh, Veena Mani, Bhawana Shukla, Anjila Kujur- and Suman Kapila	30
6.	Effect of Poly-Herbal Mixture Supplementation during Post partum Period on Feed Intake and Reproductive Performance of Sahiwal Cows  Piyali Kuri, Parveen Kumar, Nishant Kumar, Anjali Aggarwal and Mahendra Singh	35
	Non Ruminants	
7.	Effect of Supplementation of Synthetic Lysine and Methionine on Serum Biochemical Profile, Carcass Characteristics and Meat Composition in Broiler Chicken Bornalee Handique, G. Saikia, Runjun Dowarah, B.N. Saikia and S. Tamuly	40
8.	Influence of Incorporation of Azolla Meal on Performance of Laying Japanese Quails R. Kasi Sowjanya Lakshmi, Ch. Venkata Seshiah, P. Ravikanth Reddy, K. Nagaraja and I. Kumar	47
9.	Growth Performance and Nutrient Utilization of Male Broiler Chicken as Affected by Feed Restriction with or without Garlic Supplementation  Vishavdeep Singh, Udeybir Singh, A.P.S. Sethi and J.S. Lamba	51
10.	Performance of Broiler Chicken Fed Diets Supplemented with a Phytogenic Mixture V.V. Singh, V.K. Singh, D. Tewari, S. Gautam, V.B. Singh and P. Singh	58
11.	Seasonal Variations in Proximate Composition of Nine Freshwater Fish B.N. Paul, S. Bhowmick, P. Singh, S. Chanda, N. Sridhar and S.S. Giri	65

12.	Characteristics in Japanese Quails Pramod Namdeo, S.P. Tiwari, Raina Doneria, Meenu Dubey and Rajkumar Yadav	73
13.	Development of Calcium Fortified Biscuits Incorporated with Chicken Slaughter House Byproducts and Evaluation of Their Palatability in Dogs  Khushpreet Singh Virk, Om Prakash Malav, Manish Kumar Chatli, Nitin Mehta, Pavan Kumar and Rajesh V. Wagh	77
	<b>Short Communication</b>	
14.	Supplementary Effect of Different Levels of Nano Zinc Oxide on Zinc Bioavailability and Blood Metabolites in Lambs  K.K. Singh, S.B. Maity and A. Maity	83
15.	Growth Performance and Nutrient Utilization in Different Goat Breeds in the Plain Agro-cllimatic Zone of Chhattisgarh Raj Kumar Yadav, Meenu Dubey, Raina Doneria and S.P. Tiwari	88
16.	Evaluation of Berseem ( <i>Trifolium alexandrinum</i> L.) Varieties for Fodder and Seed Production  J.J. Gupta and A. Dey	91
17.	Influence of Supplementing Giloe and Cinnamon on Production Performance in Commercial Broiler Chicken Garima Tiwari, Anshu Rahal, Ashoka Kumar and Anil Kumar	94
18.	Effect of Rumen Modifier on Methanogenesis and Feed Digestibility under <i>in Vitro</i> Conditions Neeti Lakhani, Devki Nandan Kamra, Preeti Lakhani and Anju Kala	99
19.	Supplementation of Brown Seaweed ( <i>Turbinaria conoids</i> ) Powder and its Effect on Blood Metabolites and Mineral Profile in Adult Goats  Rojita Yengkhom, Putan Singh, Nirmala Muwel, Kanti Raje, Bornalee Handique and K. Venkateswaran	103

## INDIAN JOURNAL OF ANIMAL NUTRITION

	(www.indianjournals.com; www.ansi.org.in; http://epubs.icar.org.in/ejournal/index.php/IJAN)	
June	, 2019 Vol. 36	#2
	CONTENTS	
	Review	
1.	Effect of Dietary Plant Secondary Metabolites on Rumen Fermentation and Microbial Community: A Review  P.S. Banakar, Srobana Sarkar, Bhawna Tyagi, V.V. Vinay, Timmi Chugh, Sachin Kumar, Nitin Tyagi and A.K. Tyagi	107
	Ruminants	
2.	Growth Performance, Feed Efficiency and Ingestive Behavior of Sahiwal Calves Divergently Selected for Residual Feed Intake Anil Kumar Singh, Muneendra Kumar, Vinod Kumar, Debashis Roy, Raju Kushwaha, Shalini Vaswani and Avinash Kumar	113
3.	Effect of Molasses Based Multi-nutrient Supplement Containing Chromium on Nutrient Utilization, Milk Yield, Microbial Protein Flow and Antioxidant Status of Lactating Murrah Buffaloes  A.K. Patil, A.K. Verma, Putan Singh and Asit Das	122
4.	Effect of Feeding Yeast Culture on Productive and Reproductive Performance of Dairy Cows  Ajaz A. Ganie, Rameez Ali, Khurshid A Dar, Henna Hamadani, Zahid Bashir, T.K. Sarkar and J.D. Parrah	130
5.	Effect of Dietary Supplementation of Trace Minerals on Semen Production Performance of Sahiwal Bulls During Winter Season  Manjula Thakur, Asgar Ud Deen, Veena Mani, Mukesh Bhakat, Tushar Kumar Mohanty and Goutam Mondal	136
6.	Effect of Pudina ( <i>Mentha piperita</i> ) Supplementation on Nutrient Utilization and Blood Biochemical Parameters of Sheep  A. Ishfaq, A.R. Bhat, A.M. Ganai, Y.A. Beigh and G.G. Sheikh	146
7.	Evaluation of Banana Plant Waste and its Silage-based Diet on Performance of Osmanabadi Goats S.N Anjaneya, B. Ramachandra, T. Thirumalesh and P.K. Anup Kumar	153
8.	An Economic Analysis of Milk Production of Buffalo and Cow in Rajasthan G.L. Meena, Latika Sharma, Siddhartha Mishra and Suraj Choudhary	158
9.	Variation in Straw Grain Ratio and Straw Quality in Different Rice Cultivars of India M.M. Das, A.K. Misra, K.K. Singh, G.H. Pailan and T.A. Khan	164
10.	Effect of Organic and Inorganic Nutrient Sources on Yield and Quality of Fodder Cowpea [Vigna unguiculata (L.) Walp.]  Susanta Dutta, Magan Singh, Rajesh Kumar Meena, Nirmalendu Basak, Goutam Mondal and Phool Singh Hindoriya	173

11.	Productivity and Quanty of Fodder Oats (Avena sativa L.) as influenced by Sowing	
	Time, Cutting Schedules and Nitrogen Levels	179
	S.S. Kadam, N.S. Solanki, Mohd. Arif, L.N. Dashora, S.L. Mundra and B. Upadhyay	
	Non-Ruminants	
12.	Challenzyme Supplementation of High Expeller Copra Meal in Corn-animal Protein Diets for Broilers: Growth Performance, Nutrient Digestibility and Carcass Traits Ashika Devi, Siaka Seriba Diarra and Sandy Hoffman Mael	187
13.	Effect of Dietary Incorporation of <i>Coriander Seed Meal</i> on Production Performance of Japanese quail  N. Bala Chenna Reddy, D. Srinivas Kumar, K. Raja Kishore and K. Naga Raja Kumari	198
	<b>Short Communications</b>	
14.	Effect of Betaine Supplementation on Serum Metabolite Profile in Gestating Sows Alok Mishra, A.K. Verma, Asit Das, Putan Singh and N.R. Sahoo	202
15.	Replacement of Maize by other Cereals on Performance of <i>Pratapdhan</i> Chicks C.M. Yadav and H.L. Bugalia	207
16.	Effect of Feeding Turmeric and Amla on Dressing Percentage and Meat Bone Ratio of Broilers  Gyan Chandra, S.H. Mane, S.A. Dhage and A.V. Rathor	210
17.	Supplementation of Microbial Feed on Performance of Growing Goats C.M. Yadav and N.R. Meena	215

#### INDIAN JOURNAL OF ANIMAL NUTRITION

	(www.indianjournals.com; www.ansi.org.in; http://epubs.icar.org.in/ejournal/index.php/IJAN)	
Septe	mber, 2019 Vol. 36	#3
	CONTENTS	
	Review	
1.	Role of Carotenoids in Ornamental Fish Nutrition: A Review	218
	G.H. Pailan, Sujata Sahoo and D.K. Singh	
	Ruminants	
2.	Effect of Feeding Moringa (Moringa oleifera) as Green Fodder on Feed Intake,	•••
	Milk Yield, Microbial Protein Synthesis and Blood Profile in Crossbred Cows	228
	S.S. Shankhpal, C.R. Waghela, P.L. Sherasia, V. Sridhar, A.K. Srivastava and Digvijay Singh	
3.	High Level of Feeding in Addition to Dam Milk does not Improve Pre-weaning Growth	
	Performance and Kleiber Ratio of Boer × Central Highland Kids	235
	Zeleke Tesema, Mekonnen Tilahun, Asres Zegeye, Mesfin Lakew, Liuel Yizengaw,	
	Solomon Tiruneh, Getachew Worku and Shanbel Kiros	
4.	Effect of Dietary Supplementation of Rumen Protected Calcium Salts of Rapeseed	
	Oil and Encapsulated Rapeseed Oil to Lactating Dairy Cows	242
_	Subrahmanyeswar G, Senthil Murugan S, Biju Chacko and Shyama K	
5.	Effect of Dietary Supplementation of Area Specific Mineral Mixture along with	
	Hormonal Interventions on the Performances of the Anoestrous and Repeat Breeding Crossbred Cattle	247
	Kamdev Sethy, A.K. Behera, R.K. Swain, S.K. Mishra, K. Behera and P.C. Mishra	247
6.	In Vitro Metabolizable Protein and Utilizable Amino Acid Estimation of Ruminant	
•	Feed Ingredients	252
	Sonali Prusty, Vijay Kumar Sharma, U.B. Sontakke and S.S. Kundu	
7.	Effect of Fresh Banana Plant Waste and its Silage on Dry Matter Intake, Nutrient	
	Digestibility and Rumen Parameters in Osmanabadi Kids	260
	Shivaram N. Patil, T. Thirumalesh, B. Ramachandra, M.M. Appannavar, Siddhalinga	
	Swamy Hiremath and M.D. Suranagi	
8.	Body Heat Storage and Physiological Responses of Periparturient Karan Fries and	
	Sahiwal Cows During Summer and Winter Season	266
	M.M. Vaidya and S.V. Singh	
9.	Rumen Protected Choline along with Green Tea Extract Maintain Glucose	
	Homeostasis in Transition Karan Fries Cows	276
	Parag Acharya, S.S. Lathwal, Neela Madhav Patnaik and Baisakhi Moharana	
10	Non-Ruminants  Influence of Dietory Monney eligespeedberide and & Teachbard on Intestinal	
10.	Influence of Dietary Mannan-oligosaccharide and α-Tocopherol on Intestinal Microbiology and Histomorphology in Broiler Chickens	281
	Pallavi Sinha, Sanjay Kumar, Kaushalendra Kumar, Chandramoni, Manju Sinha,	201
	Deepak Kumar, Rajni Kumari and Sushma Kumari	

11.	Effect of Varying Nutrient Density of Diets on Productive Performance in Dahlem	
	Red Layers	286
	B. Prakash, S.V. Rama Rao, M.V.L.N. Raju, S.K. Verma and A.K. Panda	
12.	Optimization of Dietary Protein Requirement for the Growth, Survival and Feed Utilization of Osteobrama belangeri (Valenciennes, 1844) Fingerling Nahakpam Surjobala, Sagar C. Mandal, Arun B. Patel, Prasenjit Pal and Pramod K. Pandey	290
13.	Effect of Dietary Inclusion of Graded Levels of Toasted Guar Meal (TGM) at	
	Different Energy Efficiency on Egg Quality and Serum Parameters of White Leghorn Layers	299
	M. Hanumanth Rao, S.V. Rama Rao and Srinivas Gurram	
14.	Evaluation of Performance of Rajasri Birds under Different Management Situations V. Chinni Preetam, Srinivas Gurram and S. Qudratullah	305
<b>15.</b>	Enzyme Supplementation of Commercial Feed Diluted with Copra Meal for Laying	
	Hens	309
	Sandy Hoffman Mael, Siaka Seriba Diarra, and Ashika Devi	
16.	Optimization of Calcium and Phosphorus Levels in the Diet of Cirrhinus mrigala (Hamilton, 1822) Fingerlings  Anita Bhatnagar, Neelam Rajaharia and Oshin Dhillon	315
	Short Communication	
<b>17.</b>	Effect of Supplementation of Acidifiers with Probiotic on Performance of Broiler	
	Chicken	325
	G.M. Jadhao, S.M. Wankhede, D.H. Rekhate, A.P. Dhok, P.S. Bankar and	
	H.N. Rewatkar	

### INDIAN JOURNAL OF ANIMAL NUTRITION

			•			1 .	. , .	. 1/ 1		/TT A B T
1	WWW indian	murnale com	; www.ansi.org.	in htt	n·//er	nibe icar	org in/ei	iniirnal/ind	ev nh	n/IIAN
١	w w w.maan	ournais.com	, www.ansn.org.	1111, 1111	$\nu . \prime \prime c_{\rm i}$	Juos.icai.	OIZ.III/C	journai/mu	$\omega_{\Lambda}$ , $\rho_{\Pi}$	D/13/11

Decem	mber, 2019 Vol. 36	#4
	CONTENTS	
	Ruminants	
1.	Effect of Feeding Straw Based Densified Complete Feed Blocks Containing Dhanwantharam Oil Residue and Tapioca Starch Waste on Body Weight, Dry Matter Intake, Feed Conversion Efficiency and Nutrient Digestibility in Crossbred Heifer Calves  H.S. Sunil Gowda, Sajith Purushothaman, K. Ally, Deepa Ananth and Shibu Simon	329
2.	Nutrient Intake, Digestibility, Rumen Parameters and Blood Metabolites of Kacang Goats Fed Silage of Forage Mixture Produced from Intercropping of Sorghum Differing in Planting Space with Butterfly Pea (Clitoria ternatea)  E. Hartati, G.A.Y. Lestari, M.M. Kleden and I.G.N. Jelantik	334
3.	Effect of Supplementing Syzygium cumini (Jamun) Fruit Shreds to Total Mixed Ration on the Performance of Lactating Crossbred Cows  A.H. Mayuresh, M. Wadhwa, J.S. Hundal, M.P.S. Bakshi, Amit Sharma, Simerjeet Kaur and B.K. Bansal	342
4.	Comparative Evaluation of Oat Hay and Silage Based Rations on Nutrient Utilization and Methane Emissions in Murrah Buffaloes  U.B. Sontakke, Sonali Prusty, S.S. Kundu and Vijay Kumar Sharma	847
5.	Effect of Depotash Vinasse on Rumen Fermentation Kinetics <i>in vitro</i> Gaurav Pratap Singh, Vandana Kumari Leitanthem, Amit N. Sharma, Madhu Mohini, Naresh Arora and Goutam Mondal	353
6.	The Effects of Dietary ω-3 and ω-6 Fatty Acids on Nutrient Utilization and Growth Performance in Sahiwal Heifers Sushil Kumar, Sajjan Sihag, Zile Singh Sihag, C.S. Patil, Surender Singh Dhaka and Anand Kumar Pandey	358
	Non-Ruminants	
7.	A Newly Developed Mixture of Herbal Plants and Spices Enriched with Special Extracts and Essential Oils Enhances Feed Utilisation, Growth Performance and Lowers Harmful Caecal Bacteria in Rabbits  O.A.H. El-Ghalid, A.M. Abd El-Hady, G.M. El-Ashry, A.E. Kholif, O.H Matloup, O.A.Olafadehan and A.M. El-Raffa	865
8.	A Nursery Feed for Indian Major Carps ( <i>Labeo rohita</i> ) and its Evaluation in Farmer's  Field  Krushna Chandra Das, Kedar Nath Mohanta, Santosh Kumar Nayak, Snehalata Mohanty, Prabin Kumar Sahoo and Priyabrat Swain	377

<b>9.</b>	With or Without Supplementation of Multi-enzymes on Blood Bio-Chemical Constituents of Indigenous Chicken Ashim Kumar Saikia, Robin Bhuyan, Bibeka Nanda Saikia, Digendra Nath Sarma, Ranajit Roychaudhury, Arundhati Bora and Joga Dev Mahanta	382
10.	Effect of Inclusion of Black Pepper Powder as Natural Feed Additive on the Performance of Japanese quail V. Sri Divya, D. Srinivas Kumar and E. Raghava Rao	388
11.	Chemical Evaluation and Nutrient Digestibility of Shrimp Waste Meal in Broilers N. Mounica, J.V. Ramana, D. Srinivasa Rao, J. Suresh and P. Kavitha	393
12.	Growth Performance, Carcass Traits and Economics of Kadaknath birds Rearing under Intensive Condition in Hot and Humid Climate P.K. Jena, B. Panigrahi, N. Panda, L.M. Mohapatra, B.K. Mallik, J. Bagh, P.K. Pati and A. Baliarsingh	399
13.	Effect of Dietary Supplementation of Carrot Meal on Survival, Growth and Pigmentation of Freshwater Ornamental Fish, Koi Carp, Cyprinus Carpio (L.) Abhinka Jain, Vaneet Inder Kaur and Shathanagouda Admane Hollyappa	405
14.	Effect of Dietary Incorporation of Short Chain and Medium Chain Fatty Acid on Feed Intake and Serum metabolites in Broiler Chickens Banani Gantayat, S.M. Durge, S.A. Amrutkar and V.B. Dongre	414
	<b>Short Communication</b>	
15.	Successful Management of Nitrite Poisoning in Crossbred Dairy Calves E. Niyas, Shibu Simon, Surej Joseph Banglavan, Jith John Mathew, Ani S. Das, Reni John and S. Reshma	419
16.	In Sacco Degradability Kinetics of Cumbu Napier (COBN-5) Fodder at Two Different Stages of Harvest M. Madesh, K. Raja Kishore, D. Srinivas Kumar and A. Anitha	423

## **Author Index**

A. Anitha	423	Anshu Rahal	94
A. Baliarsingh	399	Arun B. Patel	290
A. Dey	91	Arundhati Bora	382
A. Ishfaq	146	Asgar Ud Deen	136
A. Maity	83	Ashika Devi	187, 309
A. Tikoo	17	Ashim Kumar Saikia	382
A.E. Kholif	365	Ashoka Kumar	94
A.H. Mayuresh	342	Asit Das	122, 202
A.K. Behera	247	Asres Zegeye	235
A.K. Misra	164	Avinash Kumar	113
A.K. Panda	286	B. Panigrahi	399
A.K. Pathak	17	B. Prakash	286
A.K. Patil	122	B. Ramachandra	153, 260
A.K. Srivastava	228	B. Upadhyay	179
A.K. Tyagi	107	B.K. Bansal	342
A.K. Verma	122, 202	B.K. Mallik	399
A.M. Abd El-Hady	365	B.N. Paul	65
A.M. Bhat	17	B.N. Saikia	40
A.M. El-Raffa	365	Baisakhi Moharana	276
A.M. Ganai	146	Banani Gantayat	414
A.P. Dhok	325	Bhawana Shukla	30
A.P. Singh	17	Bhawna Tyagi	107
A.P.S. Sethi	51	Bibeka Nanda Saikia	382
A.R. Bhat	146	Biju Chacko	242
A.V. Rathor	210	Bornalee Handique	40, 103
Abhilasha Singh	30	C.M. Yadav	207, 215
Abhinka Jain	405	C.R. Waghela	228
Ajaz A. Ganie	130	C.S. Patil	358
Alok Mishra	202	Ch. Venkata Seshiah	47
Amit N. Sharma	353	Chandramoni	281
Amit Sharma	342	D. Srinivas Kumar	198, 388, 423
Anand Kumar Pandey	358	D. Srinivasa Rao	393
Ani S. Das	419	D. Tewari	58
Anil Kumar Singh	113.	D.H. Rekhate	325
Anil Kumar	94	D.K. Singh	218
Anita Bhatnagar	315	Debashis Roy	113
Anjali Aggarwal	35	Deepa Ananth	329
Anjila Kujur	30	Deepak Kumar	281
Aniu Kala	1, 99	Devki Nandan Kamra	1. 99

Digendra Nath Sarma         382         Kanti Raje         103           Digvijay Singh         228         Kaushalendra Kumar         281           E. Hartati         334         Kedar Nath Mohanta         377           E. Niyas         419         Khurshid A Dar         130           E. Raghava Rao         388         Khushpreet Singh Virk         77           G. Sakika         40         Krushna Chandra Das         377           G.A.Y. Lestari         334         L.M. Mohapatra         399           G.G. Sheikh         146         L.N. Dashora         179           G.H. Pailan         164, 218         Lat Chandra Chaudhary         1           G.L. Meena         158         Latika Sharma         158           G.M. Jadhao         325         M. Hanumanth Rao         295           G.M. Jadhao         325         M. Hanumanth Rao         295           Garima Tiwari         94         M. Madesh         423           Gutara Pratap Singh         353         M.D. Suranagi         260           Gutara Worku         235         M.D. Suranagi         260           Gutara Worku         235         M.D. Suranagi         260           Gyan Chandra         210	Dhiraj K. Singh	11	Kamdev Sethy	247
E. Hartati         334         Kedar Nath Mohanta         377           E. Niyas         419         Khurshid A Dar         130           E. Raghava Rao         388         Khushpreet Singh Virk         77           G. Saikia         40         Krushna Chandra Das         377           G.A.Y. Lestari         334         L.M. Mohapatra         399           G.G. Sheikh         146         L.N. Dashora         179           G.H. Pailan         164, 218         Lal Chandra Chaudhary         1           G.M. El-Ashry         365         Liuel Yizengaw         235           G.M. El-Ashry         365         Liuel Yizengaw         235           G.M. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.D. Suranagi         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.V.L.N. Raju         266           H.S. Sunil Gowda         3	Digendra Nath Sarma	382	Kanti Raje	103
E. Niyas         419         Khurshid A Dar         130           E. Raghava Rao         388         Khushpreet Singh Virk         77           G. Saikia         40         Krushna Chandra Das         377           G.A.Y. Lestari         334         L.M. Mohapatra         399           G.G. Sheikh         146         L.N. Dashora         179           G.H. Pailan         164, 218         Lal Chandra Chaudhary         1           G.L. Meena         158         Latika Sharma         158           G.M. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M.Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Goutam Mondal         136, 173, 353         M.M. Vaidwa         260           Gyan Chandra         210         M.M. Eden         334           H.J. Bugalia         207         M.M. Kleden         34           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Henna Hamadani         1	Digvijay Singh	228	Kaushalendra Kumar	281
E. Raghava Rao         388         Khushpreet Singh Virk         77           G. Saikia         40         Krushna Chandra Das         377           G.A.Y. Lestari         334         L.M. Mohapatra         399           G.G. Sheikh         146         L.N. Dashora         179           G.H. Pailan         164, 218         Lal Chandra Chaudhary         1           G.H. Gena         158         Latika Sharma         158           G.M. El-Ashry         365         Liuel Yizengaw         235           G.M. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M. D. Suranagi         260           Goutam Mondal         136, 173, 353         M.D. Suranagi         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           H.S. Sunil Gowda         329         M.P.S. Bakshi         35           I. Kumar         47         Magan Singh         25,173           I.G.N. Jelantik         334<	E. Hartati	334	Kedar Nath Mohanta	377
G. Saikia         40         Krushna Chandra Das         377           G.A.Y. Lestari         334         L.M. Mohapatra         399           G.G. Sheikh         146         L.N. Dashora         179           G.H. Pailan         164, 218         Lal Chandra Chaudhary         1           G.H. Pailan         164, 218         Lal Chandra Chaudhary         1           G.H. Sairy         365         Luiel Yizengaw         235           G.M. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M. Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130	E. Niyas	419	Khurshid A Dar	130
GA.Y. Lestari         334         L.M. Mohapatra         399           G.G. Sheikh         146         L.N. Dashora         179           G.H. Pailan         164, 218         Lal Chandra Chaudhary         1           G.L. Meena         158         Latika Sharma         158           G.M. Jadhao         365         Liuel Yizengaw         235           G.M. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M. Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25           I.S. Jatiki         334         Mahendra S	E. Raghava Rao	388	Khushpreet Singh Virk	77
G.G. Sheikh         146         L.N. Dashora         179           G.H. Pailan         164, 218         Lal Chandra Chaudhary         1           G.H. Meena         158         Latika Sharma         158           G.M. El-Ashry         365         Liuel Yizengaw         235           G.M. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M. Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.W. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Ma	G. Saikia	40	Krushna Chandra Das	377
G.H. Pailan         164, 218         Lal Chandra Chaudhary         1           G.L. Meena         158         Latika Sharma         158           G.M. El-Ashry         365         Liuel Yizengaw         235           G.M. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M. Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L. N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Mahendra Singh         25           J. Suresh         393         Ma	G.A.Y. Lestari	334	L.M. Mohapatra	399
GL. Meena         158         Latika Sharma         158           GM. El-Ashry         365         Liuel Yizengaw         235           GM. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M. Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Mahendra Singh         35           J. Bagh         399         Manish Kumar Chatli         77           J. Suresh         393         Manish Kushwaha<	G.G. Sheikh	146	L.N. Dashora	179
G.M. El-Ashry         365         Liuel Yizengaw         235           G.M. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M. Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Mahendra Singh         35           J. Bagh         399         Manish Kumar Chatli         77           J. Suresh         393         Manish Kushwaha         25           J.D. Parrah         130         Manju Sinha	G.H. Pailan	164, 218	Lal Chandra Chaudhary	1
G.M. Jadhao         325         M. Hanumanth Rao         299           Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M. Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Mahendra Singh         35           J. Bagh         399         Manish Kumar Chatli         77           J. Suresh         393         Manish Kushwaha         25           J.D. Parrah         130         Manjula T	G.L. Meena	158	Latika Sharma	158
Garima Tiwari         94         M. Madesh         423           Gaurav Pratap Singh         353         M. Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Mahendra Singh         35           J. Bagh         399         Manish Kumar Chatli         77           J. Suresh         393         Manish Kushwaha         25           J.D. Parrah         130         Manjula Thakur         136           J.S. Hundal         342         Meenu Dubey	G.M. El-Ashry	365	Liuel Yizengaw	235
Gaurav Pratap Singh         353         M. Wadhwa         342           Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Mahendra Singh         35           J. Bagh         399         Manish Kumar Chatli         77           J. Suresh         393         Manish Kumar Chatli         77           J. Gupta         91         Manju Sinha         281           J.J. Gupta         91         Manju Sinha         281           J.S. Lumba         31         Meenu Dubey         73, 88           J.S. Lumba         51         Mekonnen Tilahun	G.M. Jadhao	325	M. Hanumanth Rao	299
Getachew Worku         235         M.D. Suranagi         260           Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.GN. Jelantik         334         Mahendra Singh         35           J. Bugh         399         Manish Kumar Chatli         77           J. Suresh         393         Manish Kushwaha         25           J.D. Parrah         130         Manju Sinha         281           J.J. Gupta         91         Manjula Thakur         136           J.S. Lamba         51         Mekonnen Tilahun         235           J.V. Ramana         393         Mesfin Lakew         235           Jith John Mathew         419         Mohd. Arif	Garima Tiwari	94	M. Madesh	423
Goutam Mondal         136, 173, 353         M.M. Appannavar         260           Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Mahendra Singh         35           J. Bagh         399         Manish Kumar Chatli         77           J. Suresh         393         Manish Kushwaha         25           J.D. Parrah         130         Manju Sinha         281           J.J. Gupta         91         Manju Sinha         281           J.S. Hundal         342         Meenu Dubey         73, 88           J.S. Lamba         51         Mekonnen Tilahun         235           J.V. Ramana         393         Mesfin Lakew         235           Jith John Mathew         419         Mohd. Arif	Gaurav Pratap Singh	353	M. Wadhwa	342
Gyan Chandra         210         M.M. Das         164           H.L. Bugalia         207         M.M. Kleden         334           H.N. Rewatkar         325         M.M. Vaidya         266           H.S. Sunil Gowda         329         M.P.S. Bakshi         342           Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Mahendra Singh         35           J. Bagh         399         Manish Kumar Chatli         77           J. Suresh         393         Manish Kushwaha         25           J.D. Parrah         130         Manju Sinha         281           J.J. Gupta         91         Manjua Thakur         136           J.S. Hundal         342         Meenu Dubey         73, 88           J.S. Lamba         51         Mekonnen Tilahun         235           J.V. Ramana         393         Mesfin Lakew         235           Jith John Mathew         419         Mohd. Arif         179           Joga Dev Mahanta         382         Mukesh Bhakat	Getachew Worku	235	M.D. Suranagi	260
H.L. Bugalia       207       M.M. Kleden       334         H.N. Rewatkar       325       M.M. Vaidya       266         H.S. Sunil Gowda       329       M.P.S. Bakshi       342         Hardev Ram       25       M.V.L.N. Raju       286         Henna Hamadani       130       Madhu Mohini       353         I. Kumar       47       Magan Singh       25, 173         I.G.N. Jelantik       334       Mahendra Singh       35         J. Bagh       399       Manish Kumar Chatli       77         J. Suresh       393       Manish Kushwaha       25         J.D. Parrah       130       Manju Sinha       281         J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198 </td <td>Goutam Mondal</td> <td>136, 173, 353</td> <td>M.M. Appannavar</td> <td>260</td>	Goutam Mondal	136, 173, 353	M.M. Appannavar	260
H.N. Rewatkar       325       M.M. Vaidya       266         H.S. Sunil Gowda       329       M.P.S. Bakshi       342         Hardev Ram       25       M.V.L.N. Raju       286         Henna Hamadani       130       Madhu Mohini       353         I. Kumar       47       Magan Singh       25, 173         I.G.N. Jelantik       334       Mahendra Singh       35         J. Bagh       399       Manish Kumar Chatli       77         J. Suresh       393       Manish Kushwaha       25         J.D. Parrah       130       Manju Sinha       281         J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Nagaraja       47       N. Panda       399	Gyan Chandra	210	M.M. Das	164
H.S. Sunil Gowda       329       M.P.S. Bakshi       342         Hardev Ram       25       M.V.L.N. Raju       286         Henna Hamadani       130       Madhu Mohini       353         I. Kumar       47       Magan Singh       25, 173         I.G.N. Jelantik       334       Mahendra Singh       35         J. Bagh       399       Manish Kumar Chatli       77         J. Suresh       393       Manish Kushwaha       25         J.D. Parrah       130       Manju Sinha       281         J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Najaraja       47       N. Panda       399 </td <td>H.L. Bugalia</td> <td>207</td> <td>M.M. Kleden</td> <td>334</td>	H.L. Bugalia	207	M.M. Kleden	334
Hardev Ram         25         M.V.L.N. Raju         286           Henna Hamadani         130         Madhu Mohini         353           I. Kumar         47         Magan Singh         25, 173           I.G.N. Jelantik         334         Mahendra Singh         35           J. Bagh         399         Manish Kumar Chatli         77           J. Suresh         393         Manish Kushwaha         25           J.D. Parrah         130         Manju Sinha         281           J.J. Gupta         91         Manjula Thakur         136           J.S. Hundal         342         Meenu Dubey         73, 88           J.S. Lamba         51         Mekonnen Tilahun         235           J.V. Ramana         393         Mesfin Lakew         235           Jith John Mathew         419         Mohd. Arif         179           Joga Dev Mahanta         382         Mukesh Bhakat         136           K. Ally         329         Muneendra Kumar         113           K. Behera         247         N. Bala Chenna Reddy         198           K. Naga Raja Kumari         198         N. Mounica         393           K. Najaraja         47         N. Panda         <	H.N. Rewatkar	325	M.M. Vaidya	266
Henna Hamadani       130       Madhu Mohini       353         I. Kumar       47       Magan Singh       25, 173         I.G.N. Jelantik       334       Mahendra Singh       35         J. Bagh       399       Manish Kumar Chatli       77         J. Suresh       393       Manish Kushwaha       25         J.D. Parrah       130       Manju Sinha       281         J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       2	H.S. Sunil Gowda	329	M.P.S. Bakshi	342
I. Kumar       47       Magan Singh       25, 173         I.G.N. Jelantik       334       Mahendra Singh       35         J. Bagh       399       Manish Kumar Chatli       77         J. Suresh       393       Manish Kushwaha       25         J.D. Parrah       130       Manju Sinha       281         J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73,88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	Hardev Ram	25	M.V.L.N. Raju	286
I.G.N. Jelantik       334       Mahendra Singh       35         J. Bagh       399       Manish Kumar Chatli       77         J. Suresh       393       Manish Kushwaha       25         J.D. Parrah       130       Manju Sinha       281         J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	Henna Hamadani	130	Madhu Mohini	353
J. Bagh       399       Manish Kumar Chatli       77         J. Suresh       393       Manish Kushwaha       25         J.D. Parrah       130       Manju Sinha       281         J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	I. Kumar	47	Magan Singh	25, 173
J. Suresh       393       Manish Kushwaha       25         J.D. Parrah       130       Manju Sinha       281         J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	I.G.N. Jelantik	334	Mahendra Singh	35
J.D. Parrah       130       Manju Sinha       281         J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	J. Bagh	399	Manish Kumar Chatli	77
J.J. Gupta       91       Manjula Thakur       136         J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	J. Suresh	393	Manish Kushwaha	25
J.S. Hundal       342       Meenu Dubey       73, 88         J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	J.D. Parrah	130	Manju Sinha	281
J.S. Lamba       51       Mekonnen Tilahun       235         J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	J.J. Gupta	91	Manjula Thakur	136
J.V. Ramana       393       Mesfin Lakew       235         Jith John Mathew       419       Mohd. Arif       179         Joga Dev Mahanta       382       Mukesh Bhakat       136         K. Ally       329       Muneendra Kumar       113         K. Behera       247       N. Bala Chenna Reddy       198         K. Naga Raja Kumari       198       N. Mounica       393         K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	J.S. Hundal	342	Meenu Dubey	73, 88
Jith John Mathew419Mohd. Arif179Joga Dev Mahanta382Mukesh Bhakat136K. Ally329Muneendra Kumar113K. Behera247N. Bala Chenna Reddy198K. Naga Raja Kumari198N. Mounica393K. Nagaraja47N. Panda399K. Raja Kishore198, 423N. Sridhar65K. Venkateswaran103N.R. Meena215	J.S. Lamba	51	Mekonnen Tilahun	235
Joga Dev Mahanta382Mukesh Bhakat136K. Ally329Muneendra Kumar113K. Behera247N. Bala Chenna Reddy198K. Naga Raja Kumari198N. Mounica393K. Nagaraja47N. Panda399K. Raja Kishore198, 423N. Sridhar65K. Venkateswaran103N.R. Meena215	J.V. Ramana	393	Mesfin Lakew	235
K. Ally329Muneendra Kumar113K. Behera247N. Bala Chenna Reddy198K. Naga Raja Kumari198N. Mounica393K. Nagaraja47N. Panda399K. Raja Kishore198, 423N. Sridhar65K. Venkateswaran103N.R. Meena215	Jith John Mathew	419	Mohd. Arif	179
K. Behera247N. Bala Chenna Reddy198K. Naga Raja Kumari198N. Mounica393K. Nagaraja47N. Panda399K. Raja Kishore198, 423N. Sridhar65K. Venkateswaran103N.R. Meena215	Joga Dev Mahanta	382	Mukesh Bhakat	136
K. Naga Raja Kumari198N. Mounica393K. Nagaraja47N. Panda399K. Raja Kishore198, 423N. Sridhar65K. Venkateswaran103N.R. Meena215	K. Ally	329	Muneendra Kumar	113
K. Nagaraja       47       N. Panda       399         K. Raja Kishore       198, 423       N. Sridhar       65         K. Venkateswaran       103       N.R. Meena       215	K. Behera	247	N. Bala Chenna Reddy	198
K. Raja Kishore198, 423N. Sridhar65K. Venkateswaran103N.R. Meena215	K. Naga Raja Kumari	198	N. Mounica	393
K. Venkateswaran 103 N.R. Meena 215	K. Nagaraja	47	N. Panda	399
	K. Raja Kishore	198, 423	N. Sridhar	65
K.K. Singh 83, 164 N.R. Sahoo 202	K. Venkateswaran	103	N.R. Meena	215
	K.K. Singh	83, 164	N.R. Sahoo	202

N.S. Solanki	179	R.K. Swain	247
Nahakpam Surjobala	290	R. Singh	17
Naresh Arora	353	Raina Doneria	73, 88
Neela Madhav Patnaik	276	Raj Kumar Yadav	88
Neelam Rajaharia	315	Rajesh Kumar Meena	25, 173
Neeta Agarwal	1	Rajesh V. Wagh	77
Neeti Lakhani	99	Rajkumar Yadav	73
Nils Teufel	11	Rajni Kumari	281
Nirmala Muwel	103	Raju Kushwaha	113
Nirmalendu Basak	173	Rakesh Kumar	25
Nishant Kumar	35	Rameez Ali	130
Nitin Mehta	77	Ranajit Roychaudhury	382
Nitin Tyagi	107	Reni John	419
O.A. Olafadehan	365	Robin Bhuyan	382
O.A.H. El-Ghalid	365	Rojita Yengkhom	103
O.H. Matloup	365	Runjun Dowarah	40
Om Prakash Malav	77	S. Bhowmick	65
Oshin Dhillon	315	S. Chanda	65
P. Kavitha	393	S. Gautam	58
P. Ravikanth Reddy	47	S. Qudratullah	305
P. Singh	58, 65	S. Reshma	419
P.C. Mishra	247	S. Tamuly	40
P.K. Anup Kumar	153	S.A. Amrutkar	414
P.K. Jena	399	S.A. Dhage	210
P.K. Pati	399	S.B. Maity	83
P.L. Sherasia	228	S.H. Mane	210
P.S. Banakar	107, 325	S.K. Mishra	247
Pallavi Sinha	281	S.K. Verma	286
Parag Acharya	276	S.L. Mundra	179
Parveen Kumar	35	S.M. Durge	414
Pavan Kumar	77	S.M. Wankhede	325
Phool Singh Hindoriya	25, 173	S.N Anjaneya	153
Piyali Kuri	35	S.P. Tiwari	73, 88
Prabin Kumar Sahoo	377	S.S. Giri	65
Pramod Namdeo	73	S.S. Kadam	179
Pramod K. Pandey	290	S.S. Kundu	252, 347
Prasenjit Pal	290	S.S. Lathwal	276
Preeti Lakhani	99	S.S. Shankhpal	228
Priyabrat Swain	377	S.V. Rama Rao	286, 299
Putan Singh	103, 122, 202	S.V. Singh	266
R. Kasi Sowjanya Lakshmi	47	Sachin Kumar	107

Sagar C. Mandal	290	V. Sridhar	228
Sajith Purushothaman	329	V.B. Dongre	414
Sajjan Sihag	358	V.B. Singh	58
Sandy Hoffman Mael	187, 309	V.K. Singh	58
Sanjay Kumar	281	V.V. Singh	58
Santosh Kumar Nayak	377	V.V. Vinay	107
Senthil Murugan S	242	Vandana Kumari Leitanthem	353
Shalini Vaswani	113	Vaneet Inder Kaur	405
Shanbel Kiros	235	Veena Mani	30, 136
Shathanagouda Admane Hollyappa	405	Vijay Kumar Sharma	252, 347
Shibu Simon	329, 419	Vijendra Kumar Meena	25
Shivaram N. Patil	260	Vinod Kumar	113
Shree Prasad Sahu	11	Vishavdeep Singh	51
Shyama K	242	Y.A. Beigh	146
Siaka Seriba Diarra	187, 309	Zahid Bashir	130
Siddhalinga Swamy Hiremath	260	Zeleke Tesema	235
Siddhartha Mishra	158	Zile Singh Sihag	358
Simerjeet Kaur	342		
Snehalata Mohanty	377		
Solomon Tiruneh	235		
Sonali Prusty	252, 347		
Srinivas Gurram	299, 305		
Srobana Sarkar	107		
Subrahmanyeswar G	242		
Sujata Sahoo	218		
Suman Kapila	30		
Suraj Choudhary	158		
Surej Joseph Banglavan	419		
Surender Singh Dhaka	358		
Susanta Dutta	173		
Sushil Kumar	358		
Sushma Kumari	281		
T. Thirumalesh	153, 260		
T.A. Khan	164		
T.K. Sarkar	130		
Timmi Chugh	107		
Tushar Kumar Mohanty	136		
U.B. Sontakke	252, 347		
Udeybir Singh	51		
V. Chinni Preetam	305		
V. Sri Divya	388		

(xii)

# Subject Index

Acidifier	325	Cattle	247
Agro-climatic regions	164	CAZymes	1
Alpha-tocopherol	281	Cereals	207
Alternative protein sources	187	Chemical composition 25, 164, 173, 3	05, 393
Amino acids	252, 393	Chicken	58
Ammonia excretion	315	Cholesterol	202
Aquarium	218	Cinnamon	94
Astaxanthin	218	Cirrhinus mrigala	315
Azolla meal	47	Clitoria ternatea	334
Bakerwal sheep	146	CNCPS protein fractions	252
Balanced feed	11	COBN-5 fodder	423
Banana plant waste silage	260	Colour enhancement	405
Banana plant waste	153	Colouration	218
Barbari	88	Composition	353
Berseem	91	Copra meal	309
Betaine	202	Coriander seed meal (CSM)	198
Bioavailability	30	Correlation	235
Biofertilizers	173	Cost	210
Biohydrogenation	107	Cow	158
Birth type	235	Crop contents	305
Black pepper	388	Crossbred cows	228
Blood biochemical parameters	40	Cr-Picolinate	122
Blood bio-chemical	382	Cutting and nitrogen	179
Blood metabolic profile	365	Daily weight gain	215
Blood metabolites	83	Dairy cows	130
Blood parameters	103	Degradation kinetics	423
Body weight	65	Densified complete feed blocks	329
Broiler chicken	40, 94	Depotash vinasse	353
Broiler performance	187	Dhanwantharam oil residue	329
Broilers 58, 207, 210,	281, 325, 393, 414	Dialyzability	30
Brown seaweed	103	Diet composition	187
Buffalo	158	Distillers dried grains with soluble (DDGS	S) 382
Bull	136	DL-methionine	40
Calcium	315	Dog biscuits	77
Carcass characteristics	40, 198, 365, 388	Dressing	210
Carcass traits	47, 399	Economics	58, 399
Carotenoids	218, 405	Efficiency	113
Carps	377	Egg production	286
Carrot meal	405	Egg quality parameters	286

Egg quality         47         In sacco degradability         423           Encapsulation         242         In vitro digestibility         353           Energy efficiency         299         In vitro true digestibility         99           Enzyme         309, 382         In vitro true digestibility         99           Exogenous enzyme         187         Indigenous chicken         382           Experimental trials         11         Ingestive behavior         113           Extrusion         377         Intention         276           FCR         377         Intention         399           Feed conversion ratio         207         Intercropping         334           Feed efficiency         242         Intestinal microbiology         281           Feed enstriction         51         IVDMD         30           Feed erstriction         21         Japanes equalis	Egg quality traits	309	IGF-1	276
Encapsulation         242         In vitro digestibility         353           Energy efficiency         299         In vitro digestion         30           Enzyme         309, 382         In vitro true digestibility         99           Exogenous enzyme         187         Indigenous chicken         382           Experimental trials         111         Ingestive behavior         113           Extrusion         377         Intensive rearing         399           Feed extrusion         207         Intercropping         334           Feed efficiency         242         Intestinal microbiology         281           Feed intake         35,414         Investment pattern         138           Feed restriction         51         IVDMD         30           Feed restriction         51         IVDMD         30           Feed restrictions         25         Kacang Goats         334           Fiber         1         Japanese quails         47, 73           Fiber fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder production         91         Kharif crops         25           Forag				
Energy efficiency         299         In vitro digestion         30           Enzyme         309, 382         In vitro true digestibility         99           Exogenous enzyme         187         Indigenous chicken         382           Experimental trials         11         Ingestive behavior         113           Extrusion         377         Insulin         276           FCR         377         Intercropping         334           Feed conversion ratio         207         Intercropping         334           Feed efficiency         242         Intestinal microbiology         281           Feed intake         35, 414         Investment pattern         158           Feed restriction         51         IVDMD         30           Feed restriction         290         Jamunapari         88           Fiber         1         Japanese quails         47, 73           Fiber fractions         25         Kacang Goats         334           Fibre fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder production         91         Kharif crops         25           Forage and nutrient yie				
Enzyme         309, 382         In vitro true digestibility         99           Exogenous enzyme         187         Indigenous chicken         382           Experimental trials         11         Ingestive behavior         113           Extrusion         377         Insulin         276           FCR         377         Intensive rearing         399           Feed conversion ratio         207         Intersive pring         334           Feed efficiency         242         Intestinal microbiology         281           Feed intake         35, 414         Investment pattern         158           Feed erstriction         51         IVDMD         30           Feed erstriction         51         IVDMD         30           Feed ettilization         290         Jamunapari         88           Fiber         1         Japanese quails         47,73           Fiber fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fordge and nutrient yield         25         Kids         266           Fresh banana plant waste	<u>*</u>		•	
Exogenous enzyme         187         Indigenous chicken         382           Experimental trials         11         Ingestive behavior         113           Extrusion         377         Insulin         276           FCR         377         Intensive rearing         399           Feed conversion ratio         207         Intercropping         334           Feed efficiency         242         Intestinal microbiology         281           Feed intake         35, 414         Investment pattern         158           Feed restriction         51         IVDMD         30           Feed restriction         290         Jamunapari         88           Fiber         1         Japanese quails         47, 73           Fibre fractions         25         Kacang Goats         334           Fibre fractions         25         Kacang Goats         334           Fibre fractions         25         Kacang Goats         334           Fibre fractions         25         Kacang Goats         392           Folder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         <			•	99
Experimental trials         11         Ingestive behavior         113           Extrusion         377         Insulin         276           FCR         377         Intensive rearing         399           Feed conversion ratio         207         Intercropping         334           Feed deficiency         242         Intestinal microbiology         281           Feed intake         35, 414         Investment pattern         158           Feed intake         35, 414         Investment pattern         158           Feed intake         35, 414         Investment pattern         158           Feed extriction         51         IVDMD         30           Feed extrictions         290         Jamunapari         88           Fiber         1         Japanese quails         47, 73           Fiber fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste	•		·	
Extrusion         377         Insulin         276           FCR         377         Intensive rearing         399           Feed conversion ratio         207         Intercropping         334           Feed efficiency         242         Intestinal microbiology         281           Feed efficiency         242         Jamunapari         38           Fised efficiency         242         Kacang Goats         334           Forber fractions         25         Kacang Goats			•	
Feed conversion ratio         207         Intercropping         334           Feed efficiency         242         Intestinal microbiology         281           Feed intake         35, 414         Investment pattern         158           Feed restriction         51         IVDMD         30           Feed et utilization         290         Jamunapari         88           Fiber         1         Japanese quails         47,73           Fibre fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe	-	377	•	276
Feed conversion ratio         207         Intercropping         334           Feed efficiency         242         Intestinal microbiology         281           Feed intake         35, 414         Investment pattern         158           Feed restriction         51         IVDMD         30           Feed utilization         290         Jamunapari         88           Fiber         1         Japanese quails         47, 73           Fibre fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         <	FCR	377	Intensive rearing	399
Feed efficiency         242         Intestinal microbiology         281           Feed intake         35, 414         Investment pattern         158           Feed restriction         51         IVDMD         30           Feed utilization         290         Jamunapari         88           Fiber         1         Japanese quails         47,73           Fibre fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Li	Feed conversion ratio	207	· ·	334
Feed intake         35, 414         Investment pattern         158           Feed restriction         51         IVDMD         30           Feed utilization         290         Jamunapari         88           Fiber         1         Japanese quails         47, 73           Fibre fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Garlic supplementation         51         Lamb growth         33           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Guast         88, 103         L-threonine         73           Green tea extract         276         Maintenanc	Feed efficiency	242		281
Feed utilization         290         Jamunapari         88           Fiber         1         Japanese quails         47, 73           Fibre fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Fresh banana plant waste         260         Kleiber ratio         235           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Growth performance         58, 73, 88, 94, 315, 377<	Feed intake	35, 414		158
Fiber         1         Japanese quails         47, 73           Fibre fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growth performance         58, 73, 88, 94, 315, 377	Feed restriction	51	IVDMD	30
Fibre fractions         25         Kacang Goats         334           Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         235	Feed utilization	290	Jamunapari	88
Fish meal         393         Kadaknath chicken         399           Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         207, 290, 358, 39	Fiber	1	Japanese quails	47, 73
Fodder cowpea         173         Karan Fries         266           Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth         207, 290, 358, 399         Mannan- oligosaccharide         281           Haemato-biochemical	Fibre fractions	25	Kacang Goats	334
Fodder production         91         Kharif crops         25           Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         207, 290, 358, 399         Mannan- oligosaccharide         281           Haemato-biochemical         146         Meat bone ratio         210           Haugh unit <td>Fish meal</td> <td>393</td> <td>Kadaknath chicken</td> <td>399</td>	Fish meal	393	Kadaknath chicken	399
Forage and nutrient yield         25         Kids         260           Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         235         Male broiler chicken         51           Growth         207, 290, 358, 399         Mannan- oligosaccharide         281           Haemato-biochemical         146         Meat bone ratio         210           Haugh unit	Fodder cowpea	173	Karan Fries	266
Fresh banana plant waste         260         Kleiber ratio         235           Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         235         Male broiler chicken         51           Growth         207, 290, 358, 399         Mannan- oligosaccharide         281           Haemato-biochemical         146         Meat bone ratio         210           Haugh unit         299         Mentha piperita         146           Haugh	Fodder production	91	Kharif crops	25
Freshwater fish         65         Koi carp         405           Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         235         Male broiler chicken         51           Growth         207, 290, 358, 399         Mannan- oligosaccharide         281           Haemato-biochemical         146         Meat bone ratio         210           Haugh unit         299         Mentha piperita         146           Haugh unit         299         Mentha piperita         202	Forage and nutrient yield	25	Kids	260
Gain         260         Lactating crossbred cows         342           Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         235         Male broiler chicken         51           Growth         207, 290, 358, 399         Mannan- oligosaccharide         281           Haemato-biochemical         146         Meat bone ratio         210           Haugh unit         299         Mentha piperita         146           Haugh unit         299         Mentha piperita         202	Fresh banana plant waste	260	Kleiber ratio	235
Garlic supplementation         51         Lamb growth         83           Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         235         Male broiler chicken         51           Growth         207, 290, 358, 399         Mannan- oligosaccharide         281           Haemato-biochemical         146         Meat bone ratio         210           Haugh unit         299         Mentha piperita         146           Hay         347         Metabolicprofile         202	Freshwater fish	65	Koi carp	405
Gestating sows         202         Laying performance         309           Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         235         Male broiler chicken         51           Growth         207, 290, 358, 399         Mannan- oligosaccharide         281           Haemato-biochemical         146         Meat bone ratio         210           Haugh unit         299         Mentha piperita         146           Hay         347         Metabolicprofile         202	Gain	260	Lactating crossbred cows	342
Giloe         94         Liver         77           Gizzard         77         Livestock productivity         11           Glucose         276         L-lysine         40           Goats         88, 103         L-threonine         73           Green tea extract         276         Maintenance cost         158           Growing goat         215         Maize         207           Growth performance         58, 73, 88, 94, 315, 377         Major minerals         30           Growth rate         235         Male broiler chicken         51           Growth         207, 290, 358, 399         Mannan- oligosaccharide         281           Haemato-biochemical         146         Meat bone ratio         210           Haugh unit         299         Mentha piperita         146           Hay         347         Metabolicprofile         202	Garlic supplementation	51	Lamb growth	83
Gizzard77Livestock productivity11Glucose276L-lysine40Goats88, 103L-threonine73Green tea extract276Maintenance cost158Growing goat215Maize207Growth performance58, 73, 88, 94, 315, 377Major minerals30Growth rate235Male broiler chicken51Growth207, 290, 358, 399Mannan- oligosaccharide281Haemato-biochemical146Meat bone ratio210Haugh unit299Mentha piperita146Hay347Metabolicprofile202	Gestating sows	202	Laying performance	309
Glucose276L-lysine40Goats88, 103L-threonine73Green tea extract276Maintenance cost158Growing goat215Maize207Growth performance58, 73, 88, 94, 315, 377Major minerals30Growth rate235Male broiler chicken51Growth207, 290, 358, 399Mannan- oligosaccharide281Haemato-biochemical146Meat bone ratio210Haugh unit299Mentha piperita146Hay347Metabolicprofile202	Giloe	94	Liver	77
Goats88, 103L-threonine73Green tea extract276Maintenance cost158Growing goat215Maize207Growth performance58, 73, 88, 94, 315, 377Major minerals30Growth rate235Male broiler chicken51Growth207, 290, 358, 399Mannan- oligosaccharide281Haemato-biochemical146Meat bone ratio210Haugh unit299Mentha piperita146Hay347Metabolicprofile202	Gizzard	77	Livestock productivity	11
Green tea extract276Maintenance cost158Growing goat215Maize207Growth performance58, 73, 88, 94, 315, 377Major minerals30Growth rate235Male broiler chicken51Growth207, 290, 358, 399Mannan- oligosaccharide281Haemato-biochemical146Meat bone ratio210Haugh unit299Mentha piperita146Hay347Metabolicprofile202	Glucose	276	L-lysine	40
Growing goat215Maize207Growth performance58, 73, 88, 94, 315, 377Major minerals30Growth rate235Male broiler chicken51Growth207, 290, 358, 399Mannan- oligosaccharide281Haemato-biochemical146Meat bone ratio210Haugh unit299Mentha piperita146Hay347Metabolicprofile202	Goats	88, 103	L-threonine	73
Growth performance58, 73, 88, 94, 315, 377Major minerals30Growth rate235Male broiler chicken51Growth207, 290, 358, 399Mannan- oligosaccharide281Haemato-biochemical146Meat bone ratio210Haugh unit299Mentha piperita146Hay347Metabolicprofile202	Green tea extract	276	Maintenance cost	158
Growth rate 235 Male broiler chicken 51 Growth 207, 290, 358, 399 Mannan- oligosaccharide 281 Haemato-biochemical 146 Meat bone ratio 210 Haugh unit 299 Mentha piperita 146 Hay 347 Metabolicprofile 202	Growing goat	215	Maize	207
Growth207, 290, 358, 399Mannan-oligosaccharide281Haemato-biochemical146Meat bone ratio210Haugh unit299Mentha piperita146Hay347Metabolicprofile202	Growth performance	58, 73, 88, 94, 315, 377	Major minerals	30
Haemato-biochemical146Meat bone ratio210Haugh unit299Mentha piperita146Hay347Metabolicprofile202	Growth rate	235	Male broiler chicken	51
Haugh unit299Mentha piperita146Hay347Metabolic profile202	Growth	207, 290, 358, 399	Mannan- oligosaccharide	281
Hay 347 Metabolicprofile 202	Haemato-biochemical	146	Meat bone ratio	210
·	Haugh unit	299	Mentha piperita	146
77. 14. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1. 1.	Hay	347	Metabolicprofile	202
Heat storage 266 Metabolizable protein 252	Heat storage	266	Metabolizable protein	252
Histomorphology 281 Metagenomics 1	Histomorphology	281	Metagenomics	1
A.M. 3.5.1	Hormones	247	Methane emission	347
	Hybrid napier	228	Methane	99, 107, 342
Hormones 247 Methane emission 347	Hybrid napier	228	Methane	99, 107, 342

Methylene blue	419	Poly-herbal mixture	35
Microbial	215	Probiotic	325
Milk components	130	Production performance	198, 388
Milk composition	11, 242	Productivity	179
Milk production	35, 130	Protein requirement	290
Milk production	35	Proximate composition	65
Milk Yield	122, 228, 235, 242, 342	Seasonal variations	65
Minerals	17, 247	Pubertal age	358
Molasses	122	Purine derivative index	122
Moringa	228	Quail	198, 388
Mortality	210	Quality	179
Multi nutrient supplemen	ntation 122	Rabbits	365
Murrah buffaloes	122, 347	Rajasri	305
Nano zinc oxide	83	Rajasthan	158
Net income	158	Rapeseed oil	242
Nitrate	419	Red gram straw	153
Nitrite	419	Reproduction	35, 247
Nitrogen retention	325	Reproductive performance	130
Nursery feed	377	Residual feed intake	113
Nutrient density	286	Rrumen microbes	1
Nutrient digestibility	51, 187, 329, 388, 393	Rumen fermentation	353
Nutrient sources	173	Rumen microbes	107
Nutrient utilization	88, 122, 146, 342, 358	Rumen modifier	99
Nylon bag degradability	164	Rumen protected choline	276
Oat silage	347	Rumen protected fat	242
Oats	179	Sahiwal calves	113
Organoleptic	228	Sahiwal cows	35
Ornamental fish feed	218	Sahiwal heifers	358
Osteobrama belangeri	290	Sahiwal	136, 266
Oxidative stress	17	Scavenging system	305
Paddy cultivars	164	Seed production	91
Pathogenic bacteria	365	Semen quality	136
Percentage	210	Semi-purified diets	290
Performance	47, 113	Sensory evaluation	77
Peripartum period	17	Serum biochemical parameters	153
Periparturient	266	Serum chemistry	414
Phosphorus	315	Serum minerals	103
Phytogenic additives mix	sture 365	Serum protein	299
Phytogenic mixture	58	Sex	235
Pigmentation	405	Sheep	17
Plant secondary metaboli	ites 107	Shell quality	299

Short chain and medium chain fatty acid	d 414	Temperature humidity index	266
Shrimp waste meal	393	Texture profile	77
Silage	153, 334	Toasted guar meal	299
Sirohi	88	Trace Mineral	136
Sodium diformate	325	Transition Karan Fries cows	276
Sodium sulphate	99	Triglyceride	202
Sodium thiosulphate	419	UMMB	17
Sorghum	334	Utilizable crude protein	252
Sowing	179	Variety	91
Stage of harvest	423	Weight gain	365
Straw grain ratio	164	Yeast	130
Supplement	215	Yields	173
Supplementation	73	Zinc bioavailability	83
Syzygium cumini fruit shreds	342	ω-3 and ω-6 fatty acids	358
Tapioca starch waste	329		

### List of referees during 2019

- 1. Dr. A. Sahoo, Avikanagar, Rajasthan
- 2. Dr. Amrish Tyagi, NDRI, Karnal, Haryana
- Dr. Anand Pathak, Sher-e-Kashmir, University of Agricultural Sciences & Technology, Jammu
- 4. Dr. Anju Kala, IVRI, Izatnagar, Uttar Pradesh
- 5. Dr. Arun Panda, Directorate of Poultry Research
- 6. Dr. Asit Das, NDRI, Karnal, Haryana
- 7. Dr. Avijit Dey, CIRB, Hisar, Haryana
- 8. Dr. Avishek Biswas, CARI, Izatnagar, Bareilly
- 9. Dr. B. Prakash, ICAR-DPR, Hyderabad
- 10. Dr. Chander Datt, NDRI, Karnal, Haryana
- 11. Dr. Daisy Wadhwa, CSKHPKV, Palampur
- Dr. Debashis Roy, DUVASU, Mathura, Uttar Pradesh
- 13. Dr. Debasis De, CIBA, West Bengal,
- 14. Dr. G.H. Pailan, CIFE, Kolkata, West Bengal
- 15. Dr. Goutam Mondal, NDRI, Karnal, Haryana
- 16. Dr. Jaspal Hundal, GADVASU, Ludhiana
- Dr. Keshab Barman, ICAR-NRC on Pig, Guwahati, Assam
- 18. Dr. Krushna Das, ICAR-CIFA, Bhubaneswar,
- 19. Dr. L.C. Chaudhary, IVRI, Izatnagar, Uttar Pradesh
- 20. Dr. M.K. Tripathi, CIRG, Makhdoom, Uttar Pradesh
- 21. Dr. M.S. Mahesh, Kemin Industries, Chennai
- 22. Dr. Madan Das, IGFRI, Jhansi
- 23. Dr. Madhu Datta, NDRI, Karnal, Haryana
- Dr. Mahipal Choubey, Navsari Agricultural University, Navsari
- 25. Dr. Med Ram Verma, IVRI, Izatnagar, Uttar Pradesh
- Dr. Muneendra Kumar, DUVASU, Mathura, Uttar Pradesh

- 27. Dr. N Dutta, ICAR-IVRI, Izatnagar, Uttar Pradesh
- 28. Dr. Niranjan Panda, Odisha University of Agriculture and Technology, Bhubaneswar
- 29. Dr. Nitin Tyagi, NDRI, Karnal, Haryana
- 30. Dr. P. Vasan, Namakkal, TANUVAS, Tamil Nadu
- 31. Dr. P.R. Pandya, Veterinary College, Anand, Gujarat
- 32. Dr. Pankaj Kumar Singh, Patna, Bihar
- 33. Dr. Parimal Sardar, CIFE, Kolkata
- 34. Dr. Prafulla Naik, ICAR-CARI, Bhubaneswar, Odisha
- 35. Dr. R.K. Dhuria, RAJUVAS, Bikaner, Rajasthan
- 36. Dr. Rajiv Chawla, NDDB, Anand, Gujarat
- 37. Dr. Raka Saxena, ICAR-NIAEPR, New Delhi
- 38. Dr. Rakesh Kumar, NDRI, Karnal, Haryana
- 39. Dr. Ram Singh, ICAR-CIRB, Hissar, Haryana
- 40. Dr. Raman Malik, NDRI, Karnal, Haryana
- 41. Dr. Ravindra Kumar, CIRG, Makhdoom, Uttar Pradesh
- 42. Dr. S.E. Jadhav, IVRI, Izatnagar, Uttar Pradesh
- 43. Dr. S.K. Bhanja, CARI, Izatnagar, Uttar Pradesh
- 44. Dr. Sachin Kumar, NDRI, Karnal, Haryana
- 45. Dr. Sanat Mahanta, IGFRI, Jhansi, Uttar Pradesh
- 46. Dr. Shalini Vaswani, DUVASU, Mathura, Uttar Pradesh
- 47. Dr. Sunil Nayak, College of Veterinary Sci and AH, Jabalpur, Madhya Pradesh
- 48. Dr. T.K. Datta, NDRI, ERS, Kalayani, West Bengal
- 49. Dr. T.K. Ghoshal, CIBA, Chennai, Tamil Nadu.
- 50. Dr. Udeybir Chahal, GADVASU, Ludhiana
- 51. Dr. Veena Mani, NDRI, Karnal, Haryana
- 52. Dr. V.B. Chaturvedi, IVRI, Izatnagar, Uttar Pradesh
- 53. Dr. Vinod Kumar Sidhu, DUVAS Mathura, Uttar Pradesh

### INDIAN JOURNAL OF ANIMAL NUTRITION

(www.indianjournals.com; www.ansi.org.in; http://epubs.icar.org.in/ejournal/index.php/IJAN)

Dece	nber, 2019 Vol. 36	#4
	CONTENTS	
	Ruminants	
1.	Effect of Feeding Straw Based Densified Complete Feed Blocks Containing Dhanwantharam Oil Residue and Tapioca Starch Waste on Body Weight, Dry Matter Intake, Feed Conversion Efficiency and Nutrient Digestibility in Crossbred Heifer Calves H.S. Sunil Gowda, Sajith Purushothaman, K. Ally, Deepa Ananth and Shibu Simon	329
2.	Nutrient Intake, Digestibility, Rumen Parameters and Blood Metabolites of Kacang Goats Fed Silage of Forage Mixture Produced from Intercropping of Sorghum Differing in Planting Space with Butterfly Pea (Clitoria ternatea) E. Hartati, G.A.Y. Lestari, M.M. Kleden and I.G.N. Jelantik	334
3.	Effect of Supplementing Syzygium cumini (Jamun) Fruit Shreds to Total Mixed Ration on the Performance of Lactating Crossbred Cows  A.H. Mayuresh, M. Wadhwa, J.S. Hundal, M.P.S. Bakshi, Amit Sharma, Simerjeet Kaur and B.K. Bansal	342
4.	Comparative Evaluation of Oat Hay and Silage Based Rations on Nutrient Utilization and Methane Emissions in Murrah Buffaloes  U.B. Sontakke, Sonali Prusty, S.S. Kundu and Vijay Kumar Sharma	347
5.	Effect of Depotash Vinasse on Rumen Fermentation Kinetics in vitro Gaurav Pratap Singh, Vandana Kumari Leitanthem, Amit N. Sharma, Madhu Mohini, Naresh Arora and Goutam Mondal	353
6.	The Effects of Dietary &-3 and &-6 Fatty Acids on Nutrient Utilization and Growth Performance in Sahiwal Heifers Sushil Kumar, Sajjan Sihag, Zile Singh Sihag, C.S. Patil, Surender Singh Dhaka and Anand Kumar Pandey	358
	Non-Ruminants	
7.	A Newly Developed Mixture of Herbal Plants and Spices Enriched with Special Extracts and Essential Oils Enhances Feed Utilisation, Growth Performance and Lowers Harmful Caecal Bacteria in Rabbits  O.A.H. El-Ghalid, A.M. Abd El-Hady, G.M. El-Ashry, A.E. Kholif, O.H Matloup, O.A.Olafadehan and A.M. El-Raffa	365
8.	A Nursery Feed for Indian Major Carps ( <i>Labeo rohita</i> ) and its Evaluation in Farmer's Field  Krushna Chandra Das, Kedar Nath Mohanta, Santosh Kumar Nayak, Snehalata Mohanty, Prabin Kumar Sahoo and Priyabrat Swain	377

9.	Effects of Feeding Graded Levels of Distillers Dried Grains with Soluble (DDGS) With or Without Supplementation of Multi-enzymes on Blood Bio-Chemical Constituents of Indigenous Chicken Ashim Kumar Saikia, Robin Bhuyan, Bibeka Nanda Saikia, Digendra Nath Sarma, Ranajit Roychaudhury, Arundhati Bora and Joga Dev Mahanta	382
10.	Effect of Inclusion of Black Pepper Powder as Natural Feed Additive on the Performance of Japanese quail V. Sri Divya, D. Srinivas Kumar and E. Raghava Rao	388
11.	Chemical Evaluation and Nutrient Digestibility of Shrimp Waste Meal in Broilers N. Mounica, J.V. Ramana, D. Srinivasa Rao, J. Suresh and P. Kavitha	393
12.	Growth Performance, Carcass Traits and Economics of Kadaknath birds Rearing under Intensive Condition in Hot and Humid Climate P.K. Jena, B. Panigrahi, N. Panda, L.M. Mohapatra, B.K. Mallik, J. Bagh, P.K. Pati and A. Baliarsingh	399
13.	Effect of Dietary Supplementation of Carrot Meal on Survival, Growth and Pigmentation of Freshwater Ornamental Fish, Koi Carp, Cyprinus Carpio (L.)  Abhinka Jain, Vaneet Inder Kaurand Shathanagouda Admane Hollyappa	405
14.	Effect of Dietary Incorporation of Short Chain and Medium Chain Fatty Acid on Feed Intake and Serum metabolites in Broiler Chickens  Banani Gantayat, S.M. Durge, S.A. Amrutkar and V.B. Dongre	414
	<b>Short Communication</b>	
15.	Successful Management of Nitrite Poisoning in Crossbred Dairy Calves  E. Niyas, Shibu Simon, Surej Joseph Banglavan, Jith John Mathew, Ani S. Das, Reni John and S. Reshma	419
16.	In Sacco Degradability Kinetics of Cumbu Napier (COBN-5) Fodder at Two Different Stages of Harvest  M. Madesh, K. Raja Kishore, D. Srinivas Kumar and A. Anitha	423